

## RESEARCH ARTICLE

# Insights into channel modulation mechanism of RYR1 mutants using $\text{Ca}^{2+}$ imaging and molecular dynamics

Toshiko Yamazawa<sup>1,2\*</sup>, Haruo Ogawa<sup>3\*</sup>, Takashi Murayama<sup>4</sup>, Maki Yamaguchi<sup>1</sup>, Hideto Oyamada<sup>5</sup>, Junji Suzuki<sup>2,6</sup>, Nagomi Kurebayashi<sup>4</sup>, Kazunori Kanemaru<sup>2,7</sup>, Katsuji Oguchi<sup>5</sup>, Takashi Sakurai<sup>4</sup>, and Masamitsu Iino<sup>2,7</sup>

**Type 1 ryanodine receptor (RYR1) is a  $\text{Ca}^{2+}$  release channel in the sarcoplasmic reticulum in skeletal muscle and plays an important role in excitation–contraction coupling. Mutations in the *RYR1* gene cause severe muscle diseases such as malignant hyperthermia (MH), which is a disorder of CICR via RYR1. Thus far, >300 mutations in *RYR1* have been reported in patients with MH. However, owing to a lack of comprehensive analysis of the structure–function relationship of mutant RYR1, the mechanism remains largely unknown. Here, we combined functional studies and molecular dynamics (MD) simulations of RYR1 bearing disease-associated mutations at the N-terminal region. When expressed in HEK293 cells, the mutant RYR1 caused abnormalities in  $\text{Ca}^{2+}$  homeostasis. MD simulations of WT and mutant RYR1s were performed using crystal structure of the N-terminal domain (NTD) monomer, consisting of A, B, and C domains. We found that the mutations located around the interdomain region differentially affected hydrogen bonds/salt bridges. Particularly, mutations at R402, which increase the open probability of the channel, cause clockwise rotation of BC domains with respect to the A domain by alteration of the interdomain interactions. Similar results were also obtained with artificial mutations that mimic alteration of the interactions. Our results reveal the importance of interdomain interactions within the NTD in the regulation of the RYR1 channel and provide insights into the mechanism of MH caused by the mutations at the NTD.**

## Introduction

The RYR plays a key role in  $\text{Ca}^{2+}$  release from the SR during excitation–contraction coupling of skeletal muscle (Ríos and Pizarro, 1991; Iino, 1999; Hwang et al., 2012; Rebbeck et al., 2014; Ríos, 2018). Mutations in the *RYR1* gene (*RYR1*) cause severe muscle disorders (Robinson et al., 2006; Snoeck et al., 2015; Witherspoon and Meilleur, 2016; Jungbluth et al., 2018; Pancaroglu and Van Petegem, 2018), such as malignant hyperthermia (MH; OMIM accession no. 145600) and central core disease (CCD; OMIM accession no. 117000). MH is a serious complication resulting from general anesthesia through commonly used inhalational anesthetics. The typical symptoms include a rapid increase in body temperature and induction of a hypermetabolic state with skeletal muscle rigidity. CCD is a rare, nonprogressive myopathy that presents in infancy and is characterized by hypotonia and proximal muscle weakness. An important feature of CCD is its close association with susceptibility to MH. Exposure to caffeine or halothane has been shown to accelerate the CICR response in the SR of striated muscles (Endo,

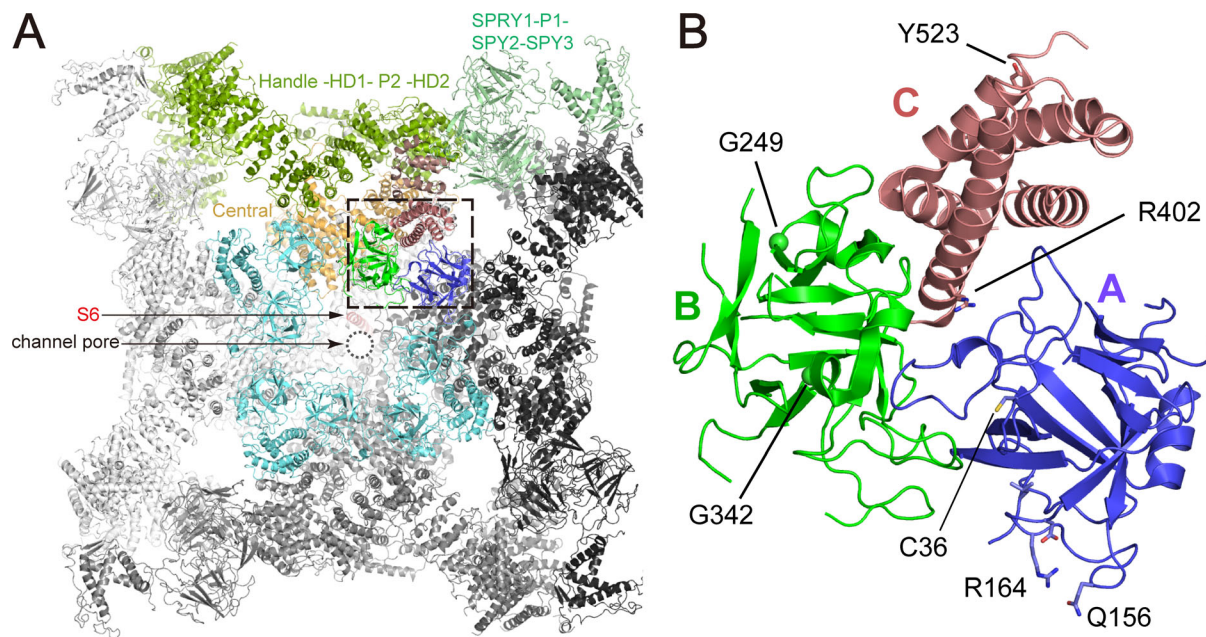
2009). It is widely believed that MH mutations cause hyperactivation of CICR, resulting in abnormal  $\text{Ca}^{2+}$  homeostasis in skeletal muscles. On the other hand, CCD mutations are of two different types: (1) gain-of-function CCD mutations found in the N-terminal (NT) and central regions (Robinson et al., 2006; Murayama et al., 2015) and (2) mutations found in the C-terminal region that result in loss of function of the channel (i.e., excitation–contraction uncoupling; Avila and Dirksen, 2001; Dirksen and Avila, 2004).

Genetic analyses of patients with MH and related diseases have revealed >300 mutations in *RYR1* (Lanner et al., 2010). There are three main mutation clusters (hotspots), which are found in the NT region (*RYR1* amino acid residues 35–614), the central region (2129–2458) in the cytoplasm, and the C-terminal region (4637–4973) near or within channel-forming segments of the *RYR1*. To understand the mechanism of the pathogenesis of MH, comprehensive study of  $\text{Ca}^{2+}$  homeostasis associated with structural properties of these mutants is necessary. However,

<sup>1</sup>Department of Molecular Physiology, The Jikei University School of Medicine, Tokyo, Japan; <sup>2</sup>Department of Pharmacology, Graduate School of Medicine, The University of Tokyo, Tokyo, Japan; <sup>3</sup>Institute for Quantitative Biosciences, The University of Tokyo, Tokyo, Japan; <sup>4</sup>Department of Pharmacology, Juntendo University School of Medicine, Tokyo, Japan; <sup>5</sup>Department of Pharmacology, School of Medicine, Showa University, Tokyo, Japan; <sup>6</sup>Department of Physiology, University of California, San Francisco, San Francisco, CA; <sup>7</sup>Division of Cellular and Molecular Pharmacology, Nihon University School of Medicine, Tokyo, Japan.

\*Toshiko Yamazawa and Haruo Ogawa contributed equally to this work; Correspondence to Toshiko Yamazawa: [toshiko1998@jikei.ac.jp](mailto:toshiko1998@jikei.ac.jp); Haruo Ogawa: [haru@iam.u-tokyo.ac.jp](mailto:haru@iam.u-tokyo.ac.jp).

© 2019 Yamazawa et al. This article is distributed under the terms of an Attribution–Noncommercial–Share Alike–No Mirror Sites license for the first six months after the publication date (see <http://www.rupress.org/terms/>). After six months it is available under a Creative Commons License (Attribution–Noncommercial–Share Alike 4.0 International license, as described at <https://creativecommons.org/licenses/by-nc-sa/4.0/>).



**Figure 1. Disease-associated mutations in the NTD RYR1.** (A) Cryo-EM structure of RYR1 tetramer viewed from the cytoplasmic side (PDB accession no. 5TB0). The crystal structure of the RYR1 NTD (PDB accession no. 2XOA; Tung et al., 2010) focused on in this study (dashed box) is superimposed on the cryo-EM structure. Each domain in the crystal structure of the NTD (dashed box) in one monomer is colored in blue (A domain), green (B domain), and red (C domain), respectively. The part corresponding to the NTD in the remaining monomers is shown in cyan. Within one monomer of the cryo-EM structure, the domains corresponding to SPRY1, P1, SPRY2, and SPRY3 are shown in light green; the domains corresponding to Handle, HD1, P2, and HD2 are shown in yellow-green; and the Central domain is shown in orange. The portion excluding the C domain in the NTDs is shown in light brown. Transmembrane helix S6 constituting the channel pore (dotted circle) is shown in red. (B) Closeup view of the NTD. The three domains are colored the same as in A. The seven mutation sites used in this study (C36, Q156, R164, G249, G342, R402, and Y523) are labeled.

due to lack of such studies, the mechanism remains largely unknown.

RYR1 exists as a huge (~2 MDa) homotetrameric channel with a large cytoplasmic structure that is involved in the regulation of the channel pore, as well as six transmembrane segments at its C-terminus, forming a cation channel pore. The crystal structure of the NT region (1–559; hereafter referred to as the N-terminal domain [NTD]) of RYR1 was first solved by Van Petegem's group (Tung et al., 2010). The NTD consists of three domains (A, B, and C) and forms a cytoplasmic vestibule (boxed area in Fig. 1 A), and it contains most of the proposed mutation hotspots underlying NT diseases (Fig. 1 B). The same group further performed structural analysis of a series of NT disease-associated mutant RYR1s and demonstrated that mutations affecting interdomain hydrogen bonds/salt bridges alter related locations of domains (Kimlicka et al., 2013a). However, the travel distance of each domain caused by the mutations was rather small (<2.5 Å), possibly due to crystal packing forces that do not occur under physiological conditions. Thus, evaluating the strength of hydrogen bonds/salt bridges between domains by MD simulation may help to improve understanding of the domain reorientations. Recently, the structures of RYR1 at near-atomic resolution were solved by single-particle cryo-EM by three groups (Efremov et al., 2015; Yan et al., 2015; Zalk et al., 2015; Willegems and Efremov, 2018). The structures revealed a complex architecture, involving a superhelical scaffold in the cytoplasmic region and a channel region of the voltage-gated ion channel superfamily. Subsequently, the RYR1 structure in an

open state was solved by many different groups (Bai et al., 2016; des Georges et al., 2016; Wei et al., 2016; Willegems and Efremov, 2018). Although three structures of RYR1 in the open state were revealed, the gating mechanism of RYR is still not fully understood. The NT region located in the cytoplasm is ~100 Å away from the cation channel pore. Therefore, it remains difficult to understand how a single disease-associated mutation at the NT region influences the opening of the channel pore.

To elucidate the molecular basis of the pathogenesis of MH, we performed functional analysis of RYR1 carrying disease-associated mutations at the NT region using a heterologous expression system in human embryonic kidney (HEK) 293 cells, which we established previously (Murayama et al., 2015). We studied eight MH-associated mutations (C36R, Q156K, R164L, G249R, G342R, R402C, R402H, and Y523C) and two MH/CCD mutations (R164C and Y523S) in the NT hotspot by live-cell  $\text{Ca}^{2+}$  imaging. Furthermore, multiple sets of MD simulation were performed to investigate the structure of the MH/CCD mutants. Since the quality of the initial models is important to obtain reliable predictions by MD simulation (Law et al., 2005), the atomic structure of the RYR1 NTD (1–559) solved by x-ray crystallography was used as the template molecule for all the simulations. Live-cell  $\text{Ca}^{2+}$  imaging in RYR1 mutants revealed abnormalities in  $\text{Ca}^{2+}$  homeostasis in terms of caffeine sensitivity, ER  $\text{Ca}^{2+}$  content, and resting cytoplasmic  $\text{Ca}^{2+}$  concentration ( $[\text{Ca}^{2+}]_{\text{cyt}}$ ). Statistical analysis of MD simulation revealed that changes in  $\text{Ca}^{2+}$  homeostasis were correlated with changes in the specific hydrogen bonds/salt bridges. These results will

clarify the functional significance of the NT region of RYR1. They might also provide clues to why an increase in the channel pore open probability occurs in mutants. Ultimately, this study may lead to the development of new therapeutic drugs for MH/CCD patients.

## Materials and methods

### Generation of stably inducible HEK293 cell lines

Disease-associated mutation in the NT region corresponding to Q156K was introduced into the rabbit RYR1 complementary DNA (cDNA) by inverse PCR. Inverse PCR was performed using a HindIII-SalI fragment (pBS-RYR1cs1) from cDNA cassettes encoding full-length rabbit skeletal muscle RYR1 (pBS-RYR1; Tong et al., 1997, 1999; Nakano et al., 2014). Other disease-associated mutations in the NT region used in this study (C36R, R164C, R164L, G249R, G342R, R402C, R402H, Y523C, and Y523S) have been previously reported (Murayama et al., 2015). In addition, as mutants for subsequent verification, E40A and D61A were also prepared by the same method as above. Stable and inducible RYR1 mutations in HEK293 cells were generated as previously described (Kakizawa et al., 2012; Nakano et al., 2014; Murayama et al., 2015) using the FLP-In T-REx system (Invitrogen). Clones with suitable RYR1 expression were selected and used in subsequent experiments.

### Time-lapse $\text{Ca}^{2+}$ imaging

$\text{Ca}^{2+}$  measurements were performed using previously described procedures (Kakizawa et al., 2012; Murayama et al., 2015, 2016). Briefly, HEK293 cells expressing WT or mutant RYR1 were cultured on glass-bottomed dishes (MatTek). 24–30 h after the addition of doxycycline at the final concentration of 2  $\mu\text{g}/\text{ml}$ , cells were loaded with 4  $\mu\text{M}$  fura-2 acetoxymethyl ester in physiological salt solution containing 150 mM NaCl, 4 mM KCl, 2 mM  $\text{CaCl}_2$ , 1 mM  $\text{MgCl}_2$ , 5.6 mM glucose, and 5 mM HEPES at pH 7.4. All experiments were performed at room temperature (23–25°C).

Fluorescence images were acquired at 420 nm using an inverted microscope (IX70; Olympus, Japan) equipped with a 40 $\times$  objective (numerical aperture, 0.95; UPlanSApo/340; Olympus), together with a cooled charge-coupled device camera (Rolera XR; Teledyne QImaging), at a rate of one frame every 1 or 2 s. Excitation wavelengths were 345 nm and 380 nm. Image analyses were performed using IPLab software (BD Biosciences Bioimaging). Regions of interest corresponding to individual cells were selected, and the mean fluorescence intensity of each region of interest minus the background intensity was calculated for each frame. We used the F345/F380 ratio (the mean fluorescence intensity at an excitation wavelength of 345 nm divided by that at an excitation wavelength of 380 nm) to estimate  $[\text{Ca}^{2+}]_{\text{cyt}}$ , as described elsewhere (Kakizawa et al., 2012; Murayama et al., 2015). The  $K_d$  (239 nM) for  $\text{Ca}^{2+}$  was determined via in vitro calibration of fura-2 fluorescence (Grynkiewicz et al., 1985).

### MD simulation of RYR1

The atomic coordinates of the NTD (1–559) of rabbit RYR1 (PDB accession no. 2XOA; Tung et al., 2010) were used as a template

for all calculations. Modeling of missing loops in 2XOA (85–96, 126–129, 185–188, 226–228, 324–327, 362–370, 427–435, 456–464, and 500–505) was performed by using the MODELLER program (version 9.12; Fiser et al., 2000; Martí-Renom et al., 2000; Webb and Sali, 2014). Construction of the initial mutant structures (C36R, Q156K, R164C, R164L, G249R, G342R, R402C, R402H, Y523C, Y523S, E40A, and D61A) and additions of hydrogen bonds/salt bridges and other missing atoms were performed using the LEaP module in the AMBER package (version 14). The AMBER package (version 14) with the parm99 force field (Case et al., 2005) was used for MD simulation.

The initial structure of the NTD was immersed in a rectangular box filled with ~11,500 TIP3P water molecules (Jorgensen et al., 1983). To ensure a physiological ionic concentration of 0.15 M and zero net charge,  $\text{K}^+$  and  $\text{Cl}^-$  ions were added adequately. The time step was 1 fs, and periodic boundary conditions were applied throughout the simulation. The atomic coordinates of the systems were saved for later analysis every 0.5 ps during MD simulations. A 10-Å cutoff distance was used for nonbonded interactions. The SHAKE algorithm (Ryckaert and Ciccotti, 1977) was applied to constrain the bond lengths of hydrogen-containing bonds. After energy minimization, the system was gradually heated in 50–60 steps from 100 K to 310 K under constant-volume conditions. Trajectories 50 ns long were obtained for each mutant and WT under a constant pressure (1 atm) and temperature (310 K) after the equilibration of the system. Twenty such trajectories were obtained for each of the initial structures of the WT and the mutants, applying different sets of initial velocities at the start of the simulation run to sample different sets of the dynamic structure of the molecule.

To evaluate the stability of the hydrogen bonds/salt bridges between interdomains during the MD simulation, we calculated and analyzed a value of the probability of forming the hydrogen bonds/salt bridges using the “ptraj” program in the AMBER package. The initial 20 ns of data were discarded as preequilibration. The VMD program (Humphrey et al., 1996) was used for analysis and visualizations.

### MD simulation using tetramer

As the initial model of the tetramer of NTDs, the cryo-EM structure of RYR1 (PDB accession no. 5GKZ; residues 1–532; Bai et al., 2016) was used. Modeling of missing loops in 5GKZ (81–99, 116–145, 182–192, 223–230, 235–242, 264–270, 320–335, 360–379, 414–437, 450–473, and 490–514) was performed by using the MODELLER program (version 9.12; Martí-Renom et al., 2000; Webb and Sali, 2014). Construction of the initial mutant structures (R402C and R402H) and additions of hydrogen bonds/salt bridges and other missing atoms were performed using the LEaP module in the AMBER package (version 14). The AMBER package (version 14) with the 14SB force field (Case et al., 2005) was used for MD simulations.

The initial structure of the NTD was immersed in a rectangular box filled with ~95,000 TIP3P (transferable intermolecular potential with 3 points) water molecules (Jorgensen et al., 1983). To ensure a physiological ionic concentration of 0.15 M and zero net charge,  $\text{K}^+$  and  $\text{Cl}^-$  ions were adequately added. The



time step was 2 fs, and periodic boundary conditions were applied throughout the simulation. The atomic coordinates of the systems were saved for later analysis every 10 ps during MD simulations. A 10-Å cutoff distance was used for nonbonded interactions. The WT and mutant systems were refined with energy minimization using the steepest descent method: A 5,000-step energy minimization was performed with harmonic constraints (force constant, 10 kcal/mol/Å<sup>2</sup>) applied to all protein atoms.

The systems were then heated to 310 K in 500 ps by MD with harmonic constraints (force constant, 1 kcal/mol/Å<sup>2</sup>) applied to  $\alpha$ -carbon (Ca) atoms only, after which the systems were equilibrated for 1 ns by MD in the number of particles, system volume, absolute temperature ensemble with the same constraints as in heating. Finally, the systems were subjected to a 60-ns production MD run in the number of particles, constant pressure, constant temperature ensemble. The SHAKE algorithm was used to constrain the bond lengths of hydrogen-containing bonds, which allows a time step of 2 fs for MD simulations. For each WT and R402C/H mutant system, three 60-ns MD trajectories were generated and combined for hydrogen bond/salt bridge analyses (after discarding the initial 30 ns for system equilibration).

### Analysis of structures

All structural figures were prepared with PyMOL (PyMOL Molecular Graphics System; Schrödinger). The Coot program was used for analysis of structures (Emsley and Cowtan, 2004). ProFit (Martin and Porter, <http://www.bioinf.org.uk/software/profit/>; McLachlan, 1982) was used for fitting of structures. As the cryo-EM structures, we used 5TBO as the closed state (in the presence of EGTA) and 5T9V as the open state (in the presence of caffeine, Ca<sup>2+</sup>, and ATP) of the RYR1 since both structures are the highest resolution in the structures solved by three groups. To create a hypothetical homotetramer of the NTD, monomer structures before or after the MD simulations are fitted to the cryo-EM structure of RYR1 in the closed state (PDB accession no. 5TBO) using the A domain of the NTD. The HBPLUS program (McDonald and Thornton, 1994) was used to identify hydrogen bonds/salt bridges in the atomic coordinates. DynDom (Hayward and Berendsen, 1998) was used to find domain movements.

### Statistics

All statistical analyses of the data were performed using Kaleidagraph software for Windows (Synergy Software). Differences between two groups were analyzed with Student's *t* test. *P* values <0.05 were considered to represent statistical significance. One-way ANOVA with Tukey's honestly significant difference post hoc test was used when comparing data from three groups.

### Online supplemental material

Fig. S1 shows the results of Western blot analysis of mutant RYR1 channels in HEK293 cells. Fig. S2 shows Ca<sup>2+</sup> homeostasis in cells expressing other mutant RYR1s. Figs. S3 and S4 show the results of the MD simulations for other mutants. Fig. S5 shows a

comparison of the strength (tightness) of the interdomain interactions in the NTD of the WT and mutant RYR1s (C36R, G249R, R402C, and R402H). Fig. S6 shows the results of the MD simulations using the NTD tetramer of the WT. Fig. S7 shows the results of the MD simulations using the NTD tetramer of the R402 mutants. Fig. S8 shows the x-ray structure of the RYRs and inositol-1,4,5-triphosphate receptor (IP<sub>3</sub>R) in the NTD. Table S1 shows the disease-associated mutations used in this study. Table S2 shows the disease-associated mutations located within the domains using a database ([https://www.cardiodb.org/paralogue\\_annotation/](https://www.cardiodb.org/paralogue_annotation/)).

## Results

### Expression of RYR1 mutants

Ten mutant RYR1 channels with mutations at seven different positions in the NT region were prepared (Fig. 1 B; and Tables S1 and S2). All the mutations have been associated with MH susceptibility, and two of them (R164C and Y523S) have also been associated with CCD phenotypes (i.e., MH/CCD; see Table S1). These mutant RYR1 channels were heterologously expressed in HEK293 cells, which do not express endogenous RYRs (Murayama et al., 2015, 2016). RYR1 expression was induced by the addition of doxycycline (2 µg/ml) and confirmed by Western blotting (Fig. S1 A).

### Functional analysis of RYR1 mutants

We first characterized the caffeine-induced Ca<sup>2+</sup> transient as an indicator of CICR activity. As the control, HEK293 cells without transfection of the RYR1 gene were used. No substantial Ca<sup>2+</sup> increase was observed upon addition of 3 mM of caffeine (Fig. S2 A). This result is consistent with our previous results (Kakizawa et al., 2012). As shown in Fig. 2 A and Fig. S2 B, these mutants showed varied responses to caffeine as compared with the WT. Since small but significant Ca<sup>2+</sup> transients were detected at 0.3 mM caffeine in WT and the responses plateaued at 10 mM caffeine in all cases (Fig. S2 C), peaks of the Ca<sup>2+</sup> transient at 0.3 and 10 mM caffeine were compared in Fig. 2, B and C. Five of the mutants studied (Q156K, R164C, R164L, R402C, and R402H) exhibited significant increases in response to 0.3 mM caffeine, indicating marked enhancement of caffeine sensitivity, which can be further appreciated by the leftward shift of dose-response relationships (Fig. 2 E and Fig. S2 C). On the other hand, the peak response at 10 mM caffeine, which represents ER Ca<sup>2+</sup> content, decreased in all 10 mutants (Fig. 2 C). In particular, the mutant Y523S, which is associated with MH/CCD phenotype, exhibited a Ca<sup>2+</sup> transient only ~10% of that in WT, despite the expression level being equivalent to that of WT (Fig. S1). While resting [Ca<sup>2+</sup>]<sub>cyt</sub> was ~40 nM in cells expressing WT RYR1, most of the disease-associated mutants caused significantly higher levels of resting [Ca<sup>2+</sup>]<sub>cyt</sub>, with values ranging between 50 and 90 nM (Fig. 2 D). Interestingly, there was an inverse correlation between resting [Ca<sup>2+</sup>]<sub>cyt</sub> and peak Ca<sup>2+</sup> transient induced by 10 mM caffeine (*R*<sup>2</sup> = 0.68), suggesting that an increase in [Ca<sup>2+</sup>]<sub>cyt</sub> was attributable mainly to leaks from the ER, in line with the results of our previous study (Murayama et al., 2015; Fig. 2 F).

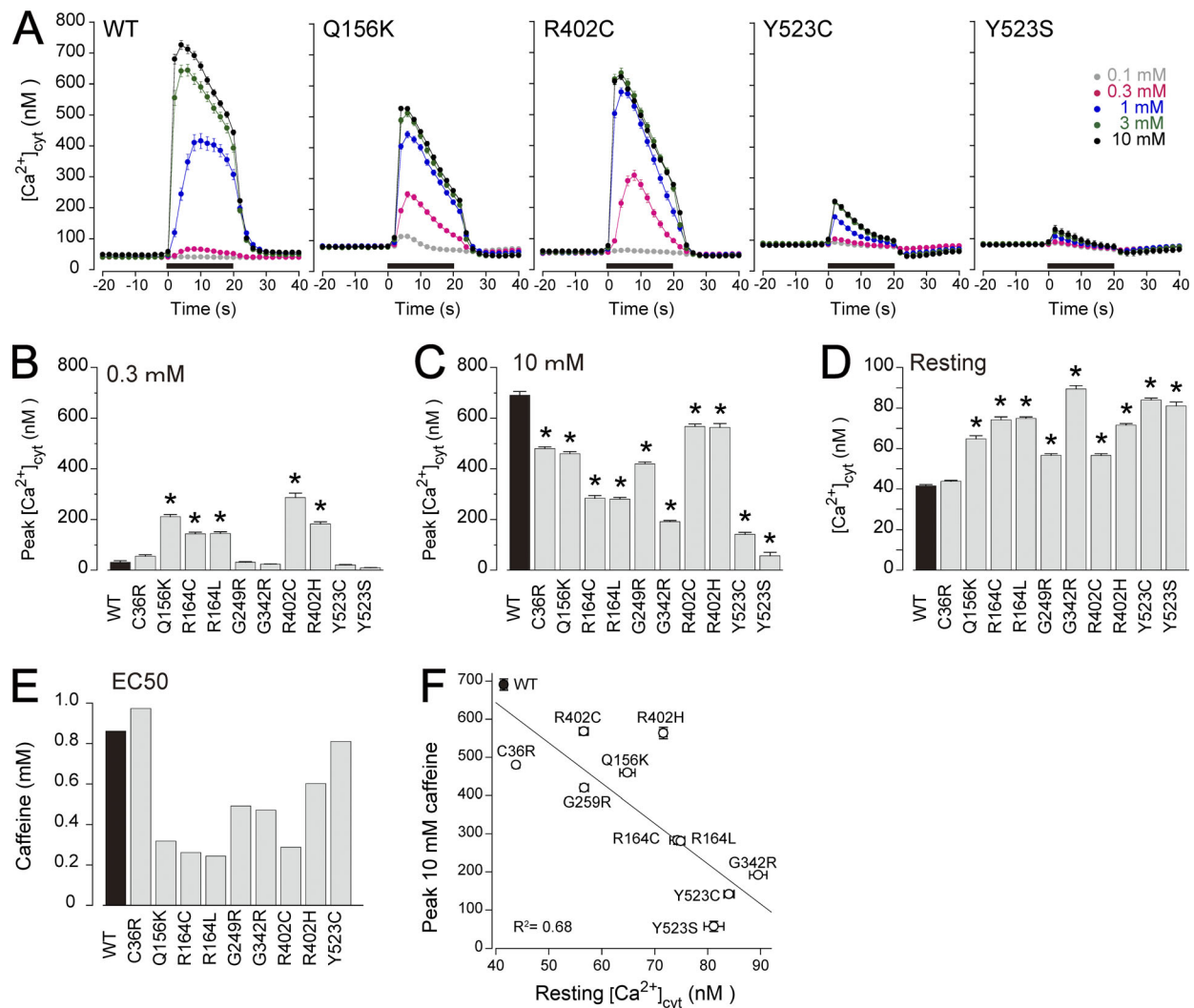


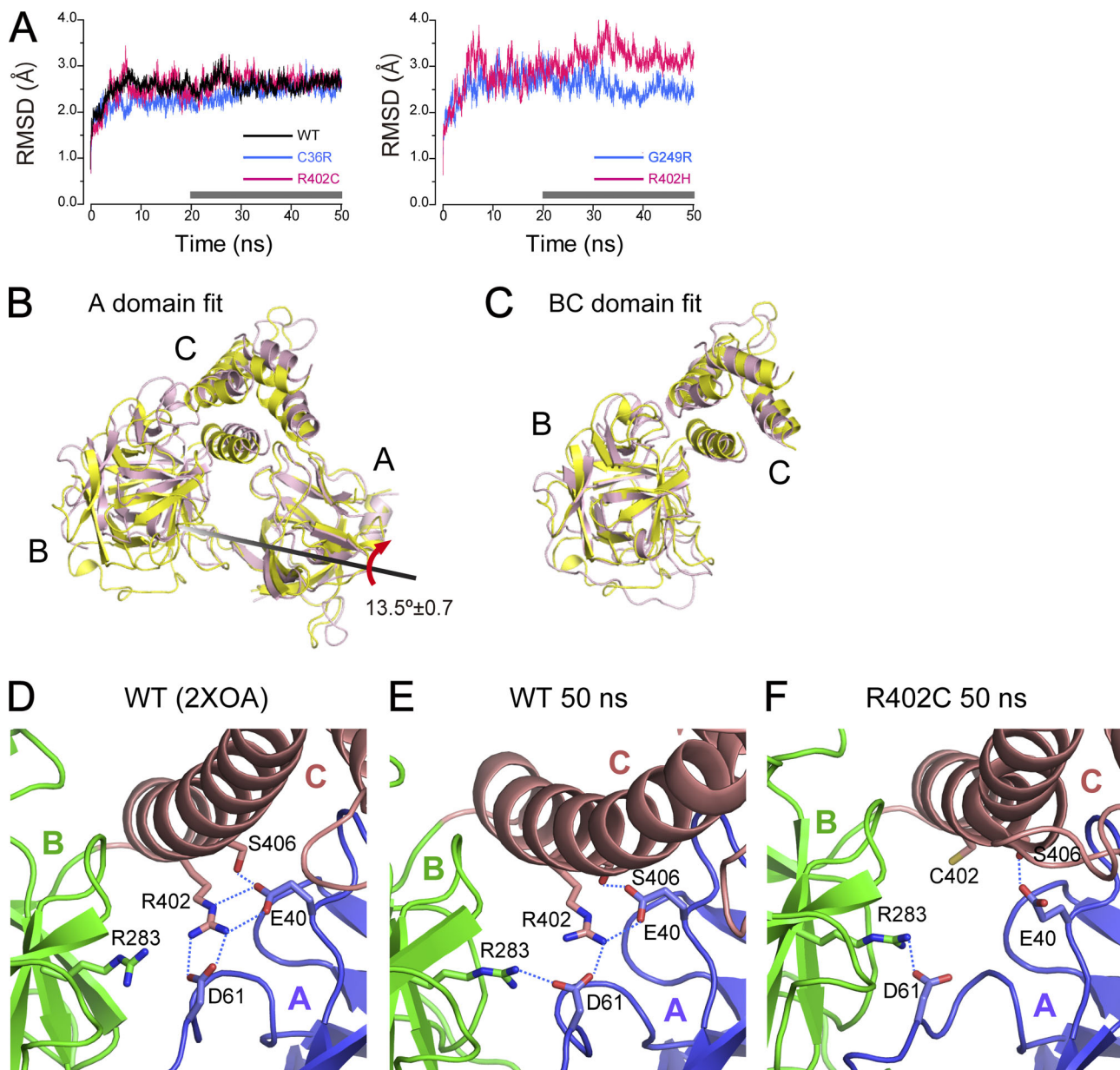
Figure 2.  $Ca^{2+}$  homeostasis in cells expressing WT or mutant RYR1s in the N-terminal region. (A) Caffeine-induced changes in  $[Ca^{2+}]_{cyt}$  determined by fura-2 in WT and mutants (Q156K, R402C, Y523C, and Y523S). Caffeine was applied at the time points indicated by the black horizontal bar. (B and C) Comparison of the peaks of  $[Ca^{2+}]_{cyt}$  transiently induced by 0.3 mM (B;  $n = 31$ –322) and 10 mM (C;  $n = 68$ –318) caffeine in the WT (black) or the mutants (gray). (D) Comparison of resting  $[Ca^{2+}]_{cyt}$  in the WT (black) and mutants (gray;  $n = 148$ –651). (E) Half-maximal effective concentration ( $EC_{50}$ ) for caffeine in the WT (black) and the mutants (gray). Y523S was excluded because caffeine's maximum and minimum response magnitudes are too small for accurate calculations. (F) The relationship between the maximum caffeine-induced  $Ca^{2+}$  transient and resting  $[Ca^{2+}]_{cyt}$  in cells expressing WT (black) or mutant (white) RYR1 ( $R^2 = 0.68$ ). Values are shown as mean  $\pm$  SEM ( $n = 63$ –651). \* $P < 0.0001$  by  $t$  test compared with the WT.

### MD simulation of the NTD of the WT and disease-associated mutants

To address the structural changes caused by disease-associated mutations, MD simulations using NTD monomers of the WT and 10 mutants were performed. Although the resulting movements may be amplified, owing to the lack of adjacent protomers that would be present in the tetramer, they may indicate some structural changes occurring because of the disease-associated mutation. After modeling the missing loops in 2XOA (the atomic coordinates of the NTD of rabbit RYR1), construction of the initial 10 mutant structures, and addition of protons to the initial structures, multiple MD simulations were performed using AMBER (version 14) with the parm99 force field (Case et al., 2005) under physiological conditions. We observed that the molecule remains stable during the simulation, because the

RMSD value of Ca atoms throughout the MD simulation was  $<3 \text{ \AA}$  (Fig. 3 A and Fig. S3 A).

We next analyzed the structural changes after the MD simulations. First, we compared the MD structures of R402C/H mutants with the structure of the WT after 50-ns simulation (R402C, Fig. 3, A–C; R402H, Fig. 3 A and Fig. S3, B–D). DynDom was used to analyze the domain movements in the R402C and R402H mutants after simulation. Fig. 3 B and Fig. S3 B display representative results from 20 calculations. Surprisingly, the B and C domains together rotated  $13.5 \pm 0.7$  degrees ( $n = 20$ ) clockwise with respect to the A domain in R402C (Fig. 3 B and Fig. S3 E) or  $14.3 \pm 1.2$  degrees ( $n = 20$ ) in the R402H mutant (Fig. S3, B and E). The travel distances of the largest moving parts for the R402C and R402H were  $\sim 8 \text{ \AA}$  and  $\sim 9.5 \text{ \AA}$ , respectively. Indeed, these distances were much



**Figure 3. Analysis of the movements in the R402C mutant after the MD simulation.** (A) Structural deviations observed in the MD simulations. Plots of the RMSD of the C $\alpha$  atoms from the initial structures of NTD RYR1s (WT, C36R; AB domain mutation, G249R; buried and R402C/H; AC domain mutation) as a function of time are shown. Calculations of each RMSD were performed using MD trajectories from the representative result. The trajectory during 20–50 ns (gray horizontal bars) was analyzed for hydrogen bonds/salt bridges (see Fig. 4, B–F). (B) Superimposition of the monomer of the WT (yellow) and R402C mutant (pink) after 50 ns of the MD simulation. The result shown here is a representative of 20 calculations. The BC domains rotated 14.3 degrees with respect to the A domain in the R402C mutant. The average rotation angle is shown in Fig. S3 E. The rotation axis is indicated by a black line. (C) Superimposition of the BC domains of the WT and R402 mutants after the simulation are shown with either structure almost overlapped. For simplicity, the upper part of the C domain is deleted. (D–F) Closeup view around residue 402. Colors of domains are the same as shown in Fig. 1. Dashed lines represent hydrogen bonds/salt bridges. (D) Crystal structure of the WT NTD (PDB accession no. 2XOA). R402 in the C domain forms hydrogen bond(s) with D61 in the A domain. Thus, through R402, the C domain has a connection with the A and B domains. (E) The same view after 50 ns of MD simulation of the WT NTD. The hydrogen bonds/salt bridges via R402 are tight and are maintained even after 50 ns of the simulation. D61 forms a hydrogen bond with R283 in the B domain (blue dotted line) after 50-ns simulation. (F) The same view after 50 ns of the MD simulation of the R402C mutant NTD. C402 cannot form a hydrogen bond with D61.

larger than the distance observed in the crystal structure of the mutants (Kimlicka et al., 2013a). In fact, the B and C domains of the R402C and R402H mutants nearly overlapped with those of the WT after 50-ns simulation (Fig. 3 C and Fig. S3 C). Therefore, R402 must have an important role in keeping a tight connection between the A and BC domains.

R402, which is located in the C domain, forms hydrogen bonds/salt bridges with D61 and E40 located in the A domain (Fig. 3 D). However, in the case of R402C/H mutants, NTD loses these connections (Fig. 3 F and Fig. S3 D), and this may initiate the rotation of the BC domains with respect to the A domain.



In contrast to the R402C/H mutants, there is no appreciable change in the MD simulations of C36R located at the AB domain interface and G249R buried in the B domain compared with the WT structure after 50-ns simulation (Fig. S4).

The residues for six other mutations (Q156K, R164C, R164L, G342R, Y523C, and Y523S) are located outside the NTD and are not involved in the interactions between domains but are involved in binding to the adjacent NTD unit. Calculations for these mutants were also performed (Fig. S4). However, there is no appreciable change compared with the WT structure after 50-ns simulation (Fig. S4). Since we used a monomer of the NTD in our MD simulation, it seems reasonable that the effect of the mutations located at the surface of NTD was not detected.

### Hydrogen bond/salt bridge analysis of the WT and mutants

One of the most frequently used approaches for demonstrating formations and movements of interfaces in the molecule during MD simulation is evaluation of the interactions between the side chains in the interdomain. The interactions are generally contributed by hydrogen bonds/salt bridges and hydrophobic interactions. Since the mutations we selected are all hydrophilic residues, we focused on analyzing the interaction of hydrogen bonds/salt bridges. Fig. 4 A shows the hydrogen bonds/salt bridges between domains in the crystal structure of the NTD (2XOA) analyzed by HBPLUS. Eight hydrogen bonds/salt bridges (R242-E481, D61-R402, R45-D447, R221-E397, D18-S207, E40-S406, T286-E481, and E40-R402) were found. Since the MD simulation of R402C/H mutants suggested that the interdomain hydrogen bonds/salt bridges may have an important role in exhibiting their structural changes, we decided to analyze all the interdomain hydrogen bonds/salt bridges during MD simulation. To do so, we defined the probability of forming hydrogen bonds/salt bridges. If the specific hydrogen bond/salt bridge is continuously maintained during the simulation, the value is defined as 100%. Since these hydrogen bonds/salt bridges are repeatedly formed and disrupted during the simulation, it is quite natural that the actual value becomes <100%. Therefore, the magnitude of this value corresponds to the stability of the hydrogen bond/salt bridge. For the analysis, 20 independent runs of MD simulation were performed, and the average values were plotted in Fig. 4, B–F. Interestingly, in addition to the eight hydrogen bonds/salt bridges that were found by HBPLUS, we found another important interaction (D61-R283; Fig. 3 E and Fig. 4 B).

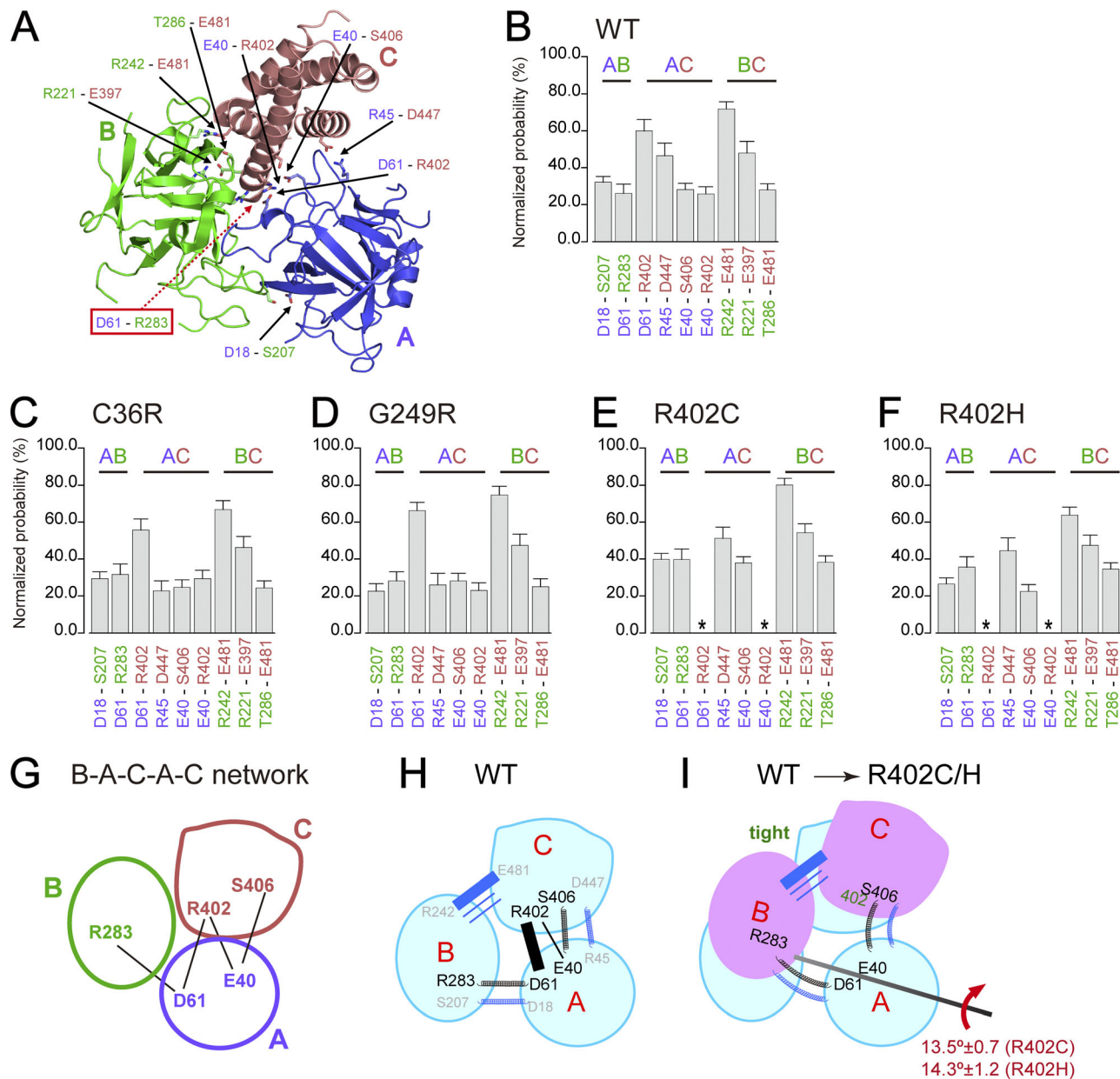
As one of the indexes indicating the tightness between domains, the standard number of hydrogen bonds/salt bridges between domains during the simulation was counted. In practice, the probability of the hydrogen bonds/salt bridges between the pair of residues constituting the interaction is considered as the number of hydrogen bonds/salt bridges between each domain. Then, the total number of hydrogen bonds/salt bridges between each domain was calculated. As shown in Fig. S5 A, the standard numbers of hydrogen bonds/salt bridges between the A and C domains and between the B and C domains are 1.7 and 1.5, respectively. However, the standard number of hydrogen bonds/salt bridges between the A and B domains is 0.7 (Fig. S5 A). Therefore, the interactions between the A and C domains and

between the B and C domains are considered to be tighter than between the A and B domains. The behavior and the total number of hydrogen bonds/salt bridges between domains in the C36R and G249R mutants are essentially the same as those in the WT (Fig. S5 B). Thus, it is reasonable to assume that the peak values of the  $\text{Ca}^{2+}$  transient of these two mutants are mild (Fig. 2). In contrast, the R402C/H mutants showed a loss of two important hydrogen bonds/salt bridges between the A and C domains (D61-R402 and E40-R402). Indeed, the standard number of hydrogen bonds/salt bridges between the A and C domains and between the A and B domains decreased drastically (0.8 and 0.8 in the R402C mutant, respectively, and 0.7 and 0.8 in the R402H mutant, respectively; Fig. S5 B). Meanwhile, the standard numbers of hydrogen bonds/salt bridges between the B and C domains in the mutants were almost the same as those of the WT (1.7 and 1.6 in the R402C and R402H mutant, respectively; Fig. S5 B). Thus, the interaction between the B and C domains is considerably tighter than interactions between the A and C domains and between the A and B domains in mutants.

R402, located in the C domain, seems to play an important role in connecting the A and B domains (Fig. 4 G). R402 forms hydrogen bonds/salt bridges with two residues (E40 and D61) in the A domain (Fig. 3 E). D61, in turn, forms a hydrogen bond/salt bridge with R283 located in the B domain, and E40 also forms a hydrogen bond/salt bridge with S406 located in the C domain. We named this B(R283)–A(D61)–C(R402)–A(E40)–C(S406) network around R402 as the B–A–C–A–C network (Fig. 3, D and E; and Fig. 4 G) because this network is critical for the connection of A, B, and C domains.

We considered why the B and C domains rotate with respect to the A domain in the R402 mutants. Fig. 4, H and I, presents schematic diagrams of the movement of the NTD in the WT and R402C/H mutants. There are four hydrogen bonds/salt bridges between the A and C domains in the WT NTD (Fig. 4 H). Among these, the D61–R402 interaction is stronger than the other three (Fig. 4 B). There are three hydrogen bonds/salt bridges between the B and C domains in the WT NTD (Fig. 4 H). Among these, the R242–E481 interaction is stronger than the other two (Fig. 4 B). Meanwhile, there are only two hydrogen bonds/salt bridges between the A and B domains, and both interactions are relatively weak (Fig. 4 B). Therefore, AC and BC domain interactions are considered to be stronger than the AB domain interaction.

In the R402C/H mutants, however, hydrogen bonds/salt bridges between the A and C domains would become drastically weak because two of the four important interactions (D61–R402 and E40–R402) were lost (Fig. 4, B, E, F, and I). The interaction between the A and B domains was weaker than other interactions in the WT, and this property was shared by the R402C/H mutants (Fig. 4, B, E, F, and I). On the other hand, interactions between the B and C domains were conserved in the R402C/H mutants, as in the WT (Fig. 4, B, E, F, and I), such that the B and C domains tend to move together (Fig. 4 I). This may cause side chain conformational changes in E40 (Fig. 3, E and F), R45, D61 (Fig. 3, E and F), and D18 in the A domain. As a result, it would become possible to bend both AC and AB domain interactions. Thus, the B and C domains would make a rotation-like movement with respect to the A domain (Fig. 4 I).



**Figure 4. Analysis of hydrogen bonds/salt bridges at the domain interfaces.** (A) Details of the hydrogen bonds/salt bridges at the domain interface detected by HBPLUS. D61–R283 interaction (red box) could not be detected by HBPLUS, but it was found by the hydrogen bond/salt bridge analysis after the MD simulation shown in B. (B–F) Analysis of probabilities of forming hydrogen bonds/salt bridges during the MD simulations in the selected mutants. Hydrogen bonds/salt bridges between AC and BC domains are considerably strong. However, hydrogen bonds/salt bridges between AB domains are relatively weak. Compared with the R402C/H mutants, the distribution of hydrogen bond/salt bridge probabilities of C36R and G249R was similar to that of WT. (G) A B(E283)–A(D61)–C(R402)–A(E40)–C(S406) network (B–A–C–A–C network) formed around R402. R402 is thought to play an important role in the connection of A, B, and C domains. R402 plays a key role in the B–A–C–A–C network. (H) The NTD in the WT after 50 ns of the MD simulation. Thick and thin lines indicate strong and weak hydrogen bonds/salt bridges, respectively. Springs indicate the hydrogen bonds/salt bridges playing important roles in the rotation of the BC domains in the R402C/H mutants, but they are weak. The interactions belonging to the B–A–C–A–C network are shown in black, and others are shown in blue. The interactions between AC domains and BC domains are considerably strong; meanwhile, the interaction between the AB domains is weak. R402 plays an important role to maintain a tight connection between the A domain and the C domain through the B–A–C–A–C network. (I) Superimposition of the R402C/H mutant (violet) on the NTD in the WT (light blue). The hydrogen bonds/salt bridges between the AB domains and BC domains are maintained similar to that in the WT. However, the interaction between the AC domains becomes drastically weak because two of four important interactions are completely lost. Two interactions (E40–S406 and R45–D447) bend, and the BC domains move together. Although the B domain moves in accordance with the movement of the C domain, two interactions (D61–R283 and D18–S207) between the AB domains bend. Thus, the BC domains together rotate clockwise by  $13.5 \pm 0.7$  degrees (R402C) or  $14.3 \pm 1.2$  degrees (R402H) with respect to the A domain. The rotation axis is indicated by a black line. Values are shown as mean  $\pm$  SEM. \* $P < 0.0001$  by Student's *t* test compared with the WT.



### Verification of the significance of the B-A-C-A-C network

The links identified as critical for establishing the B-A-C-A-C network were verified experimentally and in MD simulations as follows. We prepared two alanine-substituted mutants in the B-A-C-A-C network (E40A and D61A). Because E40 has tight interactions between R402 and S406, and because D61 has tight interactions between R283 and R402, the two mutants (E40A and D61A) are expected to have the same effect as observed in the examination of R402C/H. Two mutant RYR1 channels were heterologously expressed in HEK293 cells in the same way as described above, and expression was induced by the addition of doxycycline (2  $\mu$ g/ml) and confirmed by Western blotting (Fig. S1 B). Then, measurements of caffeine-induced  $\text{Ca}^{2+}$  transients of the mutants were performed (Fig. 5, A–D).

As shown in Fig. 5 A, two mutants showed varied responses to caffeine as compared with the WT. As in the analysis of Fig. 2, peaks of the  $\text{Ca}^{2+}$  transient at 0.3 mM and 10 mM caffeine were compared in Fig. 5, B and C. Both mutants (E40A and D61A) exhibited significant increase in response to 0.3 mM caffeine, indicating marked enhancement of caffeine sensitivity (Fig. 5 B). On the other hand, the peak response at 10 mM caffeine decreased in both mutants (Fig. 5 C). Both mutants caused significantly higher levels of resting  $[\text{Ca}^{2+}]_{\text{cyt}}$  (58–72 nM) than the resting  $[\text{Ca}^{2+}]_{\text{cyt}}$  (40 nM; Fig. 5 D). Indeed, these series of results were consistent with the results seen with R402C/H.

We next performed MD simulations (50 ns) of the two mutants (Fig. 5, E–K). The molecules remained stable during the simulation, as the RMSD value of  $\text{Ca}$  atoms throughout the MD simulation was  $<3$  Å (Fig. 5 E). DynDom was used to analyze the domain movements in the two mutants after simulation. Fig. 5, H and I, displays representative results from 20 calculations. When we analyzed the movement of the B and C domains with respect to the A domain, the BC domains in D61A and E40A rotated  $13.5 \pm 1.1$  degrees ( $n = 20$ ) and  $8.0 \pm 0.9$  degrees ( $n = 20$ ) clockwise, respectively (Fig. 5, H and I; and Fig. S3 E). Indeed, these rotations of the BC domains were consistent with that observed in the R402C/H mutants. Interestingly, each rotation angle that was observed in the four mutants (D61A, E40A, R402C, and R402H) correlated with the measurement results of resting  $[\text{Ca}^{2+}]_{\text{cyt}}$  in each mutant (Fig. 5 D). To address the molecular mechanism of these differences, the boundaries of A, B, and C domains of the three mutants were analyzed (Fig. 5, F, G, J, and K). In D61A, the interaction among A, B, and C domains via R402 (R402–D61 and R402–E41) no longer existed, due to the lack of D61, although two interactions between A and C (R402–E40 and E40–S406) remained intact. Indeed, this circumstance was almost the same as that observed in the R402C/H mutants. In E40A, as two interactions between A and C domains (R402–E40 and E40–S406) were lost, the interactions between A and C domains became weaker. However, the interactions among A, B, and C domains via R402 (R402–D61 and R402–E41) were retained. Therefore, it was considered that the rotation angle would become rather smaller in the E40A mutant compared with the other three mutants (D61A, R402C, and R402H; Fig. S3 E).

### Discussion

The NT region of the RYR1 located at the cytoplasm is  $\sim 100$  Å away from the channel pore. Nevertheless, the region contains a cluster of point mutations that cause MH and CCD. How does a single mutation at the NT region have a long-range effect on the opening of the channel pore? To clarify this question, a combination of functional studies and MD simulations of RYR1 channels carrying disease-associated mutations at the NT region were performed in this study.

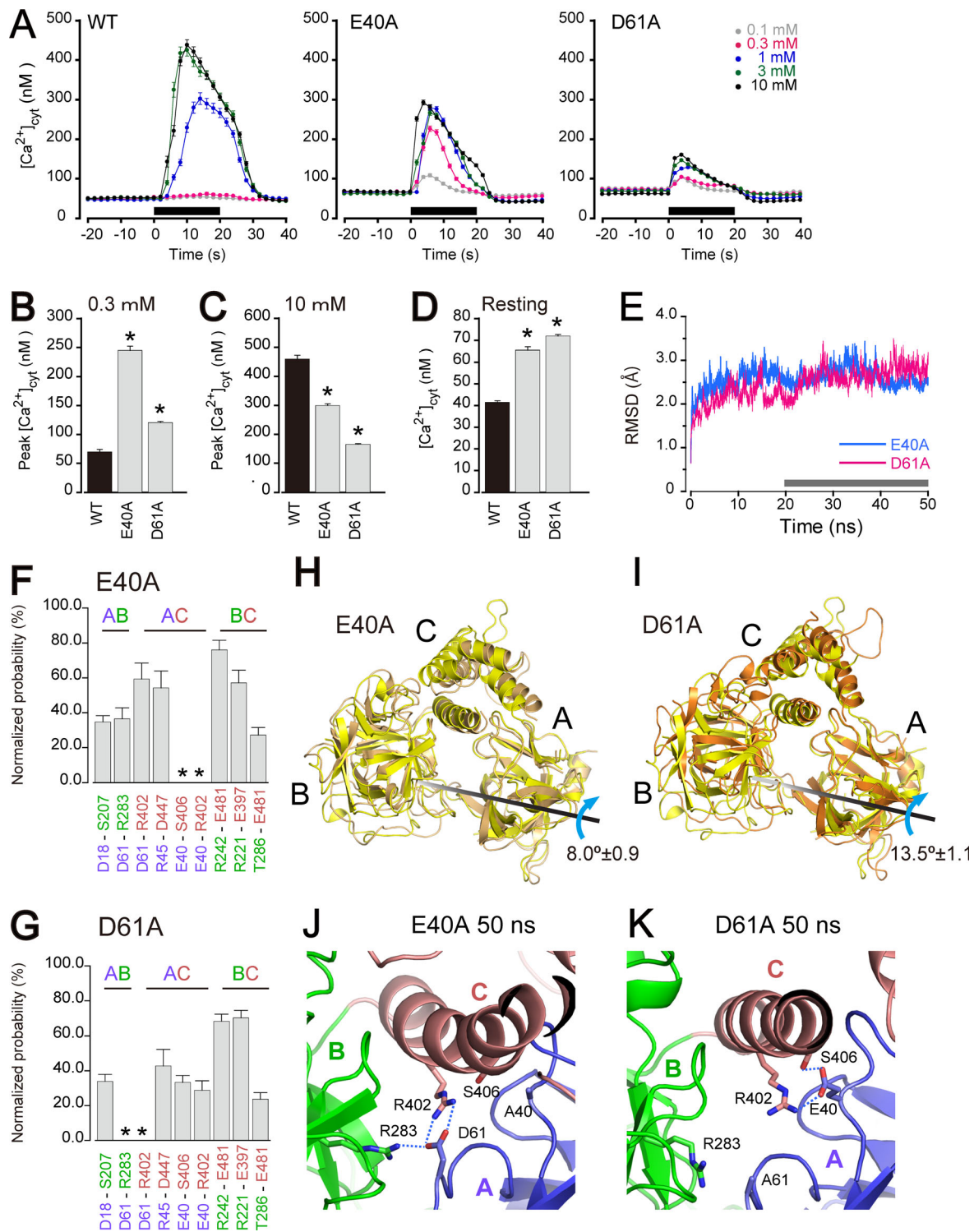
#### Importance of the B-A-C-A-C network in other RYRs

We found that the B(R283)–A(D61)–C(R402)–A(E40)–C(S406) network is critical for the connection of A, B, and C domains. Indeed, RYR1 B-A-C-A-C network mutants showed the abnormality in the caffeine-induced  $\text{Ca}^{2+}$  transient. MD simulations showed that the mutations loosen the connection between A and BC domains. The next question is whether the network is conserved in other subtypes. Comparison of the primary structures at the NTD revealed that most residues involved in the network formation are well conserved in RYR2 and RYR3: B(R298)–A(D61)–C(R417)–A(E40)–C(S421) and B(R287)–A(D62)–C(R409)–A(E41)–C(N413) for human RYR2 and RYR3, respectively. In addition, the B-A-C-A-C network is found in the crystal structures of the NTD of RYR2 (Borko et al., 2014; Fig. S6, A and B). There is an another reported crystal structure of the NTD of RYR2 (Kimlicka et al., 2013b). However this area is substantially different. There is a  $\text{Cl}^-$  bound at the interface and no B-A-C-A-C network. These findings suggest that the NTD in other subtypes may also play an important role in channel opening.

#### Interpretation of the results of the MD simulations

We performed MD simulations using the monomer structure of the NTD and found that B and C domains together rotated clockwise with respect to the A domain in the R402C/H mutants. However, because the RYR channel is a homotetramer, consideration of the movement in the tetramer structure is required. We therefore created a hypothetical homotetramer of NTD by fitting monomer structures before or after the MD simulations to the cryo-EM structure of RYR1 in the closed state (PDB accession no. 5TB0) at the A domain of the NTD monomer using ProFit. Fig. 6, A and B, shows the created hypothetical homotetramer before and after 50 ns of the simulation, respectively. The overall structure was the same before and after the simulation, verifying our MD simulation. Hypothetical homotetramers of the NTD of R402C (Fig. 6 C) and R402H (Fig. 6 D) mutants were also created using the same method as for the WT. Interestingly, the size of the central orifice of the NTD is larger in the mutants (Fig. 6 C, R402C,  $\sim 34$  Å; Fig. 6 D, R402H,  $\sim 36$  Å) than that of the WT (Fig. 6 B,  $\sim 27$  Å).

To interpret the findings, we also compared the cryo-EM structures of NTD in the closed state (PDB accession no. 5TB0; Fig. 6 E, left) and the open state (PDB accession no. 5T9V; Fig. 6 E, right). The diameter of the central orifice in the open state ( $\sim 29$  Å) was larger than that in the closed state ( $\sim 24$  Å). Importantly, there was essentially no change in the monomer structure of the NTD in the closed and open states (RMSD of  $\text{Ca}$ ,  $\sim 0.7$  Å; Fig. 6 F). Upon widening of the central orifice, NTD



**Figure 5. Verification of the B-A-C-A-C network.** (A) Caffeine-induced changes in  $[Ca^{2+}]_{cyt}$  determined by fura-2 in the WT and mutants (E40A and D61A). Caffeine was applied at the time points indicated by the black horizontal bar. (B and C) Comparison of the peaks of  $[Ca^{2+}]_{cyt}$  transiently induced by 0.3 mM (B;  $n = 161$ –233) and 10 mM (C;  $n = 161$ –245) caffeine in the WT (black) or mutants (gray). (D) Comparison of resting  $[Ca^{2+}]_{cyt}$  in the WT (black) and mutants (gray;  $n = 50$ –93). (E) Structural deviations observed in the MD simulations. Plots of the RMSD of the Ca atoms from the initial structures of NTD-RYR1s (E40A and D61A) as a function of time are shown. Calculations of each RMSD were performed using MD trajectories from the representative results. The trajectory during 20–50 ns (gray horizontal bar) was analyzed for hydrogen bonds/salt bridges (see F and G). (F and G) Analyses of probabilities of forming hydrogen bonds/salt bridges during the MD simulations. Hydrogen bonds/salt bridges between AC and BC domains are considerably strong. However, hydrogen bonds/salt bridges between AB domains are relatively weak. (H and I) Superimposition of the monomer of the WT (yellow) and of the E40A (H) or D61A (I) mutant after 50 ns of the MD simulation. The result shown here is a representative of 20 calculations. The BC domains rotated 5.7 degrees or 14.9 degrees with respect to the A domain in the E40A (H) or D61A (I) mutants, respectively. The average rotation angle is shown in Fig. S3 E. The rotation axis is indicated by a black

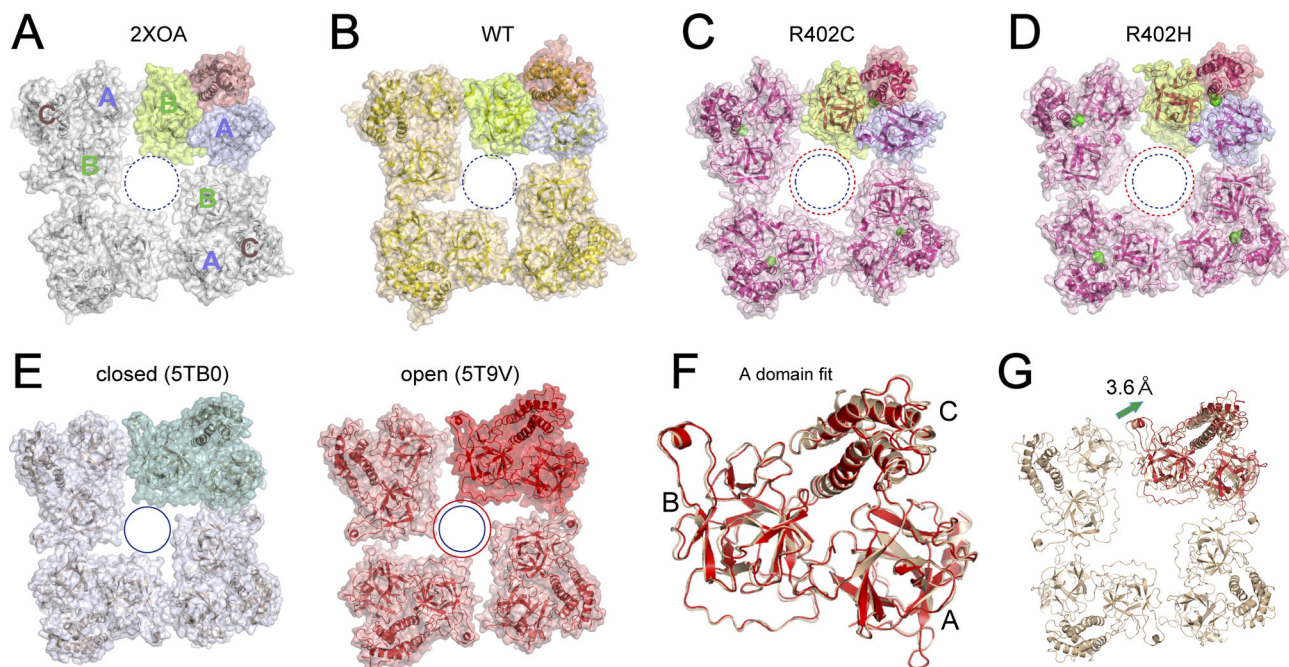
line. **(J and K)** Closeup view around residue 402 is the same as shown in Fig. 3. Colors of domains are the same as shown in Fig. 1. Dashed lines represent hydrogen bonds/salt bridges. The same view after 50 ns of the MD simulation of the E40A (J) or D61A (K) mutant is shown. Although E40A cannot form hydrogen bonds/salt bridges between R402 and S406 via E40, interaction between AB domains via D61 remains (J). However, in D61A, the interaction between AC domains via E40 remains, but there is no AB interaction. Values are shown as mean  $\pm$  SEM. \* $P < 0.0001$  by Student's  $t$  test compared with the WT.

translationally moves outward from the molecule, as indicated by the arrow in Fig. 6 G. Structural changes for widening of the central orifice might be related to the channel activity. Thus, it may be possible that the structure of the R402 is closer to the structure in the open state, but this mechanism may differ from that in the channel opening in WT.

#### MD simulation using the NTD tetramer

Recently, two papers describing MD simulations of the NTD have been reported, and they used the NTD tetramer based on the cryo-EM structure (Zheng and Liu, 2017; Xiong et al., 2018). Therefore, we also performed MD simulations of WT and R402C/H mutants using the NTD tetramer derived from cryo-EM structure (PDB accession no. 5GKZ) for 60 ns (Figs. S7 and S8). Unexpectedly, in both the simulations of WT and mutants, the RMSD value did not completely reach the plateau even after

the 60-ns calculation (Figs. S7 A and S8 A), whereas the RMSD value of each protomer reached a plateau ( $\sim 2$  Å) at 25 ns (Figs. S7 A and S8 A). This finding suggests that the structure of each protomer was not broken, but that the fourfold symmetry in the tetramer would be broken in the simulations. In the case of WT, there is not much difference in the overall structure between before (0 ns) and after 60 ns of the simulation (Fig. S7 B). However, each protomer is translationally and rotationally moved after simulation (Fig. S7 C). In the case of the mutants, in contrast, the structure of each protomer after 60 ns of simulation does not match that at 0 ns of the WT (Fig. S8 C). The BC domain rotated clockwise with respect to the A domain, although the angle in each monomer was widely distributed (Fig. S8 C). This observation is consistent with our results with monomers of the NTD (Fig. 3 B and Fig. S3 B). Hydrogen bonds/salt bridges within each protomer of the mutants were



**Figure 6. Hypothetical homotetramer structure of RYR1 after the MD simulation. (A–B)** The hypothetical homotetramer of the crystal structure of the RYR1 NTD (2XOA; Tung et al., 2010) (A) and the structure in the WT after 50 ns of the MD simulation (B) are represented by a ribbon and surface model. To create a hypothetical homotetramers of the NTDs, monomer structures are fitted to the cryo-EM structure of RYR1 in the closed state (5TB0). The blue dotted circle indicates the diameter of the central orifice. The radius of both circles shown in A and B is almost the same ( $\sim 27$  Å). **(C–D)** Hypothetical homotetramers of the R402C (C) and R402H (D) after 50 ns of the MD simulation. To create hypothetical homotetramers of the NTDs, mutants after the MD simulation were fitted to the cryo-EM structure in closed state using A domain in the NTD. The position of the mutated residue was indicated by green as the space filling model. Red dotted circles indicate the diameter of the central orifice in the R402C ( $\sim 34$  Å) and R402H ( $\sim 36$  Å) mutants, respectively. The blue dotted circle indicates the diameter of the central orifice shown in A and B ( $\sim 27$  Å). In the R402C/H mutations, the size of the central orifice becomes larger than that in the WT. **(E)** Comparison of the NTD of the cryo-EM structure in the closed (5TB0) and open (5T9V) state of RYR1. Blue and red circles indicate the diameter of the central orifice in closed ( $\sim 24$  Å) and open states ( $\sim 29$  Å), respectively. In the open state, the size of the central orifice becomes larger than that in the closed state. **(F)** Superimposition of the monomers of the cryo-EM structures in the closed state and open state. The monomer of the NTD in the open state was fitted to the A domain of the NTD in the closed state. **(G)** Superimposition of NTD of the cryo-EM structures in the closed state (light orange) and open state (red) is represented by the ribbon model. The structure in the open state is shown as a monomer. The NTD translationally moves 3.6 Å, as indicated by the arrow.



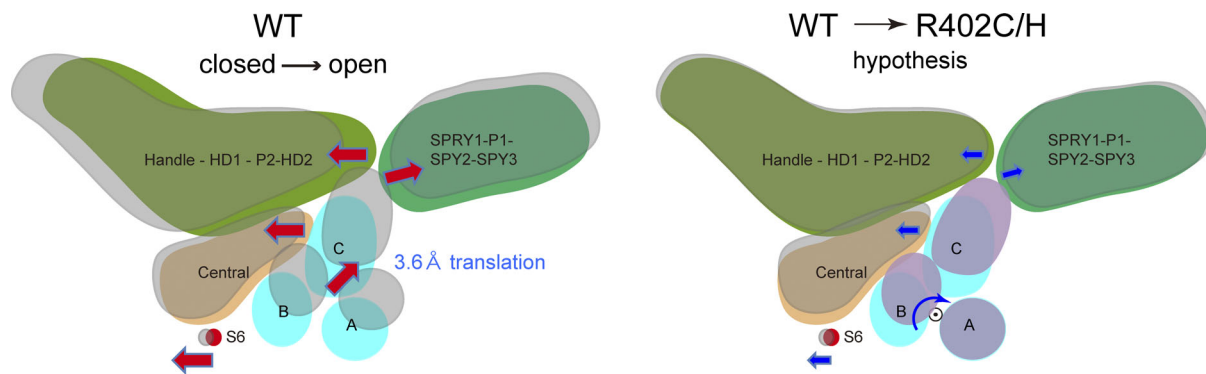


Figure 7. **Schematic diagram of the movements of each domain in the WT and hypothetical movements of each domain in the R402C/H mutants.** Each domain in the closed state is color coded: NTD is shown in light blue; Handle-HD1-P2-HD2 is shown in yellow-green; SPRY1-P1-SPRY2-SPRY3 is shown in light green; Central is shown in orange; and S6, the channel pore, is shown in red. Left: The movements of each domain in the WT. The place of each domain in the open state is shown transparently. Red arrows indicate the actual movements of the domains observed in the cryo-EM structures. Right: A hypothetical movement of each domain in the R402C/H mutants. The place of each domain in the mutants after 50 ns of the MD simulation is shown transparently. Blue arrows indicate the hypothetical movements.

essentially the same as the results using the monomer (Figs. S7 D and S8 D). These results indicate that the MD simulations themselves worked well and that what we observed in the simulations using the monomer occurred with the simulations using tetramer.

We assume that the reason for wide distribution of the angle could be the loss of cooperative movement between the monomer NTDs. In the RYR1 homotetramer, there are many communications between each monomer through transmembrane regions and other associating domains, which enable cooperative movement of NTD. However, the structure that we used in our simulations lacks any of these regions. Thus, each monomer moves independently without cooperativity. The ultimate way to resolve this problem could be calculations using the whole molecule. However, owing to the extremely large size of the molecule (a total molecular weight of 2.2 MDa) and limited resources, performing the calculation correctly is beyond the scope of our current research. An additional problem is incompleteness of the current cryo-EM model: ~15% of amino acid residues were not well identified, and ~19% of the whole model has not been built, due to its poor density.

#### Hypothesis of the underlying mechanism of the increased channel activity

How do the R402C/H mutants increase the open probability of the channel? A clue to understanding this mechanism is comprehension of the linkage between the movement of the NTD and channel pore in the WT structure. Because nearly the entire structures of RYR1 in the closed and open states solved by cryo-EM are available, we analyzed the movements of each domain when moving from the closed state to the open state. Fig. 7 (left) shows the relationship between the NTD and channel pore movement in the WT as represented by a cartoon. Along with the movement of S6, which constitutes a channel pore, the following four movements are linked, as indicated by the arrow in Fig. 7 (left): (1) Central, which is located above S6; (2) Handle-HD1-P2-HD2; (3) SPRY1-P1-SPRY2-SPRY3; and (4) the NTD. Although the exact sequence of the series of movements is not

fully clarified, it is certain that the movement of S6 is related to the movement of the NTD. As shown in Fig. 4 I, the position of the C domain moves slightly upward by the clockwise rotation of the BC domains in the R402H/C mutants. Interestingly, in terms of moving up for the C domain, a similar movement takes place in the WT during the opening of the channel pore. If so, the upward movement of the C domain in the R402C/H mutants may lead to the movement of Handle-HD1-P2-HD2, SPRY1-P1-SPRY2-SPRY3, and Central, the latter of which is located above S6. Therefore, this series of movements may ultimately increase the mobility of S6 and raise the open probability of a channel pore (Fig. 7, right).

In our MD simulations, we could not detect any effect of the mutations located around the subunit interfaces (e.g., Q156K, R164C, R164L, G342R, Y523C, and Y523S; Fig. S4). It has been reported that the mutations located around subunit interfaces may lead to reduced instability in the NTD tetramer, which may in turn serve to be structurally conducive to the open state and, simultaneously, a hindrance to the closed state (Zheng and Liu, 2017). These mutations might also affect the series of movements of the respective domains.

#### Structural comparison with the NT region of IP<sub>3</sub>R

IP<sub>3</sub>R is a ubiquitous ion channel and localized in the ER/SR, as is RYR. It is responsible for cytosolic Ca<sup>2+</sup> signaling and has crucial roles in a variety of physiological functions (Iino, 1990; Bezprozvanny et al., 1991; Berridge et al., 2003). In IP<sub>3</sub>R, inositol-1,4,5-triphosphate (IP<sub>3</sub>) binds to the NT region, which is located far from the channel pore and regulates the opening of the channel pore. This long-range regulation of the channel pore by the NT region in the IP<sub>3</sub>R is also common to RYR. Although IP<sub>3</sub>R and RYR have many structural similarities, the total number of amino acids in one monomer of IP<sub>3</sub>R is 2,700, which is ~55% of that in RYR. Thus, the structure of IP<sub>3</sub>R is considerably simple compared with that of RYR.

The x-ray crystal structures of NT of the IP<sub>3</sub>R in apo and IP<sub>3</sub>-bound states have already been determined, and the structural resemblance between the NT of the IP<sub>3</sub>R and the NTD of RYR1

has already been discussed (Seo et al., 2012). The x-ray crystal structure is composed of three domains: (1) the  $\beta$ -trefoil domain ( $\beta$ TF1) domain acting as a suppressor and (2)  $\beta$ TF2 and (3) armadillo solenoid folds (ARM) domains involved in the binding of  $\text{IP}_3$ . Surprisingly, the structures of the NT region ( $\text{IP}_3\text{R}$ ) and NTD (RYR) are very similar, and the superposition of both structures is in good agreement (Seo et al., 2012; Fig. S6 C). In  $\text{IP}_3\text{R}$ , binding of  $\text{IP}_3$  to the binding site causes movement of both  $\beta$ TF2 and ARM domains (corresponding to the B and C domains in the RYR), and the movement is considered to trigger the opening of the channel pore in the transmembrane region. In addition, a similar interaction between AC domains via R402 found in RYR (E40[A]-R402[C]-D61[A]) also exists in  $\text{IP}_3\text{R}$  between  $\beta$ TF1 and ARM (corresponding to the A and C domains in the RYR). However, in the case of the  $\text{IP}_3\text{R}$ , the charges of the three side chains are totally opposite, and those corresponding to E40(A), R402(C), and D61(A) are K127( $\beta$ TF1), D444(ARM), and R54( $\beta$ TF1), respectively (Seo et al., 2012). In  $\text{IP}_3\text{R}$ , the structural studies of the full-length receptor by cryo-EM have already been reported (Fan et al., 2015, 2018; Paknejad and Hite, 2018), and these studies also support the structural change of the NT caused by  $\text{IP}_3$  being the key to opening the channel pore in the transmembrane region.

## Conclusion

In this study, by combining  $[\text{Ca}^{2+}]_{\text{cyt}}$  measurements and MD simulations, we have shown a structural change in RYR1 with gain-of-function mutations at the domain interface of the NTD. The same line of analysis on the interdomain mutations in the NTD may lead to a deeper understanding of the mechanism enhancing channel opening.

## Acknowledgments

Eduardo Ríos served as editor.

We thank Dr. Tadashi Takemori (Tsukuba University) for valuable discussion and technical advice regarding conduct of the MD calculations, Dr. Shigeru Takemori (The Jikei University) for valuable comments, and Mariko Hanano and Shizuko Syoji (The Jikei University) for their technical assistance.

This work was supported by Japan Society for the Promotion of Science Grants-in-Aid for Scientific Research (grants 19H03198 and 19K07306 [T. Yamazawa], JP16H04748 [H. Ogawa], 19H03404 [T. Murayama], 16K08917 [K. Oguchi and H. Oyamada], 19K07105 [N. Kurebayashi], 15H05648 [K. Kanemaru], and 21229004 and 25221304 [M. Iino]), the Platform Project for Supporting Drug Discovery and Life Science Research (Basis for Supporting Innovative Drug Discovery and Life Science Research [BINDS] grants JP18am0101080 and JP19am0101080 [H. Ogawa and T. Murayama]), a grant from the Nakatani Foundation for Advancement of Measuring Technologies in Biomedical Engineering and The Jikei University School of Medicine (T. Yamazawa), and a National Center of Neurology and Psychiatry intramural research grant (29-4) for neurological and psychiatric disorders (T. Murayama).

The authors declare no competing financial interests.

Author contributions: T. Yamazawa and T. Murayama carried out the cell biological experiments. N. Kurebayashi, H. Oyamada, J. Suzuki, and K. Kanemaru provided experimental tools. T. Yamazawa, T. Murayama, K. Oguchi, T. Sakurai, and M. Iino analyzed the data. T. Yamazawa, M. Yamaguchi, and H. Ogawa performed the MD simulation and analysis. T. Yamazawa and H. Ogawa wrote the manuscript. All authors discussed the results and approved the final version of the manuscript.

Submitted: 4 October 2018

Revised: 31 July 2019

Accepted: 5 November 2019

## References

- Avila, G., and R.T. Dirksen. 2001. Functional effects of central core disease mutations in the cytoplasmic region of the skeletal muscle ryanodine receptor. *J. Gen. Physiol.* 118:277–290. <https://doi.org/10.1085/jgp.118.3.277>
- Bai, X.C., Z. Yan, J. Wu, Z. Li, and N. Yan. 2016. The central domain of RyR1 is the transducer for long-range allosteric gating of channel opening. *Cell Res.* 26:995–1006. <https://doi.org/10.1038/cr.2016.89>
- Berridge, M.J., M.D. Bootman, and H.L. Roderick. 2003. Calcium signalling: dynamics, homeostasis and remodelling. *Nat. Rev. Mol. Cell Biol.* 4: 517–529. <https://doi.org/10.1038/nrml155>
- Bezprozvanny, I., J. Watras, and B.E. Ehrlich. 1991. Bell-shaped calcium-response curves of  $\text{Ins}(1,4,5)\text{P}_3$ - and calcium-gated channels from endoplasmic reticulum of cerebellum [letter]. *Nature.* 351:751–754. <https://doi.org/10.1038/351751a0>
- Borko, L., V. Bauerová-Hlinková, E. Hostinová, J. Gašperík, K. Beck, F.A. Lai, A. Zahradníková, and J. Sevcík. 2014. Structural insights into the human RyR2 N-terminal region involved in cardiac arrhythmias. *Acta Crystallogr. D Biol. Crystallogr.* 70:2897–2912. <https://doi.org/10.1107/S1399004714020343>
- Case, D.A., T.E. Cheatham III, T. Darden, H. Gohlke, R. Luo, K.M. Merz Jr., A. Onufriev, C. Simmerling, B. Wang, and R.J. Woods. 2005. The Amber biomolecular simulation programs. *J. Comput. Chem.* 26:1668–1688. <https://doi.org/10.1002/jcc.20290>
- des Georges, A., O.B. Clarke, R. Zalk, Q. Yuan, K.J. Condon, R.A. Grassucci, W.A. Hendrickson, A.R. Marks, and J. Frank. 2016. Structural basis for gating and activation of RyR1. *Cell.* 167:145–157.
- Dirksen, R.T., and G. Avila. 2004. Distinct effects on  $\text{Ca}^{2+}$  handling caused by malignant hyperthermia and central core disease mutations in RyR1. *Biophys. J.* 87:3193–3204. <https://doi.org/10.1529/biophysj.104.048447>
- Eftremov, R.G., A. Leitner, R. Aebersold, and S. Raunser. 2015. Architecture and conformational switch mechanism of the ryanodine receptor. *Nature.* 517:39–43. <https://doi.org/10.1038/nature13916>
- Emsley, P., and K. Cowtan. 2004. Coot: model-building tools for molecular graphics. *Acta Crystallogr. D Biol. Crystallogr.* 60:2126–2132. <https://doi.org/10.1107/S0907444904019158>
- Endo, M. 2009. Calcium-induced calcium release in skeletal muscle. *Physiol. Rev.* 89:1153–1176. <https://doi.org/10.1152/physrev.00040.2008>
- Fan, G., M.L. Baker, Z. Wang, M.R. Baker, P.A. Sinyagovskiy, W. Chiu, S.J. Ludtke, and I.I. Serysheva. 2015. Gating machinery of  $\text{InsP}_3\text{R}$  channels revealed by electron cryomicroscopy. *Nature.* 527:336–341. <https://doi.org/10.1038/nature15249>
- Fan, G., M.R. Baker, Z. Wang, A.B. Seryshev, S.J. Ludtke, M.L. Baker, and I.I. Serysheva. 2018. Cryo-EM reveals ligand induced allostery underlying  $\text{InsP}_3\text{R}$  channel gating. *Cell Res.* 28:1158–1170. <https://doi.org/10.1038/s41422-018-0108-5>
- Fiser, A., R.K. Do, and A. Sali. 2000. Modeling of loops in protein structures. *Protein Sci.* 9:1753–1773. <https://doi.org/10.1110/ps.9.9.1753>
- Gryniewicz, G., M. Poenie, and R.Y. Tsien. 1985. A new generation of  $\text{Ca}^{2+}$  indicators with greatly improved fluorescence properties. *J. Biol. Chem.* 260:3440–3450.
- Hayward, S., and H.J. Berendsen. 1998. Systematic analysis of domain motions in proteins from conformational change: new results on citrate synthase and T4 lysozyme. *Proteins.* 30:144–154. [https://doi.org/10.1002/\(SICI\)1097-0134\(19980201\)30:2<144::AID-PROT4>3.0.CO;2-N](https://doi.org/10.1002/(SICI)1097-0134(19980201)30:2<144::AID-PROT4>3.0.CO;2-N)

- Humphrey, W., A. Dalke, and K. Schulten. 1996. VMD: visual molecular dynamics. *J. Mol. Graph.* 14:33–38, 27–28.
- Hwang, J.H., F. Zorzato, N.F. Clarke, and S. Treves. 2012. Mapping domains and mutations on the skeletal muscle ryanodine receptor channel. *Trends Mol. Med.* 18:644–657. <https://doi.org/10.1016/j.molmed.2012.09.006>
- Iino, M. 1990. Biphasic  $\text{Ca}^{2+}$  dependence of inositol 1,4,5-trisphosphate-induced Ca release in smooth muscle cells of the guinea pig taenia caeci. *J. Gen. Physiol.* 95:1103–1122. <https://doi.org/10.1085/jgp.95.6.1103>
- Iino, M. 1999. Molecular aspects of the excitation-contraction coupling in skeletal muscle. *Jpn. J. Physiol.* 49:325–333. <https://doi.org/10.2170/jjphysiol.49.325>
- Jorgensen, W.L., J. Chandrasekhar, J.D. Madura, R.W. Impey, and M.L. Klein. 1983. Comparison of simple potential functions for simulating liquid water. *J. Chem. Phys.* 79:926–935. <https://doi.org/10.1063/1.445869>
- Jungbluth, H., S. Treves, F. Zorzato, A. Sarkozy, J. Ochala, C. Sewry, R. Phadke, M. Gautel, and F. Muntoni. 2018. Congenital myopathies: disorders of excitation-contraction coupling and muscle contraction. *Nat. Rev. Neurol.* 14:151–167. <https://doi.org/10.1038/nrneurol.2017.191>
- Kakizawa, S., T. Yamazawa, Y. Chen, A. Ito, T. Murayama, H. Oyama, N. Kurebayashi, O. Sato, M. Watanabe, N. Mori, et al. 2012. Nitric oxide-induced calcium release via ryanodine receptors regulates neuronal function. *EMBO J.* 31:417–428. <https://doi.org/10.1038/emboj.2011.386>
- Kimlicka, L., K. Lau, C.C. Tung, and F. Van Petegem. 2013a. Disease mutations in the ryanodine receptor N-terminal region couple to a mobile inter-subunit interface. *Nat. Commun.* 4:1506. <https://doi.org/10.1038/ncomms2501>
- Kimlicka, L., C.C. Tung, A.C. Carlsson, P.A. Lobo, Z. Yuchi, and F. Van Petegem. 2013b. The cardiac ryanodine receptor N-terminal region contains an anion binding site that is targeted by disease mutations. *Structure*. 21: 1440–1449. <https://doi.org/10.1016/j.str.2013.06.012>
- Lanner, J.T., D.K. Georgiou, A.D. Joshi, and S.L. Hamilton. 2010. Ryanodine receptors: structure, expression, molecular details, and function in calcium release. *Cold Spring Harb. Perspect. Biol.* 2:a003996. <https://doi.org/10.1101/cshperspect.a003996>
- Law, R.J., C. Capener, M. Baaden, P.J. Bond, J. Campbell, G. Patargias, Y. Arinaminpathy, and M.S. Sansom. 2005. Membrane protein structure quality in molecular dynamics simulation. *J. Mol. Graph. Model.* 24: 157–165. <https://doi.org/10.1016/j.jmgm.2005.05.006>
- Marti-Renom, M.A., A.C. Stuart, A. Fiser, R. Sánchez, F. Melo, and A. Sali. 2000. Comparative protein structure modeling of genes and genomes. *Annu. Rev. Biophys. Biomol. Struct.* 29:291–325. <https://doi.org/10.1146/annurev.biophys.29.1.291>
- McDonald, I.K., and J.M. Thornton. 1994. Satisfying hydrogen bonding potential in proteins. *J. Mol. Biol.* 238:777–793.
- McLachlan, A.D. 1982. Rapid comparison of protein structures. *Acta Crystallogr. A* 38:871–873.
- Murayama, T., N. Kurebayashi, T. Yamazawa, H. Oyama, J. Suzuki, K. Kanamaru, K. Oguchi, M. Iino, and T. Sakurai. 2015. Divergent activity profiles of type 1 ryanodine receptor channels carrying malignant hyperthermia and central core disease mutations in the amino-terminal region. *PLoS One*. 10:e0130606. <https://doi.org/10.1371/journal.pone.0130606>
- Murayama, T., N. Kurebayashi, H. Ogawa, T. Yamazawa, H. Oyama, J. Suzuki, K. Kanamaru, K. Oguchi, M. Iino, and T. Sakurai. 2016. Genotype-phenotype correlations of malignant hyperthermia and central core disease mutations in the central region of the RyR1 channel. *Hum. Mutat.* 37:1231–1241. <https://doi.org/10.1002/humu.23072>
- Nakano, M., H. Oyama, T. Yamazawa, T. Murayama, H. Nanba, K. Iijima, and K. Oguchi. 2014. Construction and expression of ryanodine receptor mutants relevant to malignant hyperthermia patients in Japan. *Showa Univ. J. Med. Sci.* 26:27–38. <https://doi.org/10.15369/sujms.26.27>
- Paknejad, N., and R.K. Hite. 2018. Structural basis for the regulation of inositol trisphosphate receptors by  $\text{Ca}^{2+}$  and  $\text{IP}_3$ . *Nat. Struct. Mol. Biol.* 25: 660–668. <https://doi.org/10.1038/s41594-018-0089-6>
- Pancaroglu, R., and F. Van Petegem. 2018. Calcium channelopathies: structural insights into disorders of the muscle excitation-contraction complex. *Annu. Rev. Genet.* 52:373–396. <https://doi.org/10.1146/annurev-genet-120417-031311>
- Rebbeck, R.T., Y. Karunasekara, P.G. Board, N.A. Beard, M.G. Casarotto, and A.F. Dulhunty. 2014. Skeletal muscle excitation-contraction coupling: who are the dancing partners? *Int. J. Biochem. Cell Biol.* 48:28–38. <https://doi.org/10.1016/j.biocel.2013.12.001>
- Ríos, E. 2018. Calcium-induced release of calcium in muscle: 50 years of work and the emerging consensus. *J. Gen. Physiol.* 150:521–537. <https://doi.org/10.1085/jgp.201711959>
- Ríos, E., and G. Pizarro. 1991. Voltage sensor of excitation-contraction coupling in skeletal muscle. *Physiol. Rev.* 71:849–908. <https://doi.org/10.1152/physrev.1991.71.3.849>
- Robinson, R., D. Carpenter, M.A. Shaw, J. Halsall, and P. Hopkins. 2006. Mutations in RyR1 in malignant hyperthermia and central core disease. *Hum. Mutat.* 27:977–989. <https://doi.org/10.1002/humu.20356>
- Ryckaert, J.-P., G. Ciccotti, and H.J. Berendsen. 1977. Numerical integration of the Cartesian equations of motion of a system with constraints: molecular dynamics of n-alkanes. *J. Comput. Phys.* 23:327–341. [https://doi.org/10.1016/0021-9991\(77\)90098-5](https://doi.org/10.1016/0021-9991(77)90098-5)
- Seo, M.D., S. Velamakanni, N. Ishiyama, P.B. Stathopoulos, A.M. Rossi, S.A. Khan, P. Dale, C. Li, J.B. Ames, M. Ikura, and C.W. Taylor. 2012. Structural and functional conservation of key domains in  $\text{InsP}_3$  and ryanodine receptors. *Nature*. 483:108–112. <https://doi.org/10.1038/nature10751>
- Snoeck, M., B.G. van Engelen, B. Küsters, M. Lammens, R. Meijer, J.P. Molenaar, J. Raaphorst, C.C. Verschuren-Bemelmans, C.S. Straathof, L.T. Sie, et al. 2015. RyR1-related myopathies: a wide spectrum of phenotypes throughout life. *Eur. J. Neurol.* 22:1094–1112. <https://doi.org/10.1111/ene.12713>
- Tong, J., H. Oyama, N. Demareux, S. Grinstein, T.V. McCarthy, and D.H. MacLennan. 1997. Caffeine and halothane sensitivity of intracellular  $\text{Ca}^{2+}$  release is altered by 15 calcium release channel (ryanodine receptor) mutations associated with malignant hyperthermia and/or central core disease. *J. Biol. Chem.* 272:26332–26339. <https://doi.org/10.1074/jbc.272.42.26332>
- Tong, J., T.V. McCarthy, and D.H. MacLennan. 1999. Measurement of resting cytosolic  $\text{Ca}^{2+}$  concentrations and  $\text{Ca}^{2+}$  store size in HEK-293 cells transfected with malignant hyperthermia or central core disease mutant  $\text{Ca}^{2+}$  release channels. *J. Biol. Chem.* 274:693–702. <https://doi.org/10.1074/jbc.274.2.693>
- Tung, C.C., P.A. Lobo, L. Kimlicka, and F. Van Petegem. 2010. The amino-terminal disease hotspot of ryanodine receptors forms a cytoplasmic vestibule. *Nature*. 468:585–588. <https://doi.org/10.1038/nature09471>
- Webb, B., and A. Sali. 2014. Comparative protein structure modeling using MODELLER. *Curr. Protoc. Bioinformatics*. 47:5.6.1–32.
- Wei, R., X. Wang, Y. Zhang, S. Mukherjee, L. Zhang, Q. Chen, X. Huang, S. Jing, C. Liu, S. Li, et al. 2016. Structural insights into  $\text{Ca}^{2+}$ -activated long-range allosteric channel gating of RyR1. *Cell Res.* 26:977–994. <https://doi.org/10.1038/cr.2016.99>
- Willegems, K., and R.G. Efremov. 2018. Influence of lipid mimetics on gating of ryanodine receptor. *Structure*. 26:1303–1313.e4. <https://doi.org/10.1016/j.str.2018.06.010>
- Witherspoon, J.W., and K.G. Meilleur. 2016. Review of RyR1 pathway and associated pathomechanisms. *Acta Neuropathol. Commun.* 4:121. <https://doi.org/10.1186/s40478-016-0392-6>
- Xiong, J., X. Liu, Y. Gong, P. Zhang, S. Qiang, Q. Zhao, R. Guo, Y. Qian, L. Wang, L. Zhu, et al. 2018. Pathogenic mechanism of a catecholaminergic polymorphic ventricular tachycardia causing-mutation in cardiac calcium release channel RyR2. *J. Mol. Cell. Cardiol.* 117:26–35. <https://doi.org/10.1016/j.jmcc.2018.02.014>
- Yan, Z., X. Bai, C. Yan, J. Wu, Z. Li, T. Xie, W. Peng, C. Yin, X. Li, S.H.W. Scheres, et al. 2015. Structure of the rabbit ryanodine receptor RyR1 at near-atomic resolution. *Nature*. 517:50–55. <https://doi.org/10.1038/nature14063>
- Zalk, R., O.B. Clarke, A. des Georges, R.A. Grassucci, S. Reiken, F. Mancina, W.A. Hendrickson, J. Frank, and A.R. Marks. 2015. Structure of a mammalian ryanodine receptor. *Nature*. 517:44–49. <https://doi.org/10.1038/nature13950>
- Zheng, W., and Z. Liu. 2017. Investigating the intersubunit/subdomain interactions and motions relevant to disease mutations in the N-terminal domain of ryanodine receptors by molecular dynamics simulation. *Proteins*. 85:1633–1644. <https://doi.org/10.1002/prot.25318>



## Supplemental material

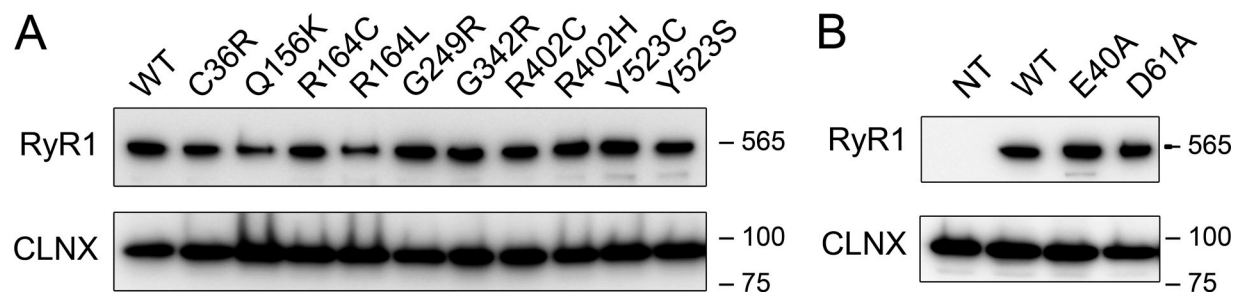
Yamazawa et al., <https://doi.org/10.1085/jgp.201812235>

Figure S1. **Western blotting of mutant RYR1 channels in HEK293 cells.** RYR1 carrying disease-associated mutations (C36R, Q156K, R164C, R164L, G249R, G342R, R402C, R402H, Y523C, and Y523S; A) or artificial mutations (E40A, D61A; B) was detected from HEK293 cell lysates. The mutant RYR1 channels showed gel mobility similar to that of WT, and the expression levels are almost the same as that of WT. Calnexin (CLNX) was used as a loading control. Numbers next to the blots are sizes of molecular mass standards (kDa).

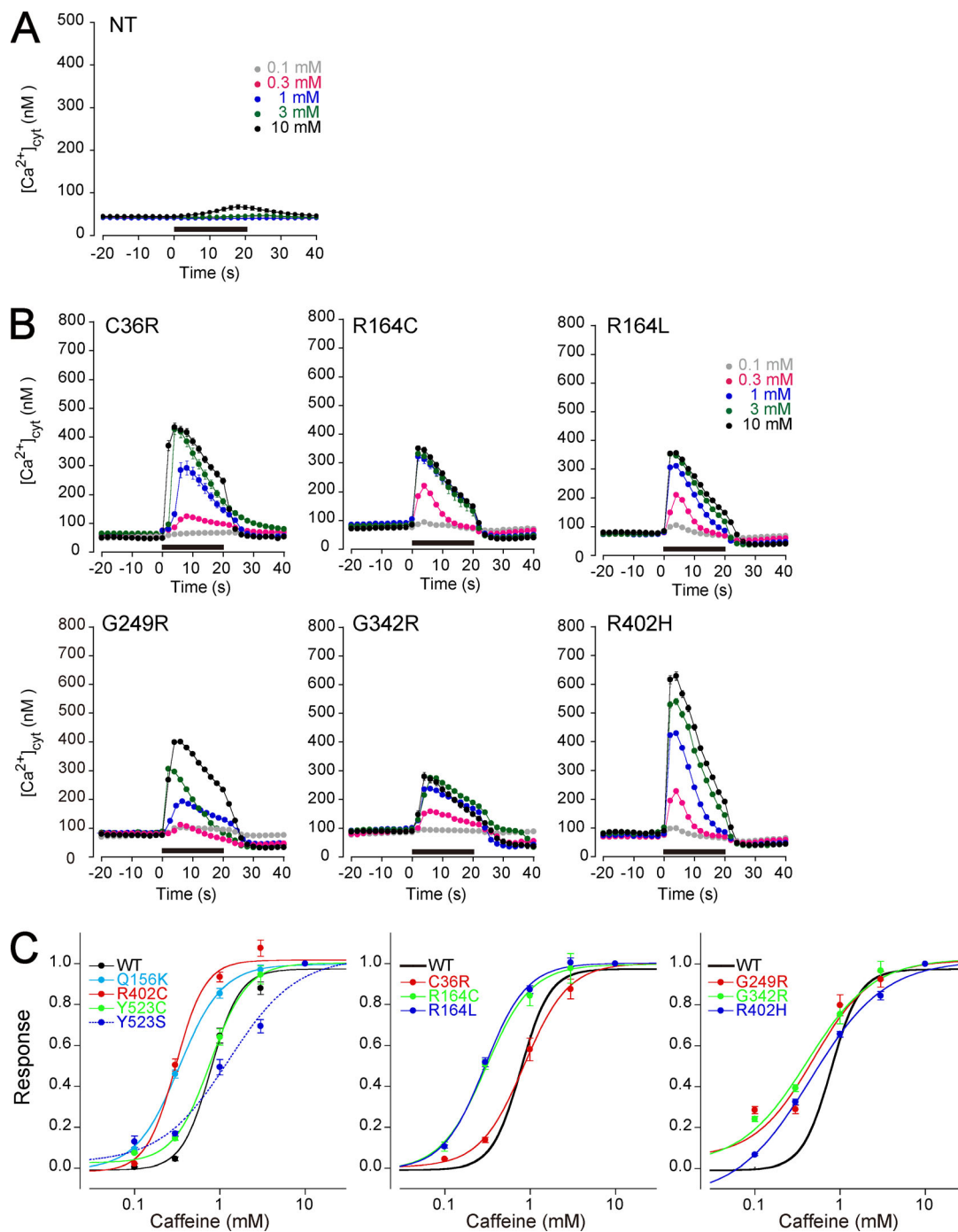


Figure S2. **Ca<sup>2+</sup> homeostasis in cells expressing WT or mutant RYR1s.** (A) Response to fura-2 signals for HEK cells without transfection of *RYR1* gene. Caffeine was applied at the time points indicated by the black horizontal bar. (B) Response to fura-2 signals in WT and mutants (C36R, R164C/L, G249R, G342R, and R402H). Caffeine was applied at the time points indicated by the black horizontal bar. (C) The magnitude of the Ca<sup>2+</sup> transients was plotted against caffeine concentrations and fitted to the dose-response curve.

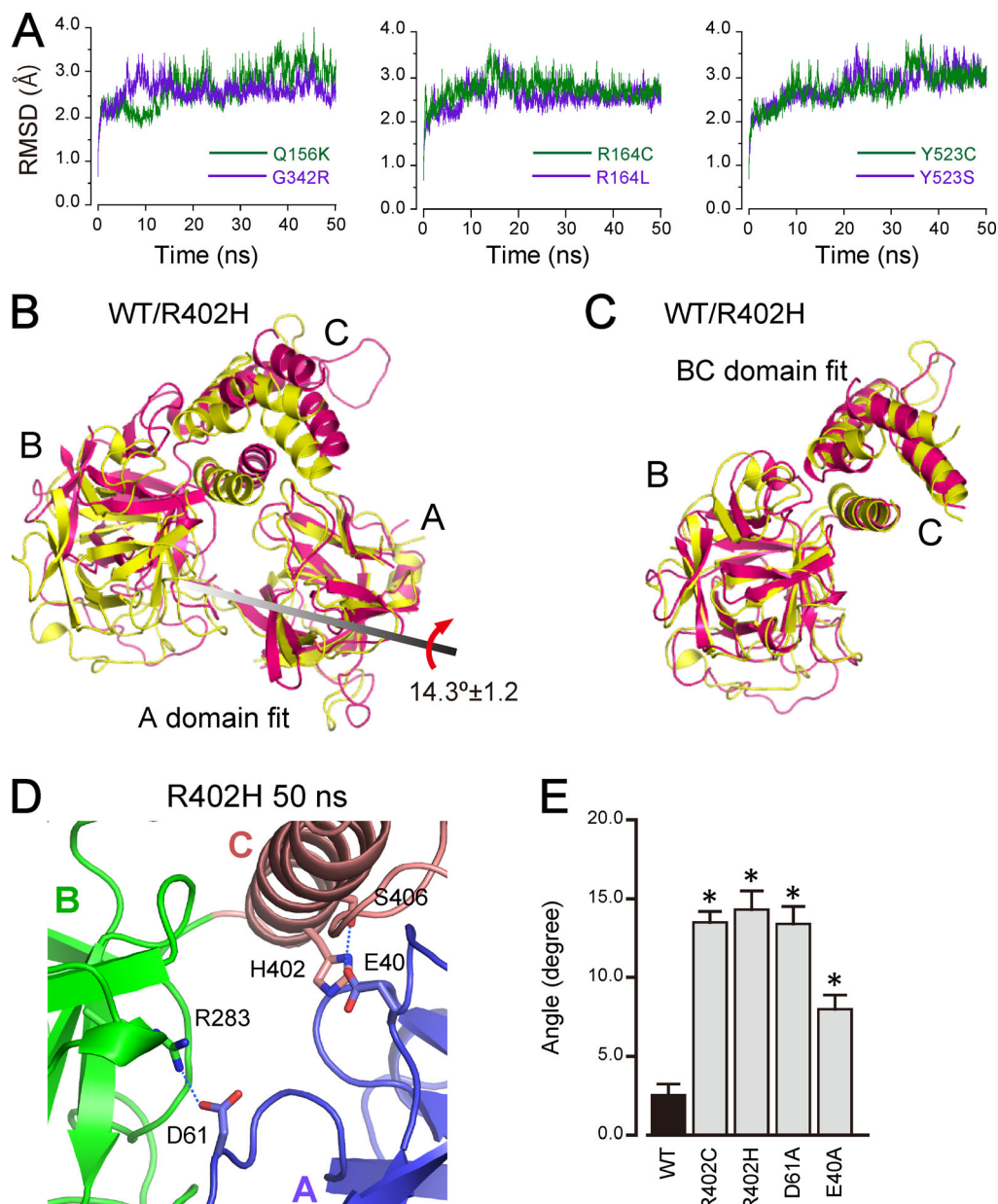


Figure S3. **Results of the MD simulations for the other mutants.** (A) RMSD plots for the MD simulation of the other mutants (Q156K, G342R, R164C, R164L, Y523C, and Y523S). (B) Superimposition of the monomers of the WT (yellow) and the R402C (pink) and R402H (magenta) mutants after 50 ns of the MD simulation. The result shown here is a representative of 20 calculations. The BC domains of the R402C mutant (pink), together, rotated clockwise by 14.3 degrees with respect to the A domain. The BC domains of the R402H mutant (magenta) rotated 21.6 degrees clockwise, which is more than that of the R402C mutant. (C) Superimposition of the BC domains of the WT and R402 mutant after the simulation. Almost all structures overlap. For simplicity, the upper portion of the C domain is deleted. (D) Closeup view around residue 402 in the R402H mutant after 50 ns of the MD simulation. Colors of domains are the same as shown in Fig. 1. H402 cannot form a hydrogen bond with D61. Thus, the C domain loses its interaction with the A domain. (E) Analysis of rotation angle in the WT and the mutants. Values are shown as mean  $\pm$  SEM. \* $P < 0.0001$  by  $t$  test compared with the WT.



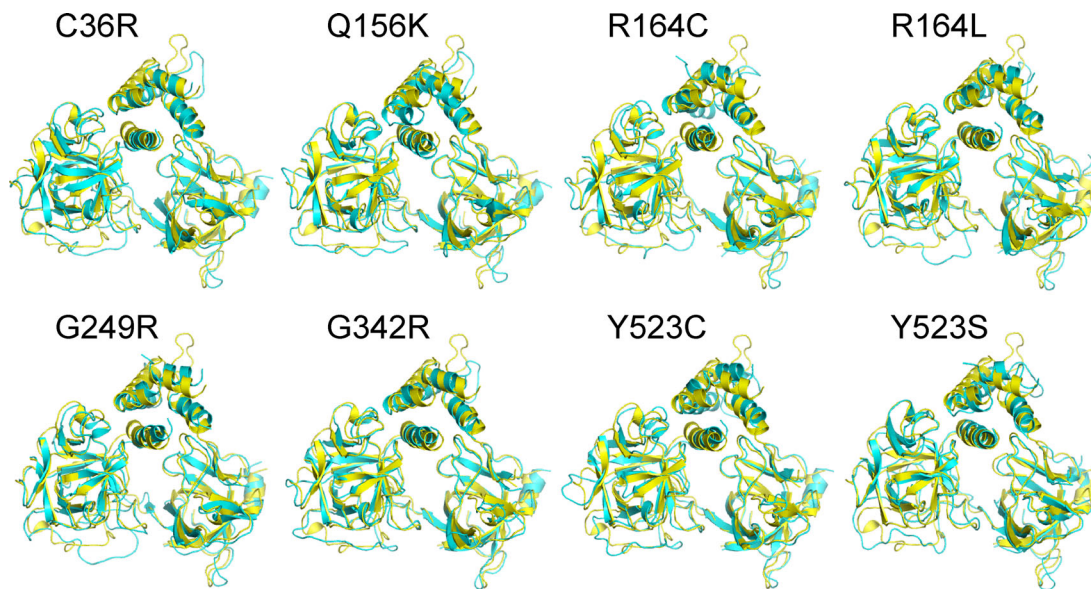


Figure S4. **Structure of the other mutants of RYR1 after the MD simulation.** Superimposition of the monomer of the WT (yellow) and other mutants (green) after 50 ns of the MD simulation.

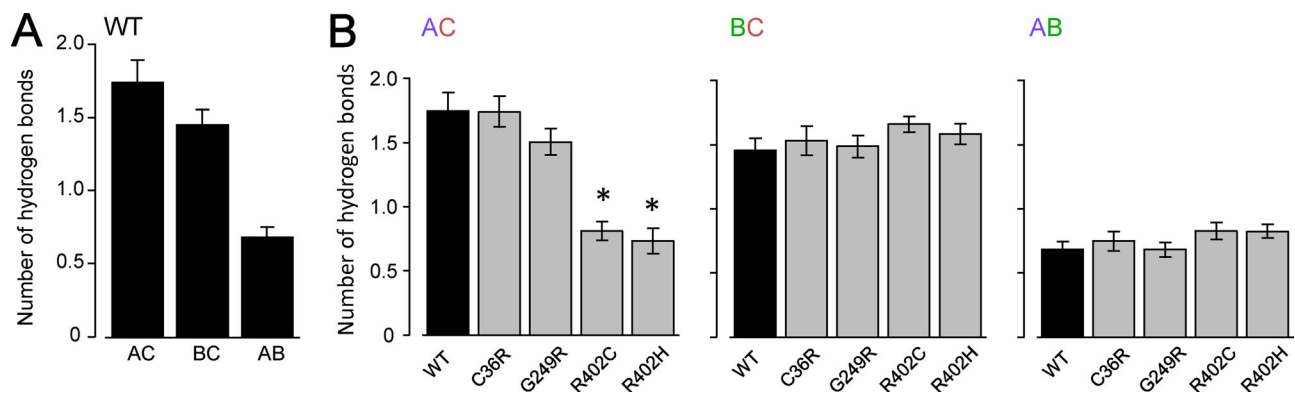


Figure S5. **Comparison of strength (tightness) of interdomain interactions in the NTD of the WT and mutant RYR1s.** Standard number of hydrogen bonds/salt bridges between domains during the simulation was counted. **(A)** In the WT, the number of hydrogen bonds/salt bridges between the A and B domains is less than that between the A and C domains or between the B and C domains. **(B)** Comparison of the standard number of hydrogen bonds/salt bridges in the WT and mutants. Note that in R402C/H mutations, the number of hydrogen bonds/salt bridges between the A and C domains is drastically decreased compared with the WT. Values are shown as mean  $\pm$  SEM. \* $P < 0.0001$  by  $t$  test compared with the WT.

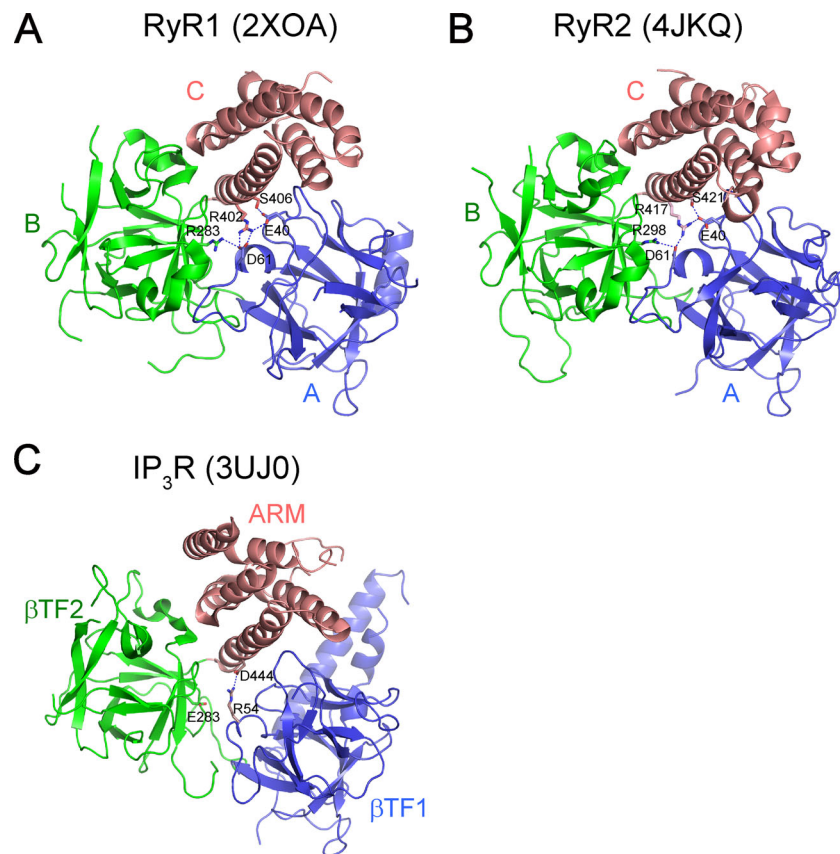


Figure S6. **The crystal structure of the RYRs and IP<sub>3</sub>R in the NTD.** **(A)** The crystal structures of the RYR1 NTD (PDB accession no. 2XOA; Tung et al., 2010). This figure is the same as Fig. 1 B. Each domain in the crystal structure of the NTD is colored in blue (A domain), green (B domain), and red (C domain). **(B)** The crystal structures of the RYR2 NTD (PDB accession no. 4JKQ; Borko et al., 2014). The three domains are colored in the same manner as in A. **(C)** The crystal structures of the IP<sub>3</sub>R NT (PDB accession no. 3UJ0; Seo et al., 2012). Each domain in the crystal structure of the IP<sub>3</sub>R is colored in blue ( $\beta$ TF1 domain corresponding to the C domain in the RYR), green ( $\beta$ TF2 domain corresponding to the C domain in the RYR), and red (ARM domain corresponding to the C domain in the RYR).

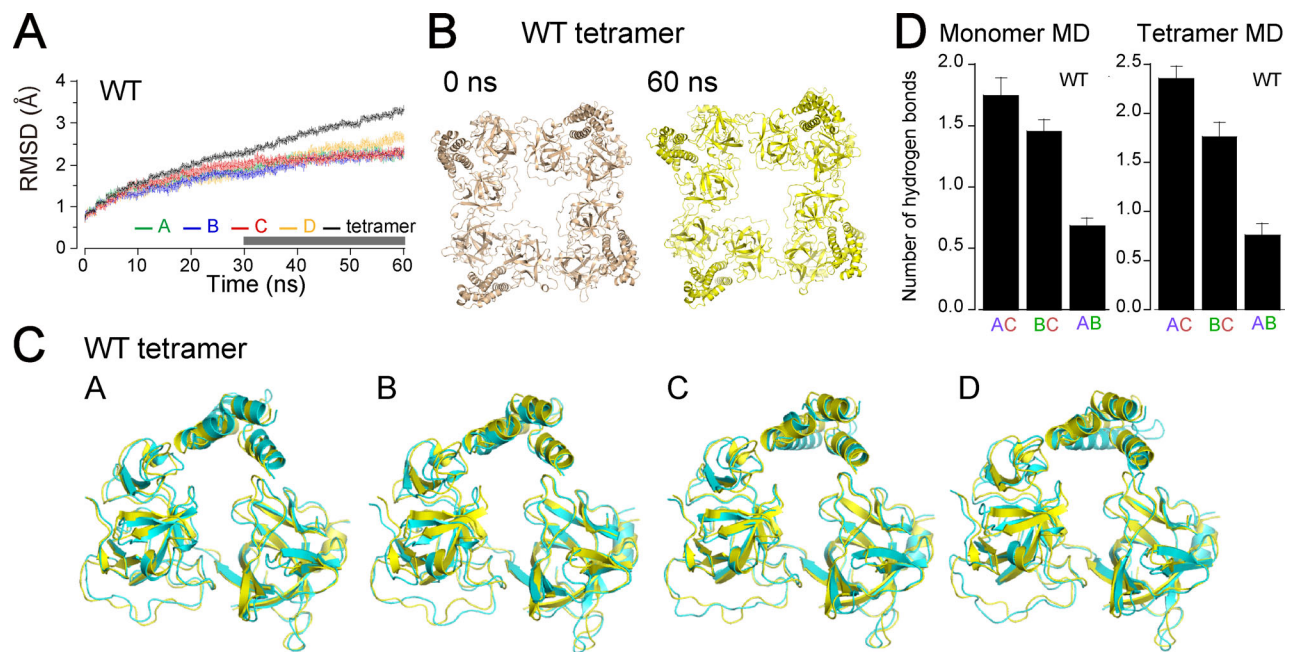


Figure S7. **MD simulation using the NTD tetramer of WT.** **(A)** Plots of the RMSD of the Ca atoms from the initial structure as a function of time. Calculations of each RMSD were performed using MD trajectories from the representative result. This fact suggests that the structure of each protomer was not broken but that the fourfold symmetry in the tetramer would be broken. **(B)** Tetramer structure of the WT before (left) and after (right) the MD simulation. **(C)** Superpositions of the structures of each protomer in the tetramer. Yellow represents the structure before the simulation, and cyan represents the structure after 60 ns of the simulation. The values of the angle of the rotation and the length of the translation for the superimposition of each monomer are as follows: protomer A (3.4 degrees, 6.1 Å), protomer B (3.1 degrees, 3.3 Å), protomer C (5.1 degrees, 17.5 Å), and protomer D (6.2 degrees, 14.4 Å). Interestingly, these values are not uniform. These results indicate that each protomer is stable, but the tetramer is unstable, during the MD simulation. **(D)** Analysis of hydrogen bonds/salt bridges at the domain interfaces in the simulation using a monomer (left) and tetramer (right). The trajectory during 30–60 ns (gray horizontal bar in A) was analyzed for hydrogen bonds/salt bridges. Values are shown as mean  $\pm$  SEM.



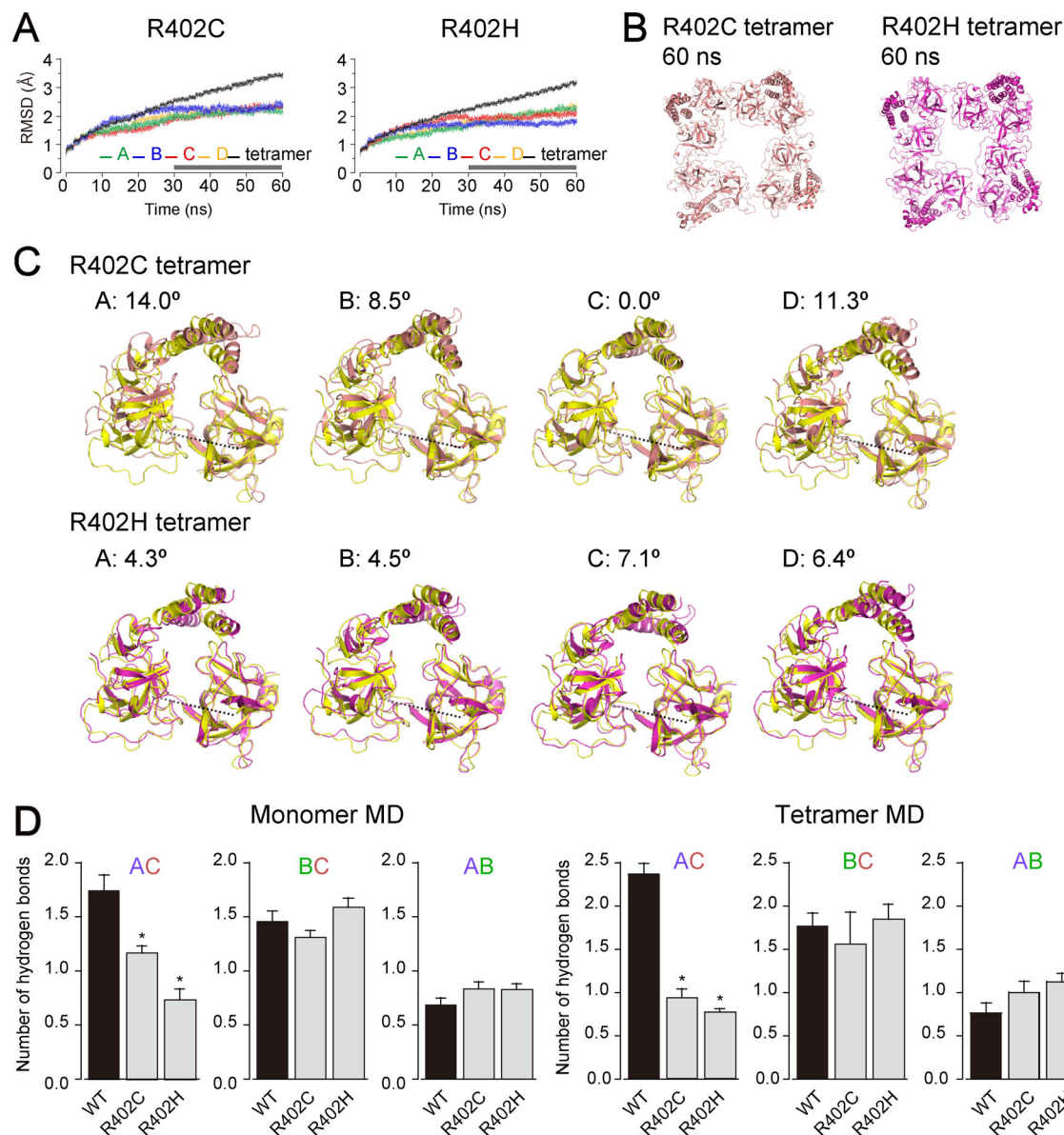


Figure S8. **MD simulation using the NTD tetramer of R402 mutants.** (A) Plots of the RMSD of the Ca atoms from the initial structures of R402 mutants as a function of time (left, R402C; right, R402H). Calculations of each RMSD were performed using MD trajectories from the representative result. (B) Tetramer structures of the R402C mutant (left) and R402H (right) after 60 ns of the MD simulation. (C) Superpositions of the structures of each protomer in the tetramer. Yellow represents the structure before the simulation. Magenta and orange represent the structures of R402C and R402H, respectively, after 60 ns of the simulation. The structure of each protomer does not match that at 0 ns of the WT. As observed in the simulation using the monomer of the R402C/H mutants, the BC domains rotate in a clockwise manner with respect to the A domain. The rotation angle of each monomer was found to be different for each protomer (the rotation angles for the R402C and R402H mutants were 0–14 degrees and 4.3–7.1 degrees, respectively). (D) Analysis of hydrogen bonds/salt bridges at the domain interfaces in the simulation using a monomer (left) and tetramer (right). The trajectory during 30–60 ns (gray horizontal bars) was analyzed for hydrogen bonds/salt bridges. Values are shown as mean  $\pm$  SEM. \* $P < 0.0001$  by Student's  $t$  test compared with the WT.

Table S1. Disease-associated mutations used in this study

Human RYR1 residue	Rabbit RYR1 residue	Disease status	Subdomain	Interface/buried	Functional characterization (References)
C35R <sup>a</sup>	C36	MH	A	AB	<a href="#">Tong et al., 1997, 1999; Murayama et al., 2015</a>
Q155K	Q156	MH	A	Interface 1	<a href="#">Nakano et al., 2014</a>
R163C	R164	MH/CCD	A	Interface 1, 4	<a href="#">Tong et al., 1997, 1999; Avila and Dirksen, 2001; Yang et al., 2003, 2007; Sato et al., 2010, 2013; Nakano et al., 2014; Murayama et al., 2015</a>
R163L	R164	MH	A	Interface 1, 4	<a href="#">Monnier et al., 2005; Murayama et al., 2015</a>
G248R <sup>a</sup>	G249	MH	B	Buried	<a href="#">Tong et al., 1997, 1999; Sato et al., 2010; Murayama et al., 2015</a>
G341R	G342	MH	B	Interface 5	<a href="#">Tong et al., 1997, 1999; Yang et al., 2003; Murayama et al., 2015</a>
R401C <sup>a</sup>	R402	MH	C	AC	<a href="#">Sato et al., 2013; Murayama et al., 2015</a>
R401H <sup>a</sup>	R402	MH	C	AC	<a href="#">Murayama et al., 2015</a>
Y522C	Y523	MH	C	Interface 3	<a href="#">Murayama et al., 2015</a>
Y522S	Y523	MH/CCD	C	Interface 3	<a href="#">Tong et al., 1997, 1999; Avila and Dirksen, 2001; Robinson et al., 2002; Dirksen and Avila, 2004; Brini et al., 2005; Murayama et al., 2015</a>

Classification of the domain and the interface is based on the structure of the rabbit NTD of RYR1 by Tong et al. (2010). The residue numbers of each domain are as follows: A domain (1–205), B domain (206–394), and C domain (395–532).

<sup>a</sup>Mutants used for the MD simulation.

Table S2. Disease-associated mutations located within the domains

Rabbit RYR1 residue	Human RYR1 residue	Subdomain	Mutations
D18	D17	A	D→N
E40	E39	A	E→K
R45	R44	A	R→C, R→H
D61	D60	A	D→N, D→Y
S207	S206	B	None
R221	R220	B	R→H
R242	R241	B	R→C, R→H
R283	R282	B	R→Q
T286	T285	B	None
E397	E396	C	E→A
R402	R401	C	R→C, R→H, R→G, R→L, R→S
S406	S405	C	None
D447	D446	C	None
E481	E480	C	E→L

Classification of the domain and the interface is based on the structure of the rabbit NTD of RYR1 by Tong et al. (2010). The residue numbers of each domain are as follows: A domain (1–205), B domain (206–394), and C domain (395–532). Mutant residues indicate human mutations associated with MH/CCD disease ([https://www.cardiodb.org/paralogue\\_annotation/](https://www.cardiodb.org/paralogue_annotation/)). No mutation has been reported for S207, T286, S406, and D447.

## References

- Avila, G., and R.T. Dirksen. 2001. Functional effects of central core disease mutations in the cytoplasmic region of the skeletal muscle ryanodine receptor. *J. Gen. Physiol.* 118:277–290. <https://doi.org/10.1085/jgp.118.3.277>
- Brini, M., S. Manni, N. Pierobon, G.G. Du, P. Sharma, D.H. MacLennan, and E. Carafoli. 2005. Ca<sup>2+</sup> signaling in HEK-293 and skeletal muscle cells expressing recombinant ryanodine receptors harboring malignant hyperthermia and central core disease mutations. *J. Biol. Chem.* 280:15380–15389. <https://doi.org/10.1074/jbc.M410421200>

- Carpenter, D., R.L. Robinson, R.J. Quinnell, C. Ringrose, M. Hogg, F. Casson, P. Booms, D.E. Iles, P.J. Halsall, D.S. Steele, et al. 2009. Genetic variation in RYR1 and malignant hyperthermia phenotypes. *Br. J. Anaesth.* 103:538–548. <https://doi.org/10.1093/bja/aep204>
- Dirksen, R.T., and G. Avila. 2004. Distinct effects on  $\text{Ca}^{2+}$  handling caused by malignant hyperthermia and central core disease mutations in RyR1. *Biophys. J.* 87: 3193–3204. <https://doi.org/10.1529/biophysj.104.048447>
- Monnier, N., G. Kozak-Ribbens, R. Krivosic-Horber, Y. Nivoche, D. Qi, N. Kraev, J. Loke, P. Sharma, V. Tegazzin, D. Figarella-Branger, et al. 2005. Correlations between genotype and pharmacological, histological, functional, and clinical phenotypes in malignant hyperthermia susceptibility. *Hum. Mutat.* 26: 413–425. <https://doi.org/10.1002/humu.20231>
- Murayama, T., N. Kurebayashi, T. Yamazawa, H. Oyamada, J. Suzuki, K. Kanemaru, K. Oguchi, M. Iino, and T. Sakurai. 2015. Divergent activity profiles of type 1 ryanodine receptor channels carrying malignant hyperthermia and central core disease mutations in the amino-terminal region. *PLoS One.* 10: e0130606. <https://doi.org/10.1371/journal.pone.0130606>
- Nakano, M., H. Oyamada, T. Yamazawa, T. Murayama, H. Nanba, K. Iijima, and K. Oguchi. 2014. Construction and expression of ryanodine receptor mutants relevant to malignant hyperthermia patients in Japan. *Showa Univ. J. Med. Sci.* 26:27–38. <https://doi.org/10.15369/sujms.26.27>
- Robinson, R.L., C. Brooks, S.L. Brown, F.R. Ellis, P.J. Halsall, R.J. Quinnell, M.A. Shaw, and P.M. Hopkins. 2002. RYR1 mutations causing central core disease are associated with more severe malignant hyperthermia in vitro contracture test phenotypes. *Hum. Mutat.* 20:88–97. <https://doi.org/10.1002/humu.10098>
- Sato, K., N. Pollock, and K.M. Stowell. 2010. Functional studies of RYR1 mutations in the skeletal muscle ryanodine receptor using human RYR1 complementary DNA. *Anesthesiology.* 112:1350–1354. <https://doi.org/10.1097/ALN.0b013e3181d69283>
- Sato, K., C. Roesl, N. Pollock, and K.M. Stowell. 2013. Skeletal muscle ryanodine receptor mutations associated with malignant hyperthermia showed enhanced intensity and sensitivity to triggering drugs when expressed in human embryonic kidney cells. *Anesthesiology.* 119:111–118. <https://doi.org/10.1097/ALN.0b013e31828cebfe>
- Tong, J., H. Oyamada, N. Demareux, S. Grinstein, T.V. McCarthy, and D.H. MacLennan. 1997. Caffeine and halothane sensitivity of intracellular  $\text{Ca}^{2+}$  release is altered by 15 calcium release channel (ryanodine receptor) mutations associated with malignant hyperthermia and/or central core disease. *J. Biol. Chem.* 272:26332–26339. <https://doi.org/10.1074/jbc.272.42.26332>
- Tong, J., T.V. McCarthy, and D.H. MacLennan. 1999. Measurement of resting cytosolic  $\text{Ca}^{2+}$  concentrations and  $\text{Ca}^{2+}$  store size in HEK-293 cells transfected with malignant hyperthermia or central core disease mutant  $\text{Ca}^{2+}$  release channels. *J. Biol. Chem.* 274:693–702. <https://doi.org/10.1074/jbc.274.2.693>
- Yang, T., T.A. Ta, I.N. Pessah, and P.D. Allen. 2003. Functional defects in six ryanodine receptor isoform-1 (RyR1) mutations associated with malignant hyperthermia and their impact on skeletal excitation-contraction coupling. *J. Biol. Chem.* 278:25722–25730. <https://doi.org/10.1074/jbc.M302165200>
- Yang, T., E. Esteve, I.N. Pessah, T.F. Molinski, P.D. Allen, and J.R. López. 2007. Elevated resting  $[\text{Ca}^{2+}]_i$  in myotubes expressing malignant hyperthermia RyR1 cDNAs is partially restored by modulation of passive calcium leak from the SR. *Am. J. Physiol. Cell Physiol.* 292:C1591–C1598. <https://doi.org/10.1152/ajpcell.00133.2006>