

**HUMAN T LYMPHOCYTE SUBPOPULATIONS DEFINED
BY Fc RECEPTORS AND MONOCLONAL ANTIBODIES**

A Comparison*

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Several approaches have provided evidence for the existence of distinct T lymphocyte subsets in man as defined by cell-surface markers. To a large extent, these studies have employed either differential Fc-receptor binding, or antisera reactive selectively with subpopulations of T cells. With the former method, it was shown that two human T cell subsets, T cells bearing receptors for the Fc portion of IgG (T γ) and T cells bearing receptors for the Fc portion of IgM (T μ), could be isolated via surface receptors capable of binding the Fc portion of IgG or IgM, respectively (1, 2). Approximately 60–70% of T lymphocytes in peripheral blood were defined as T μ , whereas a smaller portion representing <20% were T γ cells. Functionally, T γ and T μ populations were distinct with respect to phytohemagglutinin responsiveness and capacity to effect help or suppression of B cell-immunoglobulin production in a pokeweed-mitogen-driven system (3, 4).

Hetero- and autoimmune antisera and hybridomas that secrete monoclonal antibodies directed at subsets of human T cells have provided an alternative method for their identification (5–16). 20% of peripheral T cells were found to be reactive with TH₂ heteroantisera (TH₂⁺) or a monoclonal antibody termed OKT5 (OKT5⁺) and both defined the cytotoxic/suppressor populations (6, 7, 17). The remaining 80% of T cells were unreactive with both TH₂ and OKT5 antisera and defined as TH₂⁻ (OKT5⁻). A second monoclonal antibody, OKT4, reacted with 60% of the total T cell population and was restricted in its reactivity to the TH₂⁻ (OKT5⁻) T cell subpopulation (12–16). Only the OKT4⁺ T cell population responded with proliferation to soluble antigen (12). More importantly, this population was the helper population because it was required for the optimal development of the TH₂⁺ (OKT5⁺) cytotoxic cell in cell-mediated lympholysis, induction of B cell differentiation, proliferation and immunoglobulin synthesis in a pokeweed-mitogen-driven system, and production of helper factors (13–16). Thus, the OKT4⁺ and OKT5⁺ T cells belonged to reciprocal subsets, and defined the human inducer (helper), and suppressor, T cell subpopulations, respectively. Because of the importance of relating T lymphocyte

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subsets defined by both monoclonal antibodies and Fc receptors to one another, these populations were isolated and compared.

Materials and Methods

Separation of T Cell Subpopulations by Differential Fc-Receptor Binding. Separation of unfractionated T cells, T γ -enriched, T γ -depleted, T μ , and T cells depleted of T γ and T μ subpopulations (T_{Null}) populations was performed as described in detail elsewhere (4). Purity of T γ and T μ subpopulations was >96% by analysis of Fc-receptor rosetting.

Separation of OKT4- and OKT5-reactive Populations. Production, growth, and characterization of the hybridoma-secreted monoclonal antibodies OKT4 and OKT5 were the subject of prior reports (12, 17). With fluorescence-activated cell sorter (FACS-I) (Becton, Dickinson, FACS Systems, Mountain View, Calif.), T cells were separated into OKT4⁺ and OKT5⁺ populations as previously described, and then cultured in 20% fetal calf serum overnight (12). Purity and viability of the FACS-separated T cell subsets were both >95%. Enumeration of T γ and T μ cells within the unfractionated T cell population, the unfractionated T cell population incubated with monoclonal antibody and fluorescein-conjugated goat anti-mouse IgG (G/M FITC) (antibody treated [Rx]), and FACS-separated OKT4⁺ and OKT5⁺ subsets was determined as previously described (2).

Surface Markers on Separated Lymphocyte Subpopulations. T cell subpopulations obtained by Fc-receptor rosetting were analyzed after 24 h with monoclonal antibodies. In addition to OKT4 and OKT5, other monoclonal antibodies were utilized; one, termed OKT3, was previously shown to react with 100% of peripheral T cells and no other lymphoid elements (13, 14); a second, termed OKI1, was shown to react with the bimolecular glycoprotein complex of 29,000- and 34,000-dalton subunits of the Ia molecule in man (14); and a third, termed OKM1, was utilized to define monocytes because it reacts selectively with virtually all adherent macrophages required for antigen presentation in T cell proliferation and a population of nonadherent monocytes.¹ Indirect immunofluorescence was performed with monoclonal antibodies at a 1:250 dilution as described in prior reports. The negative control ascites was OKT6, a monoclonal antibody reactive selectively with human thymocytes but not peripheral lymphocytes (18).

Results and Discussion

Unfractionated T cells and T γ - and T μ -enriched subpopulations were analyzed from a single individual by the use of monoclonal antibodies and indirect immunofluorescence on the FACS. As shown by the representative case in Fig. 1, ~95% of the unfractionated E-rosetting population was reactive with the OKT3 monoclonal antibody, whereas 65% were reactive with OKT4 and 20% were reactive with OKT5. The 5% of cells forming rosettes with sheep erythrocytes (E-RFC⁺) unreactive with OKT3 were reactive with OKM1, a hybridoma antibody that defines an antigen present on monocytes. In addition, 3% of cells in the same unfractionated T cell population were specifically reactive with OKI1 defining the Ia antigen in man.

The T μ population was similar to the unfractionated T cell population with respect to reactivity with OKT3, OKT4, and OKT5 (Fig. 1). However, unlike the unfractionated T cell population, OKI1- and OKM1-reactive cells had been depleted after the removal of the T γ population. In contrast, the T γ -enriched population contained only 10% T cells as defined by OKT3 monoclonal antibody. Within this T cell fraction, there was no selectivity for either the OKT4⁺ inducer or OKT5⁺ suppressor T cell subset, because both were represented in the same ratio as found in the unseparated T cell population. More importantly, it was found that the major portion of lymphocytes in the T γ -enriched population were mononuclear cells that bore the

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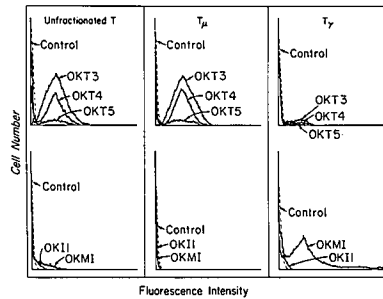


FIG. 1. Reactivity of unfractionated T cells and T cell subsets defined by Fc receptors with monoclonal antibodies utilizing indirect immunofluorescence on FACS. See Results and Discussion for additional information.

OKM1 marker. In addition, it should be noted that the T_{γ} cells were predominantly Ia^{-} as determined by reactivity with OKI1.

Table I provides results from two of five additional experiments in which T_{γ} -enriched, T_{γ} -depleted, T_{μ} , T_{Null} , and unfractionated T populations were reacted with these monoclonal antibodies. The data clearly demonstrate the reactivities of T_{γ} -depleted, T_{μ} , and T_{Null} subpopulations were virtually identical to the unfractionated population. Again, the T_{γ} -enriched population was largely composed of cells that lacked reactivity with all of these T cell monoclonal antibodies and that shared reactivity with a monocyte-reactive monoclonal antibody, OKM1 (18).

Thus, the T_{γ} -enrichment procedure did not select for subsets of T cells as defined by these monoclonal antibodies. Three additional experiments gave similar results and showed that the T_{γ} -enriched population was heterogeneous because anywhere from 5 to 50% of cells were OKT3 reactive, 50 to 90% were OKM1 reactive, and 3 to 25% were OKI1 reactive. It should be noted that a prior study demonstrated the existence of OKM1⁺ mononuclear cells that were predominantly Ia^{-} .¹

To determine the relationship of separated OKT4⁺ and OKT5⁺ T cell subsets to the T_{γ} and T_{μ} lymphocyte subpopulations, unfractionated T cells or FACS-separated T cell subsets were rosetted with ox erythrocytes that were coated with either rabbit IgG or IgM antibodies. Subsequently, the percentages of T_{γ} and T_{μ} cells were enumerated. As shown in Table II, the unfractionated T cells, the unfractionated T cells treated with monoclonal antibody and G/M FITC (OKT4 Rx or OKT5 Rx), and OKT4⁺ or OKT5⁺ T cell subsets were all comparable in their percentage of T_{γ} -reactive cells. Thus, isolation of either OKT4⁺ or OKT5⁺ T cell subsets provided no significant enrichment for the T_{γ} population. However, Table II also shows that there was a partial enrichment of the T_{μ} -rosetting cells in the OKT4⁺ subset. These latter results supported the notion that the population of T cells defined by the monoclonal antibody OKT4 and that defined by T_{μ} were in part overlapping. This was not surprising in light of prior functional studies which demonstrated that both T_{μ} and OKT4⁺ T cell subsets contained the T inducer population for B cell-immunoglobulin production (4, 15).

Other than the partial enrichment of T_{μ} cells within the OKT4⁺ subset, there was generally little correlation in the cell subpopulations defined by monoclonal antibodies and those defined by Fc receptors. Moreover, the T_{γ} -depleted, T_{μ} , and T_{Null} subpopulations defined by Fc receptors were virtually indistinguishable from the unfraction-

TABLE I
Reactivity of T Cell Subsets Defined by Fc Receptors with Monoclonal Antibodies

Experimental number	Monoclonal antibody	Unfractionated T	T γ enriched	T γ depleted	T μ	T _{Nat}
1	OKT3	93	14	93	95	89
	OKT4	65	8	64	69	58
	OKT5	25	7	25	25	25
	OKI1	5	3	2	1	3
	OKM1	6	50	3	1	8
2	OKT3	84	15	90	95	85
	OKT4	61	10	68	73	55
	OKT5	22	7	25	18	25
	OKI1	4	8	1	1	0
	OKM1	18	88	3	2	7

ated T cell population in reactivity with OKT3, OKT4, and OKT5. The single exception was the T γ -enriched population, which had few T cells as defined by OKT3 and had a large number of OKM1⁺ cells. In further support of these latter findings were the previous functional studies that demonstrated that the unfractionated T cell population and the T μ subpopulation were similar. Thus, both unfractionated T cells and T μ cells could provide help for B cell-immunoglobulin secretion, effect cell-mediated lympholysis, and be induced by concanavalin A (Con A) to suppress (4, 19, 20). In contrast, it has been shown that the OKT4⁺ subset alone could provide help in T-T and T-B interactions, whereas the OKT5⁺ subset alone could effect cell-mediated lympholysis and was induced by Con A to suppress (12, 15, 17).

The T γ population of cells was distinct from the unfractionated T cells and all other subsets of T cells isolated with Fc rosetting by virtue of its small percentage (15%) that showed reactivity with the T cell-specific monoclonal antibodies and its large percentage (50–88%) that showed reactivity with the OKM1 monoclonal-antibody defining monocytes. Although both T γ and OKT5⁺ subpopulations can be induced to suppress with Con A, there are functional differences of note between these populations (17, 19, 20). Specifically, OKT5⁺ cells proliferate in mixed lymphocyte cultures and effect cell-mediated-lympholysis, whereas T γ cells do not. Conversely, T γ cells effect natural-killer activity and antibody-dependent cellular-cytotoxic function that are not properties of unactivated OKT5⁺ cells (17, 21, 22).

Our results indicate that circulating lymphoid cells identified as T γ cells (i.e., nonadherent E-RFC⁺ and Fc-receptor-portion of IgG positive) include at least two cell types: (a) a small population reactive with OKT3, and (b) a large population reactive with OKM1. Those cells expressing T cell antigens recognized by our monoclonal antibodies averaged only 22% of the T γ preparations in these six experiments. The lack of reactivity of most T γ cells with three monoclonal antibodies thought to define all peripheral T cells and a monoclonal antibody specific for human thymocytes (OKT6) suggests that the majority that displayed distinct morphologic and histochemical features (23) are neither mature nor immature cells of T lineage (12–17, 18). Moreover, the reactivity with the OKM1 monoclonal antibody that defined an antigen shared by monocytes and granulocytes implies that they may rather be of monocyte-myeloid lineage. We suggest that these cells are the natural-killer cells.

TABLE II
Capacity of Monoclonal-Antibody-separated T Cell Subsets to Rosette with Ox
Erythrocytes Coated with IgG and IgM

Cell Population	Experiment 1		Experiment 2	
	Fc γ †	Fc μ §	Fc γ †	Fc μ §
	%			
E-RFC ⁺	4.0	57	7	58
OKT4 R _x *	5.5	48	7	61
OKT4 ⁺	2.5	88	1	90
OKT5 R _x *	6.5	56	6.5	58
OKT5 ⁺	2.5	34	7	50

* Control T cell populations were reacted with either OKT4 or OKT5 and G/M FITC but not sorted on FACS.

Summary

Human T cell subpopulations have been defined on the basis of differential expression of either Fc receptors or specific cell-surface antigens. In this study, we utilized a series of monoclonal antibodies reactive with T cells, monocytes, and Ia antigens to characterize isolated subpopulations of T cells bearing receptors for the Fc portion of IgG (T γ) and subpopulations of T cells bearing receptors for the Fc portion of IgM (T μ). The results showed that the T μ population contained both inducer (OKT4⁺) and cytotoxic/suppressor (OKT5⁺) populations and was similar to the unfractionated T cell population, whereas the T γ subset contained few T lymphocytes (OKT3⁺) and was not enriched for either T cell subset defined by these monoclonal antibodies. Rather, the T γ population was comprised largely of Ia⁻ cells possessing a monocyte antigen (OKM1⁺). In reciprocal studies, it was found that both isolated OKT4⁺ and OKT5⁺ T cell subsets contained few T γ cells, whereas both subsets were mainly comprised of T μ cells. We conclude that there is little correlation between T cell subsets defined by these monoclonal antibodies and those defined by Fc receptors.

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