

Excitation-Contraction Coupling

Orail as a potential "fits-all approach" therapeutic target for the treatment of DMD

Arthur J. Cheng¹, Ferdinand von Walden², and Johanna T. Lanner³

Duchenne muscular dystrophy (DMD) is an X-linked disorder caused by loss-of-function mutations in the dystrophin gene. DMD is a progressive disease that affects 1 in 3,500–5,000 male births and results in muscle weakness, respiratory and cardiac failure, and, in due course, premature death commonly in the third decade of life (Mendell and Lloyd-Puryear, 2013). Histopathological hallmarks of DMD muscles include central nucleation, fibrosis, inflammation, and muscle fiber pseudohypertrophy amongst others (Zweyer et al., 2022). The dystrophin gene was discovered in 1987 (Hoffman et al., 1987), and since then there have been numerous efforts to develop therapies to delay disease progression and enhance muscle function in DMD patients.

Present widespread therapeutic recommendations include corticosteroids, cardiac medications, and, eventually, assisted ventilation to slow down disease progression, reduce symptoms, and improve quality of life, but do not target the underlying mechanism of the disease. However, pharmacological advances using oligonucleotide-mediated exon skipping to bypass variant exons and gene editing to provide truncated forms of dystrophin have gained interest in recent years, and several clinical trials are under way to assess their efficacy (Eser and Topaloğlu, 2022). Although these are promising advancements, there are still challenges to overcome and room for improvement in the area. For example, the TREAT-NMD DMD global database contains over 7,000 dystrophin mutations (Bladen et al., 2015), raising significant challenges for the development of gene correction therapies that might be applicable to large cohorts of patients rather than single individuals. Moreover, present exonskipping therapies focusing on restoring dystrophin at low levels are suggested to have the potential to benefit ~30% of all patients (Eser and Topaloğlu, 2022) and hence also struggle to treat a large cohort of DMD individuals. However, the largest challenge to the field of therapeutic genome editing might be the

high cost of these drugs (Segal, 2022). Nevertheless, there is still a need for curative treatments suitable for a large group of DMD individuals, and pharmacological interventions with a "fits-all approach" are desired. This is one of the reasons the recent publication by García-Castañeda et al. (2022) and colleagues in Dr. Robert T. Dirksen's lab is attractive with its obvious transferability into drug development.

Chronically elevated levels of myoplasmic free [Ca²+] is an acknowledged explanation for the observed skeletal muscle fiber deterioration and loss-of-function in DMD. Two main pathomechanisms have been proposed to cause sustained abnormalities in myoplasmic Ca²+ in DMD (see Fig. 1); enhanced RyR1-mediated Ca²+ leak from the sarcoplasmic reticulum (SR; Bellinger et al., 2009) and excessive extracellular Ca²+ influx via membrane tears and/or store-operated Ca²+ entry (SOCE; Edwards et al., 2010; Zhao et al., 2012). The Ca²+ sensor stromal interaction molecule-1 (STIM1) in the ER/SR membrane and the highly Ca²+-selective Orai channel in the plasma membrane forms the basis for SOCE (Emrich et al., 2022).

Here, García-Castañeda and colleagues used skeletal muscle-specific tamoxifen-inducible Orail-knockout mice crossed with mdx mice to elucidate the impact of Orail-dependent Ca²⁺ entry in the DMD muscle pathology (Fig. 1). This approach allowed them to knockout Orail in the skeletal muscle of young, post-developmental mice and thus establish the effects of inducing a change into Ca²⁺ entry after the disease manifestation. Excitingly, eliminating Orail-mediated Ca²⁺ influx in young (2–3-mo-old) mdx mice resulted in considerable functional and morphological improvements. Skeletal muscle from mdx mice lacking Orail-mediated Ca²⁺ influx exhibited normalized intracellular Ca²⁺ homeostasis and improved muscle strength. Obliterating Orail expression also had a protective effect on the plasma membrane (sarcolemma) in mdx mice as it protected muscles from eccentric contraction-induced damage. Further, muscles from mdx mice

¹School of Kinesiology and Health Sciences, York University, Toronto, ON, Canada; ²Women's and Children's Health, Karolinska Institute, Stockholm, Sweden; ³Physiology and Pharmacology, Karolinska Institute, Stockholm, Sweden.

Correspondence to Johanna T. Lanner: johanna.lanner@ki.se

This work is part of a special issue on excitation-contraction coupling.

© 2023 Cheng et al. This article is distributed under the terms of an Attribution–Noncommercial–Share Alike–No Mirror Sites license for the first six months after the publication date (see http://www.rupress.org/terms/). After six months it is available under a Creative Commons License (Attribution–Noncommercial–Share Alike 4.0 International license, as described at https://creativecommons.org/licenses/by-nc-sa/4.0/).





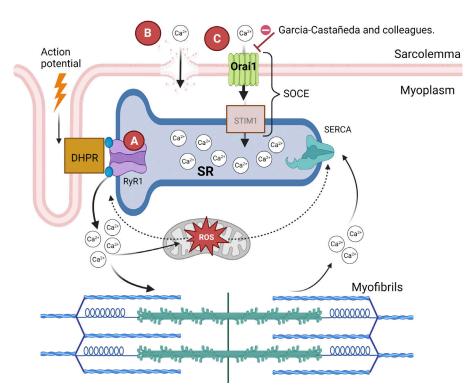


Figure 1. A schematic illustration of the role of Ca²⁺ in the development of skeletal muscle dysfunction in DMD. The two proposed pathways of Ca²⁺-induced muscle dysfunction include (A) increased SR Ca²⁺ leak and extracellular Ca²⁺ entry via (B) sarcolemmal membrane damage or (C) SOCE. Dihydropyridine receptor (DHPR), ryanodine receptor 1 (RyR1), sarcoplasmic reticulum Ca²⁺ ATPase (SERCA), mitochondrial reactive oxygen species (ROS), Orai Ca²⁺ release-activated Ca²⁺ modulator 1 (Orai1), stromal interaction molecule 1 (STIM1). Figure created with Biorender.

with markedly reduced Orail expression displayed fewer histopathological hallmarks, i.e., normalized fiber cross-sectional area (CSA) and reduced muscle fibrosis. However, central nucleation was still present in muscles of mdx mice lacking Orail expression, which indicates ongoing regenerative processes in the mdx muscle fibers that are not reversed by restoring the intracellular Ca^{2+} homeostasis.

As addressed by the authors, many questions remain to be answered including the molecular link explaining how removal of muscle-specific Orai1-mediated Ca2+ influx has these beneficial effects in mdx muscle. Here the nuclear-factor κΒ (NfκΒ) and transforming growth factor β (TGF- β) signaling pathways might be of interest, given their known DMD pathomechanistic links, and that their pathway activities have been associated with Orai expression levels (Berry et al., 2018; Kang et al., 2021). Orail also plays an important role in maintaining Ca2+ homeostasis in healthy skeletal muscle, and the potential limitations of Orail-targeted pharmacological therapy for chronic diseases, such as decreased muscle fatigue resistance with Orai1 knockout as shown by the authors need to be investigated further. Also, patients with Orail deficiency display muscle hypotonia and severe combined immunodeficiency coupled with autoimmunity (McCarl et al., 2009), highlighting Orail's important role for health. Until the physiological function of Orail in skeletal muscle and other tissues is better understood, it is still too early to suggest it as a promising drug target in humans. Drug target or not, Orail is also expressed in a wide variety of tissues, which stresses that a potential future intervention needs to be tissue specific and carefully dosed to avoid detrimental and dangerous side-effects. Another DMD-related aspect that would be interesting to assess is a potential Orail involvement in the DMDinduced diaphragm and cardiac dysfunction. Although DMD

clinically manifests as progressive muscle weakness, it is the respiratory failure and heart problems (often develop in the form of dilated cardiomyopathy) that ultimately causes the premature death. The diaphragm is a skeletal muscle with a similar cytoskeleton, extracellular matrix, and excitationcontraction (EC) coupling as the limb muscles examined by García-Castañeda and colleagues, and hence one could speculate that it is likely that the removal of Orai1-mediated Ca2+ influx has beneficial effects in DMD muscle. On the other hand, there is more uncertainty whether eliminating Orail-mediated Ca2+ influx in cardiac muscle would have any effect given that cardiac muscle is designed to handle Ca2+ influx as its EC coupling, and thus every heartbeat, is initiated by Ca2+ influx via the voltage-gated Ca²⁺ channel (Cav2.1, L-type Ca²⁺ channel). Hopefully this and additional aspects of Orai physiology and pathophysiology in striated muscle will be addressed by the Dirksen laboratory or others in the near future. Nevertheless, the results of García-Castañeda et al. (2022) demonstrate an important role of enhanced Orail-mediated Ca2+ entry in exacerbating the dystrophic phenotype of mdx mice, rendering Orail a potential therapeutic target with a "fits-all approach" for the treatment of DMD. On that note, it is promising that there is a continuous development of Orail inhibitors (Azimi et al., 2020) for various indications, and some have entered clinical trials with DMD potentially included in the near future.

Acknowledgments

Eduardo Ríos served as editor.

The authors declare no competing financial interests.

J.T. Lanner is supported by the Swedish Research Council.



Author contributions: A.J. Cheng, F. von Walden and J.T. Lanner drafted the manuscript and approved the final draft of the manuscript.

References

- Azimi, I., R.J. Stevenson, X. Zhang, A. Meizoso-Huesca, P. Xin, M. Johnson, J.U. Flanagan, S.B. Chalmers, R.E. Yoast, J.S. Kapure, et al. 2020. A new selective pharmacological enhancer of the Orail Ca²⁺ channel reveals roles for Orail in smooth and skeletal muscle functions. ACS Pharmacol. Transl. Sci. 3:135–147. https://doi.org/10.1021/acsptsci.9b00081
- Bellinger, A.M., S. Reiken, C. Carlson, M. Mongillo, X. Liu, L. Rothman, S. Matecki, A. Lacampagne, and A.R. Marks. 2009. Hypernitrosylated ryanodine receptor calcium release channels are leaky in dystrophic muscle. Nat. Med. 15:325–330. https://doi.org/10.1038/nm.1916
- Berry, C.T., M.J. May, and B.D. Freedman. 2018. STIM- and Orai-mediated calcium entry controls NF-κB activity and function in lymphocytes. *Cell Calcium*. 74:131–143. https://doi.org/10.1016/j.ceca.2018.07.003
- Bladen, C.L., D. Salgado, S. Monges, M.E. Foncuberta, K. Kekou, K. Kosma, H. Dawkins, L. Lamont, A.J. Roy, T. Chamova, et al. 2015. The TREAT-NMD DMD global database: Analysis of more than 7,000 Duchenne muscular dystrophy mutations. *Hum. Mutat.* 36:395–402. https://doi.org/10.1002/humu.22758
- Edwards, J.N., O. Friedrich, T.R. Cully, F. von Wegner, R.M. Murphy, and B.S. Launikonis. 2010. Upregulation of store-operated Ca²⁺ entry in dystrophic mdx mouse muscle. *Am. J. Physiol. Cell Physiol.* 299:C42–C50. https://doi.org/10.1152/ajpcell.00524.2009
- Emrich, S.M., R.E. Yoast, and M. Trebak. 2022. Physiological functions of CRAC channels. Annu. Rev. Physiol. 84:355-379. https://doi.org/10.1146/ annurev-physiol-052521-013426

- Eser, G., and H. Topaloğlu. 2022. Current outline of exon skipping trials in Duchenne muscular dystrophy. *Genes.* 13:1241. https://doi.org/10.3390/genes13071241
- García-Castañeda, M., A. Michelucci, N. Zhao, S. Malik, and R.T. Dirksen. 2022. Postdevelopmental knockout of Orail improves muscle pathology in a mouse model of Duchenne muscular dystrophy. J. Gen. Physiol. 154: e202213081. https://doi.org/10.1085/jgp.202213081
- Hoffman, E.P., R.H. Brown Jr, and L.M. Kunkel. 1987. Dystrophin: The protein product of the Duchenne muscular dystrophy locus. *Cell.* 51:919–928. https://doi.org/10.1016/0092-8674(87)90579-4
- Kang, Q., X. Peng, X. Li, D. Hu, G. Wen, Z. Wei, and B. Yuan. 2021. Calcium channel protein ORAII mediates TGF-β induced epithelial-to-mesenchymal transition in colorectal cancer cells. Front. Oncol. 11:649476. https://doi.org/10.3389/fonc.2021.649476
- McCarl, C.A., C. Picard, S. Khalil, T. Kawasaki, J. Röther, A. Papolos, J. Kutok, C. Hivroz, F. Ledeist, K. Plogmann, et al. 2009. ORAII deficiency and lack of store-operated Ca²⁺ entry cause immunodeficiency, myopathy, and ectodermal dysplasia. J. Allergy Clin. Immunol. 124:1311–1318.e7. https://doi.org/10.1016/j.jaci.2009.10.007
- Mendell, J.R., and M. Lloyd-Puryear. 2013. Report of MDA muscle disease symposium on newborn screening for Duchenne muscular dystrophy. Muscle Nerve. 48:21–26. https://doi.org/10.1002/mus.23810
- Segal, D.J. 2022. The promise of gene editing: So close and yet so perilously far. Front. Genome Ed. 4:974798. https://doi.org/10.3389/fgeed.2022 974798
- Zhao, X., J.G. Moloughney, S. Zhang, S. Komazaki, and N. Weisleder. 2012. Orail mediates exacerbated Ca²⁺ entry in dystrophic skeletal muscle. PLoS One. 7:e49862. https://doi.org/10.1371/journal.pone.0049862
- Zweyer, M., H. Sabir, P. Dowling, S. Gargan, S. Murphy, D. Swandulla, and K. Ohlendieck. 2022. Histopathology of Duchenne muscular dystrophy in correlation with changes in proteomic biomarkers. Histol. Histopathol. 37:101-116. https://doi.org/10.14670/hh-18-403