

BPAG1n4 is essential for retrograde axonal transport in sensory neurons

Jia-Jia Liu,¹ Jianqing Ding,¹ Anthony S. Kowal,¹ Timothy Nardine,¹ Elizabeth Allen,¹ Jean-Dominique Delcroix,¹ Chengbiao Wu,¹ William Mobley,¹ Elaine Fuchs,² and Yanmin Yang¹

¹Department of Neurology and Neurological Sciences, Stanford University School of Medicine, Stanford, CA 94305

²Howard Hughes Medical Institute, Laboratory of Mammalian Cell Biology and Development, The Rockefeller University, New York, NY 10021

Disruption of the BPAG1 (bullous pemphigoid antigen 1) gene results in progressive deterioration in motor function and devastating sensory neurodegeneration in the null mice. We have previously demonstrated that BPAG1n1 and BPAG1n3 play important roles in organizing cytoskeletal networks *in vivo*. Here, we characterize functions of a novel BPAG1 neuronal isoform, BPAG1n4. Results obtained from yeast two-hybrid screening, blot overlay binding assays, and coimmunoprecipitations demonstrate that BPAG1n4 interacts directly with dynactin

p150^{Glued} through its unique ezrin/radixin/moesin domain. Studies using double immunofluorescent microscopy and ultrastructural analysis reveal physiological colocalization of BPAG1n4 with dynactin/dynein. Disruption of the interaction between BPAG1n4 and dynactin results in severe defects in retrograde axonal transport. We conclude that BPAG1n4 plays an essential role in retrograde axonal transport in sensory neurons. These findings might advance our understanding of pathogenesis of axonal degeneration and neuronal death.

Introduction

The BPAG1 (bullous pemphigoid antigen 1) null mouse has a fascinating and complex phenotype, displaying sensory ataxia and devastating sensory neurodegeneration (Guo et al., 1995). This degeneration is characterized by neurofilament disorganization, microtubule network abnormality, and intermittent accumulation of membranous organelles along sensory axons. The BPAG1 gene encodes multiple isoforms with BPAG1e expressed in the epidermis and the others (BPAG1n1, BPAG1n2, and BPAG1n3) expressed in neurons. Throughout normal development, mRNAs of BPAG1 neuronal isoforms are present in a variety of neurons. Postnatally, this expression is absent or reduced in motor neurons and restricted to sensory neurons (Brown et al., 1995; Dowling et al., 1997). It has been demonstrated that BPAG1n1 links intermediate filaments to actin microfilaments and that BPAG1n3 can stabilize microtubules (Yang et al., 1996, 1999). However, the mechanisms underlying the devastating sensory neuron degeneration in BPAG1 null mice await further characterization.

Impaired axonal transport in neurons has long been implicated as a mechanism underlying axonal degeneration

and neuronal death. In neurons, motor protein of kinesin superfamily drives anterograde transport, whereas cytoplasmic dynein/dynein powers retrograde transport. The mutations discovered in critical components of transport pathways provide evidence for the notion that axonal transport is essential for neuronal survival. Transgenic mice carrying mutant superoxide dismutase-1, mouse models of amyotrophic lateral sclerosis, show deficits in slow axonal transport early in the disease course (Williamson and Cleveland, 1999). Additionally, in some superoxide dismutase-1 mutant lines, an early up-regulation of the kinesin superfamily motor protein KIF1A was detected in spinal motor neurons (Dupuis et al., 2000). Mutations in the gene encoding KIF1B β cause an axonal form of a hereditary neuropathy (Zhao et al., 2001). A recent work has linked mutations in cytoplasmic dynein heavy chain with defects in retrograde transport that lead to motor neuron degeneration (Hafezparast et al., 2003). Furthermore, a mutation in the p150^{Glued} subunit of dynactin has been identified in a family with a slowly progressive, autosomal-dominant form of lower motor neuron disease in which sensory symptoms are absent (Puls et al., 2003).

Address correspondence to Yanmin Yang, Department of Neurology, Stanford University School of Medicine, 1201 Welch Rd., MSLS P207, Stanford, CA 94305-5489. Tel.: (650) 736-1032. Fax: (650) 498-6262. email: yanmin.yang@stanford.edu

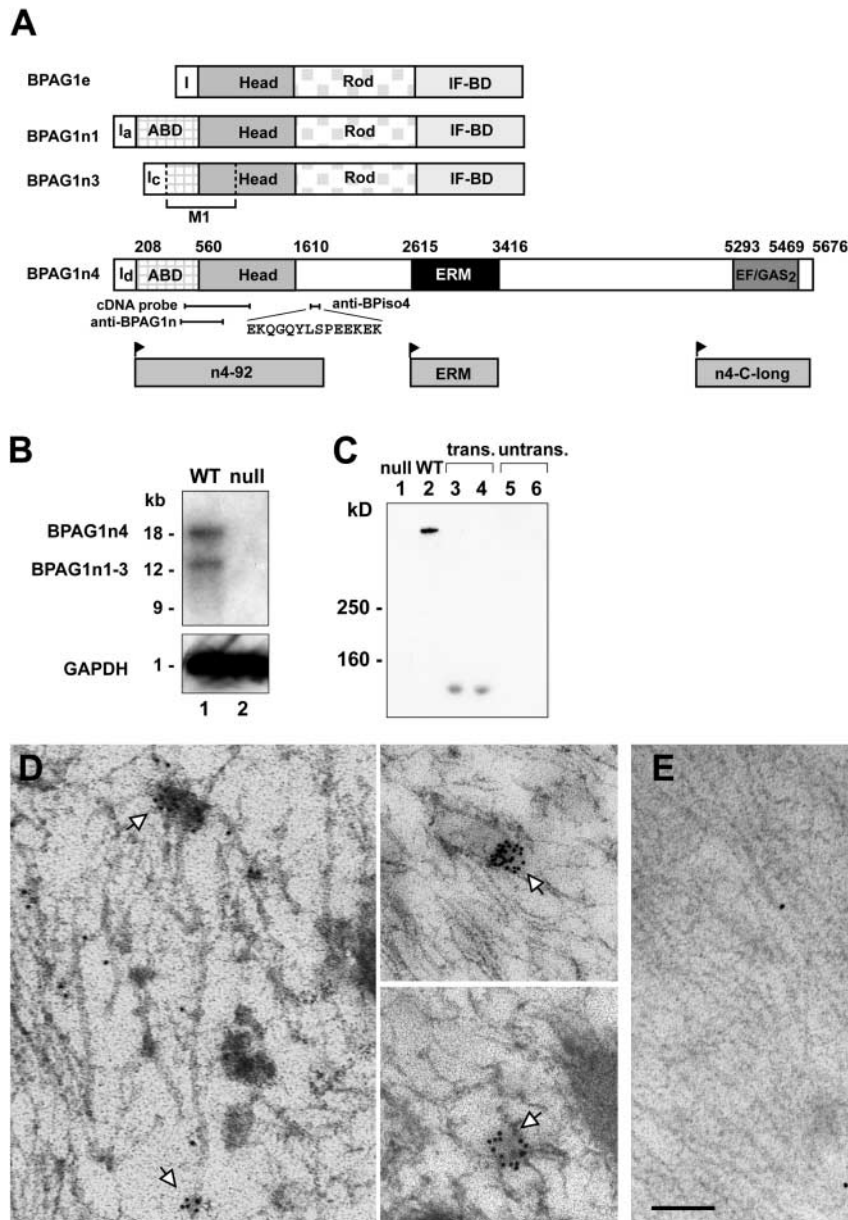
Key words: cytoskeleton; BPAG1n4–dynactin interaction; axonal transport; neurodegeneration; ERM domain

Abbreviations used in this paper: BPAG1, bullous pemphigoid antigen 1; co-IP, coimmunoprecipitation; DRG, dorsal root ganglion; ERM, ezrin/radixin/moesin; immunoEM, immunoelectron microscopy; Tf-TR, transferrin conjugated with Texas red; WT, wild type.

Figure 1. Characterization of BPAG1n4.

(A) Schematic of the domain structures of BPAG1 isoforms. I, first exon of each isoform; BPAG1e, the epithelial isoform; ABD, actin-binding domain; IF-BD, intermediate filament-binding domain; M1, microtubule-binding domain. Amino acid residue numbers denote functional domain boundaries in BPAG1n4. Individual domains and regions used in this work are illustrated below. Anti-BPAG1n recognizes all BPAG1 isoforms.

(B) Northern blot analysis of mouse dorsal root ganglion (DRG) RNA. (probes) A 1.6-kb cDNA head domain fragment (top; 4-d exposure); GAPDH as internal control (bottom; 1-h exposure). (C) Protein expression of BPAG1n4 in mouse brain. Blot was probed with anti-BPiso4. COS-7 (lane 3) and NIH 3T3 cells (lane 4) were transfected with an expression construct for n4-92 (A, bottom) encompassing the epitope for anti-BPiso4. (untrans.) Untransfected COS-7 (lane 5) and NIH cells (lane 6). (D–E) Immunocytochemistry reveals subcellular localization of BPAG1n4 in dorsal roots. (D) BPAG1n4 labeling with anti-BPiso4 was visualized with 12-nm gold particles conjugated with secondary antibody. White arrows indicate BPAG1n4 on vesicle-like structures in association with microtubules. (E) Negative control, secondary antibodies alone. Bar: (D) 300 nm; (E) 400 nm.



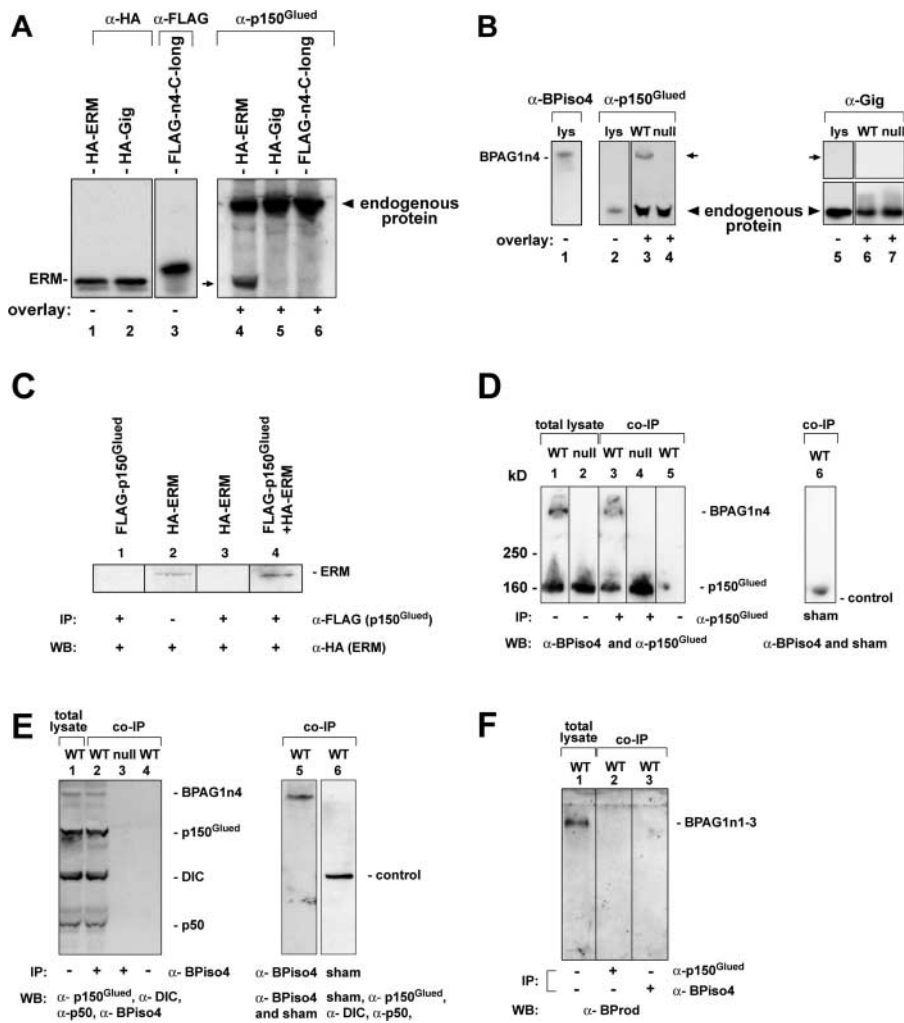
In this work, we analyze the functions of BPAG1n4, the fourth neuronal isoform of the BPAG1 gene family. BPAG1n4, referred to as BPAG1a by Leung et al. (2001), harbors a structurally and functionally unique ezrin/radixin/moesin (ERM) domain (Burrige and Mangeat, 1984; Anderson and Marchesi, 1985; Lankes and Furthmayr, 1991). We provide biochemical evidence that the ERM domain of BPAG1n4 directly interacts with dynactin, and show that this isoform physiologically colocalizes with the dynactin–dynein complex in vivo in sensory axons. Finally, we demonstrate that disruption of the interaction between p150^{Glued} dynactin and BPAG1n4 leads to a failure of retrograde axonal transport in sensory neurons.

Results and discussion

BPAG1n4 localizes to vesicle-like structures

Although BPAG1n4 and BPAG1n1 share a conserved NH₂-terminal actin-binding domain, BPAG1n4's COOH termi-

nus harbors a distinctive functional microtubule-binding domain (Leung et al., 1999; unpublished data). This domain features two EF-hand Ca²⁺-binding motifs (Ikura, 1996) immediately upstream of a short stretch of GAS2 motif (EF-hand/GAS2; Schneider et al., 1988). Furthermore, a central ERM domain (Burrige and Mangeat, 1984; Anderson and Marchesi, 1985; Lankes and Furthmayr, 1991) is flanked by two functional cytoskeleton-binding domains (Yang et al., 1996; Leung et al., 1999; Fig. 1 A). Human BPAG1n4 shares ~82% sequence identity with a recently published mouse form, BPAG1a (Leung et al., 2001). The expression of BPAG1n4 was detected at both the mRNA and protein levels in wild type (WT), but not in null tissues (Fig. 1, B and C). The specificity of anti-BPiso4 was confirmed in cells transiently expressing a FLAG epitope-tagged BPAG1n4 NH₂-terminal segment (Fig. 1 C, lanes 3 and 4) that was also recognized by anti-FLAG (not depicted). Immunohistochemical studies using anti-BPiso4 antibody recapitulated our previously published staining pattern charac-



coimmunoprecipitate BPAG1n4 (D, lane 3). Conversely, anti-BPiso4 but not the sham antibody was able to coimmunoprecipitate the dynactin complex (E, lane 2). Antibodies against BPAG1n4, p150^{Glued}, p50, DIC, and control protein (sham) were used for immunoblotting. (F) Neither anti-p150^{Glued} nor anti-BPiso4 could coimmunoprecipitate other BPAG1 neuronal isoforms (BPAG1n1–3). Immunoblot was probed with rabbit antibody against the common rod domain shared by BPAG1n1–3 (anti-BProd).

teristic of sensory neurons using anti-BPAG1n antibody, which recognizes all isoforms (Yang et al., 1996; Dowling et al., 1997). No significant staining was found in motor neurons of postnatal animals (Yang et al., 1996).

As a first step toward understanding the function of this new isoform, anti-BPiso4 antibody was used to determine the subcellular localization of BPAG1n4 in the spinal cord dorsal column by immunoelectron microscopy (immunoEM). Intriguingly, ~72% of BPAG1n4-associated particles localized to vesicle-like structures associated with microtubules (Fig. 1 D, gold particles). No significant labeling was detected in negative controls using secondary antibodies only (Fig. 1 E).

BPAG1n4 interacts directly with dynactin p150^{Glued}

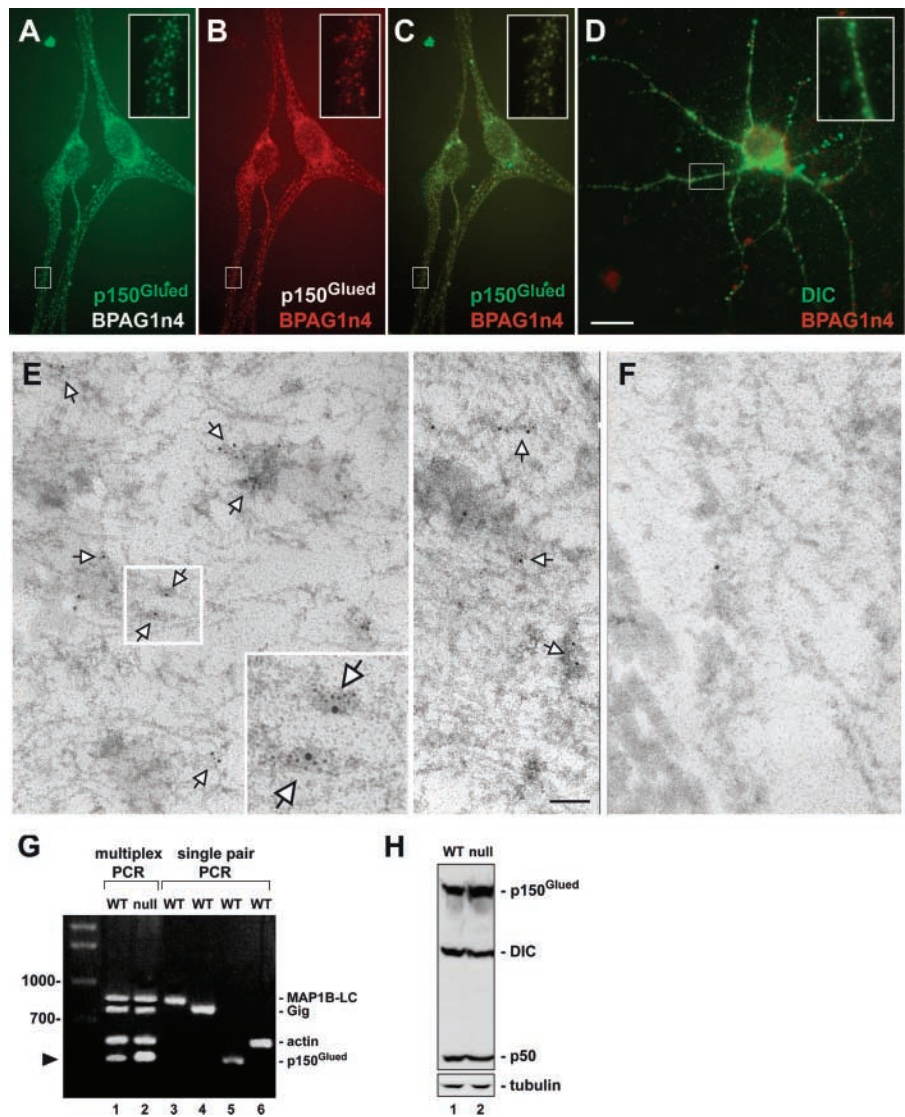
ERM domain containing proteins are known to modify the interaction of cytoskeletal and integral membrane proteins (Burrige and Mangeat, 1984; Anderson and Marchesi, 1985; Lankes and Furthmayr, 1991). We used the ERM domain of BPAG1n4 as bait in a yeast two-hybrid screen (Ding et al., 2002) of a human brain cDNA library. 28 pu-

tative positive clones exclusively identified the COOH-terminal region of dynactin p150^{Glued} (Gill et al., 1991) as the binding partner for BPAG1n4. Consistent with our two-hybrid results, HA-tagged ERM could be coimmunoprecipitated by FLAG-tagged p150^{Glued} when coexpressed in cells (Fig. 2 C, lane 4). Additional assays using tissue extracts corroborated the interaction of BPAG1n4 and p150^{Glued}, including in vitro blot overlay (Fig. 2, A and B) and coimmunoprecipitations (co-IPs; Fig. 2, D–F). In brief, by in vitro blot overlay binding assay, p150^{Glued} bound only to the immobilized HA-tagged ERM domain (Fig. 2 A, lane 4) or to BPAG1n4 (Fig. 2 B, lane 3), but not to control proteins (Fig. 2 A, lanes 5 and 6; and Fig. 2 B, lanes 4, 6, and 7). In co-IP experiments from tissues, BPAG1n4 was specifically detected in the complex coimmunoprecipitated by anti-p150^{Glued} (Fig. 2 D, lane 3). Conversely, p150^{Glued} and p50, another dynactin subunit, as well as DIC but not a control protein, were specifically coimmunoprecipitated by anti-BPiso4 (Fig. 2 E, lane 2 and 5, respectively). Neither the secondary alone nor a sham antibody pulled down BPAG1n4 (Fig. 2 D, lanes 5 and 6) or dynactin (Fig. 2 E, lane 4 and 6).

Figure 2. Blot overlay and co-IP assays show BPAG1n4–dynactin interaction.

(A) Blot overlay assays with ERM domain, 85-kD epitope-tagged ERM (HA-ERM), 78-kD gigaxonin (HA-Gig), or 90-kD BPAG1n4 COOH terminus (FLAG-n4-C-long) from transfected CHO cells were immobilized on membranes. Membranes were incubated with (+) or without (–) total lysates of mouse spinal cord and sciatic nerve and probed with anti-HA (HA.11; CRP, Inc.), anti-FLAG (Sigma-Aldrich), and anti-p150^{Glued}, respectively. In addition to its endogenous band (arrowhead), p150^{Glued} was also detected at the position of HA-ERM (lane 4, arrow) as a result of its binding to the ERM domain of BPAG1n4. (B) Blot overlay assays with BPAG1n4. BPAG1n4 was immobilized on membrane and overlaid with (+) or without (–) total lysates of spinal cord and sciatic nerve. Blots were probed with anti-p150^{Glued} and antigigaxonin (Gig, control). (lys) total brain lysates. p150^{Glued} (lane 3), but not gigaxonin (lane 6), binds to full-length BPAG1n4 (arrows). (C) Co-IP of ERM domain and p150^{Glued} in COS-7 cells. (WB) Western blotting; (IP) immunoprecipitation. Cell lysates of COS-7 transfected with pHA-ERM (lanes 2 and 3), or pFLAG-p150^{Glued} (lane 1), or both (lane 4) were incubated with anti-FLAG M2 affinity gel. Protein samples were analyzed by immunoblotting with anti-HA antibody. (D–E) co-IP of BPAG1n4 and the dynactin–dynein complex from mouse nerve tissue extracts. A rabbit anti-p150^{Glued} (Santa Cruz Biotechnology, Inc.) antibody, but not the sham antibody, was able to

Figure 3. Colocalization of BPAG1n4 with dynein/dynein and up-regulation of p150^{Glued} in BPAG1 null mouse. (A–D) Double immunofluorescence staining of cultured DRG neurons shows colocalization of BPAG1n4 with dynein/dynein. Cells were stained with anti-BPiso4/anti-p150^{Glued} (BD Biosciences) in A–C and anti-BPiso4 /anti-DIC (Chemicon) in D. AlexaFluor-conjugated secondary antibodies (Molecular Probes) were used for detection. Insets show colocalizations at higher magnification. (E) Double ImmunoEM labeling of mouse sciatic nerve sections with anti-BPiso4 (18 nm) and anti-DIC (6 nm). Arrows indicate colocalization of BPAG1n4 and dynein. (F) Negative control with secondary antibodies only. (G) RT-PCR analysis of DRG mRNAs. Multiplex gene-specific primers (lanes 1 and 2) were used on cDNAs from WT and null DRG. Lanes 3–6 indicate each individually amplified primer pair product from WT: p150^{Glued} (410 bp), microtubule associated protein 1B light chain (MAP1B-LC; 850 bp), gigaxonin (Gig, 780 bp), and actin (560 bp). Arrowhead indicates up-regulation of p150^{Glued} in BPAG1 null (lane 2) versus WT (lane 1). (H) Protein analysis on mouse spinal cord and sciatic nerve lysates using antibodies to p150^{Glued}, DIC, and p50 (BD Biosciences). Antitubulin served as loading control. Bars: (A–D) 10 μ m; (E and F) 400 nm.



Neuronal isoforms BPAG1n1–3 were detected in total lysates (Fig. 2 F, lane 1) using anti-BPprod antibody, which recognizes the domain common to these isoforms, but not in complexes precipitated by either anti-p150^{Glued} or anti-BPiso4 (Fig. 2 D, lanes 2 and 3). Together, our results provide strong evidence that BPAG1n4 associates with dynein p150^{Glued} via its ERM domain.

The dynein complex is required for functional cytoplasmic dynein (Waterman-Storer et al., 1997; Holleran et al., 1998; King and Schroer, 2000; Deacon et al., 2003), a motor protein that powers retrograde transport. co-IP results suggested that, through interaction with p150^{Glued}, BPAG1n4 might indirectly associate with dynein in vivo. We performed double immunofluorescence staining on cultured dorsal root ganglion (DRG) neurons and observed striking colocalizations of BPAG1n4 with p150^{Glued} (Fig. 3, A–C) and with DIC (Fig. 3 D) in neurites. Double immunocytochemistry on mouse sciatic nerve provided additional in vivo evidence for colocalization of BPAG1n4 and dynein. Approximately 71% of the BPAG1n4-associated gold particles colocalized with 44% of the dynein-associated particles (Fig. 3 E).

Intriguingly, quantitative multiplex RT-PCR analysis on DRG extracts from WT and BPAG1 null mice revealed a specific up-regulation of p150^{Glued} mRNA expression in null mice (Fig. 3 G, lane 2). Actin, gigaxonin (Bomont et al., 2000; Ding et al., 2002), and MAP1B-LC were used as controls and showed no corresponding up-regulation (Fig. 3 G, lane 2). Similarly, analysis of protein extracts from spinal cord and sciatic nerves revealed a substantial increase of p150^{Glued} expression (but not of p50 or DIC) in BPAG1 null mice (Fig. 3 H). Specific up-regulation of p150^{Glued} expression in the null mice might represent a compensatory response for loss of BPAG1n4 as an interaction partner.

BPAG1n4 is required for retrograde axonal transport in sensory neurons

At the ultrastructural level, vesicles, multivesicular bodies, mitochondria, and other membranous organelles were found to accumulate in BPAG1 null sensory axons, a phenotype characteristic of impaired axonal transport (Fig. 4 A). In contrast, WT axons displayed no such accumulations (Fig. 4 B). We analyzed axonal transport by double-ligation

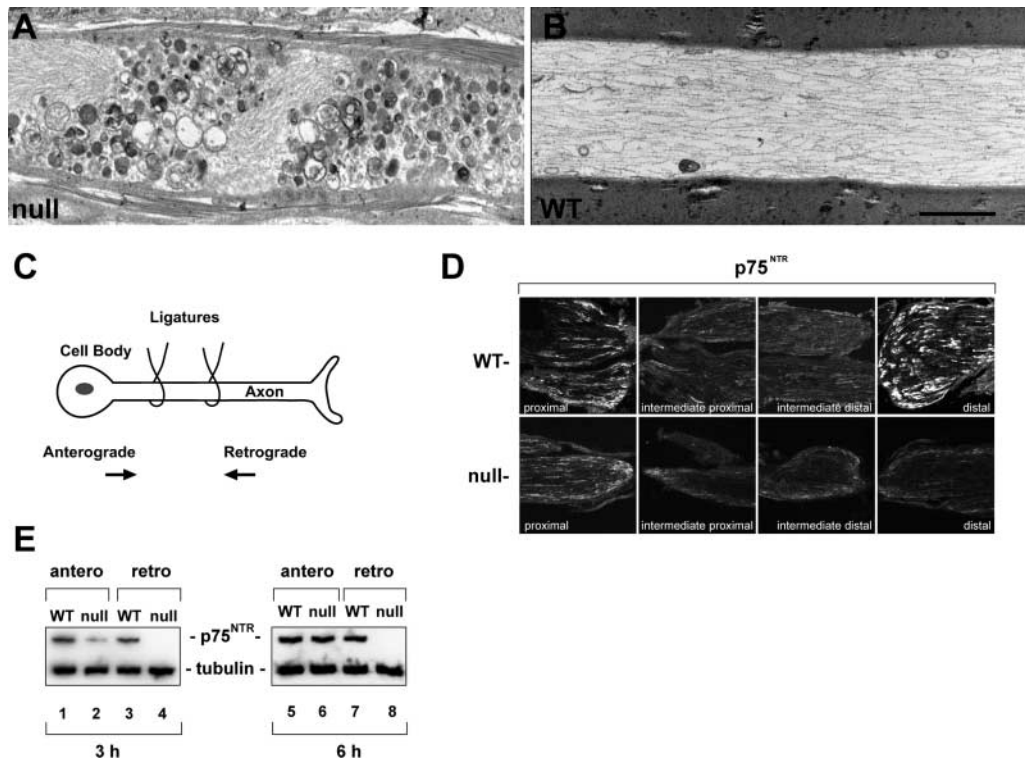


Figure 4. Retrograde axonal transport is disrupted in BPAG1 null mice. (A and B) EM of longitudinal sections of spinal cord dorsal column reveals accumulation of vesicles and other organelles in BPAG1 null (A), but not in WT (B). (C–E) Proteins failed to accumulate on the distal side of a double ligation (schematic shown in C) of BPAG1 null sciatic nerves. Shown are confocal images of nerve sections stained with anti-p75^{NTR} (D), and immunoblots of p75^{NTR} from ~3-mm sections proximal or distal to double ligatures (E). Anti- α -tubulin served as loading control. Bar: (A and B) 500 nm; (D) 140 μ m.

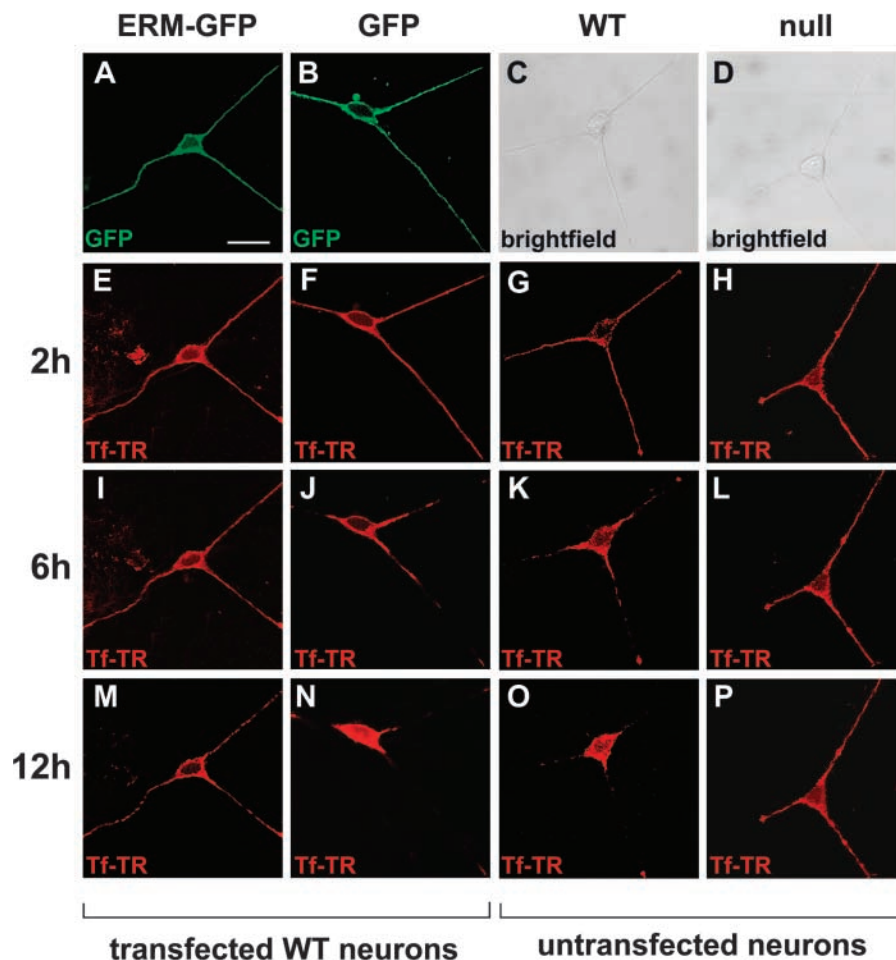
of sciatic nerves (Fig. 4 C), monitoring a sensory neuron protein, p75^{NTR} (Raivich et al., 1991), for any subsequent accumulation. Immunofluorescence staining revealed that in BPAG1 null mice p75^{NTR} failed to accumulate at distal sites 6 h after ligation (Fig. 4 D, bottom, distal), but exhibited proximal accumulations (Fig. 4 D, bottom, proximal) surprisingly similar to those observed in WT (Fig. 4 D, top). Quantitative biochemical assays were conducted on ~3-mm segments proximal and distal to the double ligatures at 3- and 6-h time points. In WT mice, a substantial accumulation of p75^{NTR} was observed bidirectionally at both time points (Fig. 4 E, lanes 1 and 5, and lanes 3 and 7, respectively). In contrast, in the null mice, the accumulation of p75^{NTR} was barely detectable at both directions 3 h after ligation (Fig. 4 E, lanes 2 and 4). Interestingly, at the 6-h time point the retrograde accumulation remained undetectable in the null mice (Fig. 4 E, lane 8), but at anterograde direction the difference appears less marked (Fig. 4 E, lane 6). These results suggest that, whereas the axonal transport in BPAG1 null mice is bidirectionally impaired, the retrograde direction is apparently more severely affected.

The gene targeting strategy in BPAG1 null mice ablated all isoforms (Guo et al., 1995), all of which could collectively play important roles bidirectionally in axonal transport. However, the specific interaction of BPAG1n4 and dynein/dynein *in vivo* pointed to a potential role for the BPAG1n4 isoform in retrograde axonal transport. To test this hypothesis, we overexpressed an ERM-GFP fusion pro-

tein in cultured mouse DRG neurons. ERM-GFP and GFP proteins were efficiently transported to the neurite tips in transfected neurons (Fig. 5 A–B). To examine retrograde transport, the neurons were incubated for 2 h with media containing the tracer transferrin conjugated with Texas red (Tf-TR). The media were replaced with tracer-free media, and retrograde transport of Tf-TR was subsequently monitored at 2-h time points. In the first 2 h after removing Tf-TR, no significant differences were observed in the cultured neurons (Fig. 5, E–H). However, after 4–6 h, the WT and GFP-expressing neurons had begun to accumulate Tf-TR in their cell bodies (Fig. 5, K and J, respectively). After 12–14 h, ~94% of these neurons had completed retrograde transport of the Tf-TR pulse (Fig. 5, N and O). In contrast, in three independent experiments, 82% of the ERM-GFP-overexpressing neurons showed severe defects identical to those seen in 91% of BPAG1 null neurons, namely, a nearly complete failure to transport Tf-TR to the cell bodies (Fig. 5, I and M, and L and P, respectively). Together, these results demonstrate that the isolated ERM is sufficient to disrupt BPAG1n4's function in sensory neurons in a dominant-negative fashion, leading to a failure of retrograde axonal transport that could account for the impaired retrograde axonal transport found in the BPAG1 null mouse. We conclude that BPAG1n4 is essential for retrograde axonal transport in sensory neurons. Because the postnatal expression of BPAG1 neuronal isoforms is restricted to sensory neurons (Yang et al., 1996), it is likely that a related molecule ex-

Figure 5. Disruption of retrograde transport is BPAG1n4 isoform specific.

Mouse DRG neurons were transfected with pEGFP-ERM-N1 or pEGFP-N1. Retrograde transport of transferrin conjugated with Texas red (Tf-TR) was examined as described in Materials and Methods. Cells expressing ERM-GFP (A) or GFP (B) were first incubated with Tf-TR for 2 h. After removal of the Tf-TR, the cells were observed and pictured every 2 h. Shown are the cells after 2 h (E–H), 6 h (I–L), and 12 h (M–P). Untransfected DRG neurons of WT (C, G, K, and O) and BPAG1 null mice (D, H, L, and P) served as controls. The ERM-GFP-expressing neurons mimicked BPAG1 null neurons in their failure to transport Tf-TR retrogradely into the cell bodies. Bar, 23 μ m.



pressed in motor neurons, such as ACF7 (Byers et al., 1995; Karakesiosoglou et al., 2000), may play a corresponding role in axonal transport in those cells. Alternatively, motor neurons may rely on different mechanisms for such a process.

Defects in axonal transport have been implicated in many human neurodegenerative disorders (Williamson and Cleveland, 1999; Hafezparast et al., 2003; Puls et al., 2003). The extraordinary length of axons and unique features of neurons may require a more complex system to meet the unusual transport challenges that are far beyond those of non-neuronal cells. Our findings pertaining to BPAG1n4 may contribute to a better understanding of relations between neurodegeneration and axonal transport. The next challenge will be to understand how the interactions of BPAG1n4 with dynactin/dynein and cytoskeletons facilitate retrograde axonal transport in sensory neurons.

Materials and methods

Molecular cloning of BPAG1n4

Complementary DNAs of BPAG1n4 were amplified by rapid amplification of cDNA ends-PCR in human brain Marathon-Ready cDNA (CLONTECH Laboratories, Inc.). Sequencing information from three overlapping clones of long-range PCR products (A, 8.1 kb; B, 7.2 kb; and C, 7.8 kb) confirmed that we obtained the full-length cDNA sequence of BPAG1n4.

Blot overlay binding assays and co-IPs

Nerve tissues (spinal cord and sciatic nerves) were homogenized in PBS with protease inhibitors (Roche), and then briefly centrifuged at 2,300 g for

30 s to remove tissue debris and any particulate materials. The supernatants were divided into two portions, one for further centrifugation at 109,000 g to collect pellets, and two for directly serving as total lysate for overlay and immunoprecipitations. Pellets were resolved on 5% SDS-PAGE and transferred on polyvinylidene fluoride membranes to immobilize the full-length BPAG1n4. The blots were incubated with the total lysates overnight, followed by immunoblot analyses using the indicated antibodies. For co-IP, the rabbit anti-dynactin (H-300; Santa Cruz Biotechnology, Inc.) was used to coimmunoprecipitate BPAG1n4, and anti-BPiso4 was used to coimmunoprecipitate p150^{Glued}. The lysates were incubated with different antibodies and protein A-Sepharose 4B beads (Zymed Laboratories) at 4°C overnight. Beads were washed a few times with PBS. Bound proteins were eluted with SDS sample buffer. Protein samples were resolved through 4–15% gradient SDS-PAGE (Bio-Rad Laboratories) and analyzed by immunoblotting.

ImmunoEM

WT animals were killed by intravenous perfusion with 2% PFA and 0.05% glutaraldehyde. The dissected samples of dorsal roots and sciatic nerves were postembedded as described previously (Yang et al., 1999). The antibody incorporations on ultrathin sections were visualized with 12 nm anti-rabbit for single labeling or 18 nm anti-rabbit and 6 nm anti-mouse gold-conjugated particles (Jackson ImmunoResearch Laboratories). After staining with uranyl acetate, followed by lead citrate, the sections were analyzed under an electron microscope (model CM10; Philips).

Double ligation and immunostaining of sciatic nerves

WT control and BPAG1 null mice (13–17 postnatal days) were anesthetized with a mixture of xylamine/ketamine. On the right sciatic nerve of each mouse, two ligatures 5 mm apart were placed at mid-thigh. For immunostaining, 6 h after ligation, mice were perfused with 10 ml of 0.1 M phosphate buffer, pH 7.4, and 40 ml of fixation solution (4% PFA in PB). Sciatic nerves were postfixed for 2 h and placed overnight in a cryoprotective solution (PB with 15% sucrose). After cryoprotection, sciatic nerves

were embedded in OTC and frozen at -80°C . 8–10- μm sections were cut in a cryostat at -20°C , mounted on glass slides, and stained with anti-p75^{NTR} (Promega), followed by fluorescent dye-conjugated secondary antibodies (Jackson ImmunoResearch Laboratories). Samples were analyzed and images were captured using a confocal microscope (model Radiance200; Bio-Rad Laboratories).

Primary sensory neuron transfection and transferrin transport

DRG neurons from newborn WT mice were transfected with pEGFP-ERM-N1 or the control construct pEGFP-N1. Transfection was performed using the Mouse Neuron Nucleofector™ kit and the Nucleofector™ device (Amaxa). Transfection rates were $\sim 20\%$. Cells were cultured in Neurobasal™ complete medium (Invitrogen) on collagen-coated glass coverslips. For the transferrin transport assay, cells were incubated with medium containing 50 $\mu\text{g}/\text{ml}$ of human transferrin conjugated with Texas red (Tf-TR; Molecular Probes) for 2 h at 37°C to allow for uptake. After removal of the transferrin-containing medium, coverslips were rinsed with PBS and incubated with Tf-TR-free medium at 37°C . Confocal fluorescent images were taken every 2 h with a confocal laser scanning microscope (model LSM 510; Carl Zeiss MicroImaging, Inc.) to monitor the transport of Tf-TR.

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