

Kinetics of M₁ muscarinic receptor and G protein signaling to phospholipase C in living cells

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G protein-coupled receptors (GPCRs) mediate responses to external stimuli in various cell types. Early events, such as the binding of ligand and G proteins to the receptor, nucleotide exchange (NX), and GTPase activity at the G α subunit, are common for many different GPCRs. For G $_q$ -coupled M₁ muscarinic (acetylcholine) receptors (M₁Rs), we recently measured time courses of intermediate steps in the signaling cascade using Förster resonance energy transfer (FRET). The expression of FRET probes changes the density of signaling molecules. To provide a full quantitative description of M₁R signaling that includes a simulation of kinetics in native (tsA201) cells, we now determine the density of FRET probes and construct a kinetic model of M₁R signaling through G $_q$ to activation of phospholipase C (PLC). Downstream effects on the trace membrane lipid phosphatidylinositol 4,5-bisphosphate (PIP₂) and PIP₂-dependent KCNQ2/3 current are considered in our companion paper in this issue (Falkenburger et al. 2010. *J. Gen. Physiol.* doi:10.1085/jgp.200910345). By calibrating their fluorescence intensity, we found that we selected transfected cells for our experiments with $\sim 3,000$ fluorescently labeled receptors, G proteins, or PLC molecules per μm^2 of plasma membrane. Endogenous levels are much lower, 1–40 per μm^2 . Our kinetic model reproduces the time courses and concentration–response relationships measured by FRET and explains observed delays. It predicts affinities and rate constants that align well with literature values. In native tsA201 cells, much of the delay between ligand binding and PLC activation reflects slow binding of G proteins to receptors. With M₁R and G β FRET probes overexpressed, 10% of receptors have G proteins bound at rest, rising to 73% in the presence of agonist. In agreement with previous work, the model suggests that binding of PLC to G α_q greatly speeds up NX and GTPase activity, and that PLC is maintained in the active state by cycles of rapid GTP hydrolysis and NX on G α_q subunits bound to PLC.

INTRODUCTION

G protein-coupled receptors (GPCRs) transduce extracellular signals to initiate intracellular signaling cascades. This large receptor family serves diverse physiological functions and constitutes a good portion of pharmaceutical targets (Sautel and Milligan, 2000). We describe kinetic studies as one way to understand how GPCR signaling works. We focus on activation of the G $_q$ -coupled M₁ muscarinic (acetylcholine) receptor (M₁R) and its consequences: activation of PLC, depletion of phosphatidylinositol 4,5-bisphosphate (PIP₂), and closure of PIP₂-dependent KCNQ channels (Suh et al., 2004; Jensen et al., 2009). M₁R signaling increases excitability in mammalian sympathetic neurons (Brown, 1983) and augments hippocampal long-term potentiation (e.g., Shinoe et al., 2005).

Using Förster resonance energy transfer (FRET) in an expression system, we have previously measured time courses of individual steps in the M₁R signaling cascade

(Jensen et al., 2009). The first steps include the binding of the muscarinic agonist to the M₁R, activation of G proteins and the binding of the G α_q subunit to PLC (Fig. 1). FRET allowed us to track the interactions of signaling proteins within intact cells. We monitored G protein activity by measuring both the interaction of G β subunits with receptors, which increased in the presence of agonist, and the interaction of G β with G α , which decreased in the presence of agonist. The time courses of these interactions were different, and we interpreted them in the classical sense, that G proteins are recruited to the receptors with agonist and dissociate upon nucleotide exchange (NX). It is nonetheless possible that conformational rearrangement instead of binding/unbinding underlies these changes.

The FRET probes needed to be overexpressed, with each signaling step being reconstituted using a different set of transfected proteins. Such changes in the amounts of signaling molecules affected the timing. To better analyze our measurements, we have therefore determined the amounts of expressed and endogenous

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Abbreviations used in this paper: CFP, cyan fluorescent protein; FRET, Förster resonance energy transfer; FSc, cyan fluorescent beads; FSy, yellow fluorescent beads; GAP, GTPase accelerating protein; GEF, guanine nucleotide exchange factor; GPCR, G protein-coupled receptor; M₁R, M₁ muscarinic (acetylcholine) receptor; NX, nucleotide exchange; PH, pleckstrin homology domain (binds PIP₂); PIP₂, phosphatidylinositol 4,5-bisphosphate; YFP, yellow fluorescent protein.

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signaling molecules and constructed a kinetic model of the M₁R signaling cascade. In quantifying FRET probes, the present study follows the lead of several studies that determined the density of fluorescent molecules by comparing intensities of cells to fluorescent beads and to solutions of recombinant fluorescent proteins (Chiu et al., 2001; An and Almers, 2004; Sugiyama et al., 2005).

We then combine this molecular census with our kinetic observations (Jensen et al., 2009) to develop a comprehensive, quantitative description for M₁R signaling. Previous models of GPCR signaling have been helpful in explaining high-affinity and low-affinity binding sites for ligands at GPCRs (De Lean et al., 1980), inverse agonism (Kinzer-Ursem and Linderman, 2007), GTPase-activating properties of PLC (Bornheimer et al., 2004; Turcotte et al., 2008), and modulation by regulators of G protein signaling (Hao et al., 2003). As our FRET time courses (Jensen et al., 2009) were collected from different steps of the M₁R signaling cascade in living cells, the model informed by these measurements allows us to extract kinetic rate constants for many steps of M₁R signaling, to ask whether our understanding of signaling is self-consistent and sufficient to explain the observed delays, and to suggest more general mechanisms by which GPCR signaling may function. Our companion paper (see Falkenburger et al. in this issue) makes new measurements of lipid metabolism and extends the model to regulation of phosphoinositides and KCNQ channels.

MATERIALS AND METHODS

Cell culture and plasmids

Cells (tsA201) cultured in DMEM (Invitrogen) with 10% serum and 0.2% penicillin/streptomycin were passaged once a week. Cells were transfected at 75% confluency, plated on polylysine-coated glass chips 24 h after transfection, and used for experiments the next day. Cells were transfected with Lipofectamine 2000 (10 μ l for a 3-cm dish; Invitrogen) and 0.1–1.2 μ g DNA per plasmid: mouse M₁R (provided by N. Nathanson, University of Washington, Seattle, WA); cerulean-tagged M₁R (M₁R-CFP) and M₁R-YFP-CFP (Jensen et al., 2009); mouse G α_q -eCFP (provided by C. Berlot, Geisinger Clinic, Danville, PA); bovine eYFP-G β 1 (provided by S. Ikeda, National Institutes of Health [NIH], Rockville, MD); rat eYFP-PLC β 1 (provided by L. Runnels, University of Medicine and Dentistry of New Jersey, Piscataway, NJ); human eCFP-PH(PLC δ 1) and eYFP-PH(PLC δ 1) (provided by K. Jalink, The Netherlands Cancer Institute, Amsterdam, Netherlands); human KCNQ2 and rat KCNQ3 (provided by D. McKinnon, State University of New York, Stony Brook, NY); and unlabeled human G α_q , G β 1, and G γ 2 (The Missouri S&T cDNA Resource Center). Hereafter, we refer to fluorophores simply as cyan fluorescent protein (CFP) or yellow fluorescent protein (YFP), regardless of whether regular or enhanced fluorescent proteins were used.

Flow cytometry

Cells were harvested, resuspended in Ringer's buffer, and analyzed by a FACScan (BD) flow cytometer using a 488-nm laser and a 530/30-nm emission filter. Parameters were FSC E00, SSC 359,

and FL1 401. Cells were selected based on forward and sideward scatter, using the same gate for all samples. At least 10,000 cells were counted per sample. A different gate and FSC E01 were used to measure fluorescent beads. Data were analyzed offline using FlowJo software (Tree Star, Inc.) and visualized with IGOR Pro 6.0 (Wavemetrics). Fluorescence intensity histograms were computed and displayed on a logarithmic scale for fluorescence intensity, which more closely mimics our visual impression than a linear scale.

Western blot

Cells were harvested in PBS, centrifuged, and resuspended in 50 μ l of lysis buffer: PBS with 1% Triton X-100 and protease inhibitor cocktail (Thermo Fisher Scientific). Lysates were cleared by centrifugation (13,000 *g* for 20 min at 4°C), and the supernatant was transferred to new tubes. Protein concentration was determined by Bradford assay (Bio-Rad Laboratories). 20 μ g of protein was separated by SDS-PAGE using standard techniques (as in Krenz et al., 2009). The primary antibody against PLC β 1 was used at 1:1,000 (BD Transduction Laboratories). The secondary antibody was coupled to horseradish peroxidase and visualized by chemiluminescence (AlphaImager; AlphaInnotech).

Recombinant CFP and YFP protein

His-tagged ECFP and EYFP DNA were cloned into pETGQ and transformed into *Escherichia coli* BL-21 (DE3) cells, which were grown at 37°C in Luria broth for 4 h. To induce expression, isopropyl-1-thio- β -D-galactopyranoside was added to a concentration of 1 μ M, and cells were grown overnight at 18°C. Bacteria were collected by centrifugation and resuspended in ice-cold lysis buffer. The lysate was harvested using a homogenizer and cleared by centrifugation, and the constructs were purified by Ni²⁺-NTA chromatography. Proteins were checked for contaminants by comparing Coomassie-stained bands on SDS-PAGE from 20 and 1 μ g of protein. Absorbance was measured with a spectrophotometer (DU640; Beckman Coulter). For comparison of fluorescence intensities between CFP/YFP solutions and bead suspensions, emission spectra were acquired with a fluorometer (10-nm excitation slit, 3-nm emission slit, and 1-s sampling per point; Fluorolog FL-1039; Horiba Jobin Yvon).

Photometric measurement of proteins

To estimate expressed protein amounts, the fluorescence of live cells was measured in the epifluorescence photometry setup used in Jensen et al. (2009). In brief, a xenon arc lamp was used for excitation with 440/20- or 500/20-nm excitation filters; 480/30- and 535/25-nm emission filters were used for CFP and YFP channels. Fluorescence was detected by photon-counting photomultiplier tubes. We estimated the number of fluorescent proteins from the brightness of each cell. To be consistent with our previous work and to be able to compare the relative amounts of CFP and YFP, we co-transfected both CFP- and YFP-tagged proteins. When appropriate, untagged G α , G β , and G γ were included to complement any tagged G protein subunits (see legend to Fig. 3 A). As FRET is expected to reduce CFP fluorescence, we took the values for YFP fluorescence before acceptor photobleaching and for CFP fluorescence after acceptor photobleaching. To bleach \sim 95% of YFP, we illuminated with the YFP filter set for 5 min, omitting the neutral density filter (0.2 ND) used for time course measurements. FRET efficiency was determined from the increase in CFP fluorescence upon acceptor photobleaching. The time constant for YFP bleaching was 51 s with continuous illumination. Using a 350 linear/log optometer (United Detector Technology Instruments), the light energy of the CFP-measuring light at the stage was ninefold less than that of the YFP-bleaching light.

We assume that we sample light from the entire cell thickness when our imaging setup (40 \times oil objective; N.A. 1.3) is focused on

a cell, as with a 1- μm diameter fluorescent bead, recorded fluorescence intensity changed only 5% within a 10- μm focus range around the z-plane of maximal intensity.

Modeling

A kinetic model of M_1R signaling was formulated as a compartmental model in the Virtual Cell framework (University of Connecticut). The Virtual Cell Model “FalkenburgerJGP2010” is available at <http://www.vcell.org/> under shared models/hillelab. Model species and reactions are shown in Fig. 4. Model initial conditions are listed in Table II. They are mostly derived from measurements. Model rate constants are listed in Table IV. They were chosen manually and were not optimized by a statistical fit. When comparing model outputs to FRET data, we combine all molecular species that include the tagged pair of molecules. Thus, a term such as “receptor with ligand,” for example, refers to the sum of all species that include both receptor (R) and ligand (L).

Statistics

Summarized data include one data point per cell. Bars and markers represent mean \pm SEM.

RESULTS

The signaling steps from M_1R excitation to PIP_2 depletion and channel modulation are shown diagrammatically in Fig. 1. KCNQ channels need PIP_2 to function. Ligand binding to M_1R s allows the G protein G_q to activate the enzyme PLC, which hydrolyzes PIP_2 , thus shutting off the channel. Our laboratory has already formulated a preliminary kinetic description of these steps (Suh et al., 2004). It was based on published test tube measurements of the G protein cycle and plausible estimates of the subsequent steps from the final modulation of KCNQ channels from our studies. Subsequently, we measured the kinetics of four of the intermediate reactions in living cells using transfected FRET reporters for each step (Jensen et al., 2009). Nevertheless, to refine a more realistic kinetic model still requires several more measurements, particularly better estimates of the concentrations of reactants.

Transfected cells we select for study express roughly 3,000 proteins per μm^2 of plasma membrane

The study of Jensen et al. (2009) transfected CFP- and YFP-labeled signaling proteins into cells for optical measurements of the progression of muscarinic signals through intermediate signaling steps. We now want to estimate how many copies of the transfected molecules were expressed in those cells. Because no method seemed ideal, we accumulated determinations by numerous techniques based on the following five measurements. (1) We calibrated fluorescent beads (1- μm diameter) as fluorescence intensity standards by comparing their fluorescence to purified CFP and YFP solutions in a fluorometer. Transfected cells could then be compared with the calibrated beads. (2) We allowed the cytoplasm of untransfected cells to equilibrate with a CFP solution of known concentration in a whole cell patch pipette and measured the fluorescence intensity. (3) We estimated the density of expressed pleckstrin homology domain (PH)-YFP acceptor molecules from FRET efficiency. (4) We compared whole populations of transfected cells with fluorescent beads by flow cytometry. (5) For PLC expression, we compared levels of transfected proteins with levels of endogenous proteins using Western blot.

The calibration studies began with generation and purification of recombinant CFP and YFP proteins in bacteria (Fig. 2 A). To determine protein concentration several ways, we measured the absorbance (A_{280}) of tyrosine and tryptophan residues at 280 nm, absorbance of the fluorochromes at their peaks (440 nm for CFP and 505 nm for YFP), and absorbance of the alkali-denatured fluorochrome of YFP (Fig. 2, B and C). From published extinction coefficients, these values corresponded to $\sim 650 \mu\text{mol}$ of CFP and $400 \mu\text{mol}$ of YFP protein per liter of stock solution (Table I). We then compared the calibrated solutions to suspensions of fluorescent beads (cyan fluorescent [FSc] and yellow fluorescent [FSy]) in cuvettes using a fluorometer (Fig. 2, D and E). One

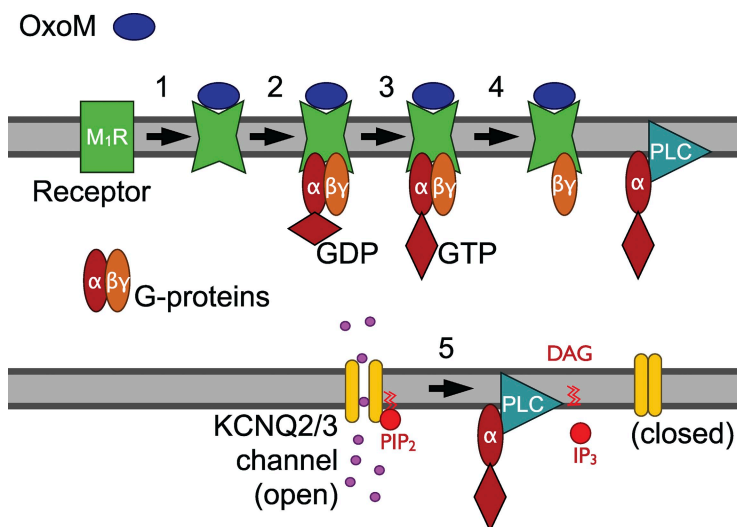


Figure 1. Schematic representation of classical M_1R signaling. (1) Binding of the agonist Oxonolone to the M_1R increases receptor affinity for G proteins. (2) Binding of G proteins to the receptor leads to NX (3) on the G_{α_q} subunit. (4) G protein $\alpha\beta\gamma$ trimers dissociate, and G_{α_q} binds to PLC. (5) KCNQ2/3 potassium channels require PIP_2 to open. Activated PLC cleaves PIP_2 into DAG and IP_3 . PIP_2 depletion turns off KCNQ2/3 channels.

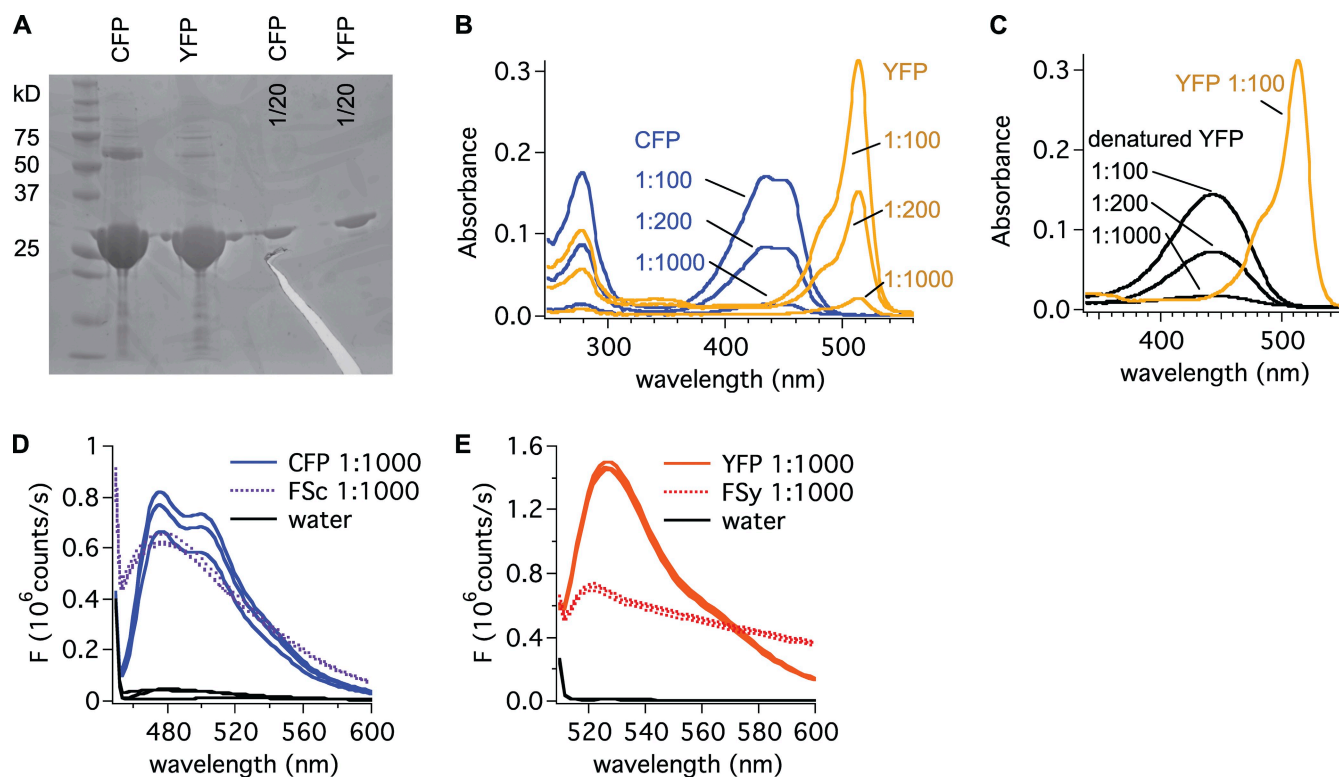


Figure 2. Properties of purified recombinant CFP and YFP proteins. (A) Purity of protein solutions is >95%: 20 or 1 μg (1/20) of purified protein (CFP and YFP) was separated by SDS-PAGE and stained with Coomassie. The thick band above 25 kD corresponds to the molecular weight of CFP and YFP (27 kD). Impurities are less abundant than the 1/20 dilution. Upper band for CFP possibly reflects dimerization. (B) Absorbance spectra for CFP and YFP. CFP and YFP solutions were diluted at 1:100, 1:200, and 1:1,000, and absorbance was determined (1) at the peak around 280 nm (absorbance of tyrosine and tryptophan) and (2) at the peak of fluorophore absorbance (440 nm for CFP and 515 nm for YFP). Results are summarized in Table I. (C) Absorbance spectra for YFP denatured in 0.1 N NaOH as compared with native YFP. (D) Emission spectra for diluted CFP solution (CFP; 3.9×10^{11} molecules/ μl) and a suspension of FSc beads (FSc; 10^4 beads/ μl) suggest that 1 FSc bead is as bright as 4×10^7 CFP molecules. Three different samples measured in the same experiment are shown. Similar results were found in three independent experiments. (E) Emission spectra from diluted YFP solution (YFP; 2.4×10^{11} molecules/ μl) and a suspension of FSy beads (FSy; 10^4 beads/ μl) indicate that 1 FSy bead is as bright as 10^7 YFP molecules. Similar results were found in three independent experiments.

FSc bead was as bright as 4×10^7 CFP molecules, and one FSy bead was bright as 10^7 YFP molecules. In the previous FRET work, we had selected cells to study by quantitative criteria based on their brightness (Jensen

TABLE I
Absorbance of CFP and YFP solutions

	ϵ	A	C
	mM^{-1}		μM
CFP, 280 nm	25.9 ^a	16.7 \pm 0.7	646 \pm 27
CFP, 440 nm	26 ^b	16.0 \pm 0.9	617 \pm 33
YFP, 280 nm	23.4 ^a	10.8 \pm 0.3	460 \pm 11
YFP, 515 nm	84 ^b	30.5 \pm 0.4	363 \pm 5
YFP, denatured	44 ^c	13.8 \pm 1.3	313 \pm 31

Measurements on three different days, mean \pm SEM. ϵ , published extinction coefficients; A, measured absorbance of the stock solution; C, protein concentration of the stock solution estimated as A/ϵ .

^aCalculated from molar extinction coefficients for tyrosine (1.49 mM^{-1}) and tryptophan (5.5 mM^{-1} ; from Oregon Medical Laser Center).

^bFrom Patterson et al. (2001).

^cFrom Ward (2006).

et al., 2009). Using the same epifluorescence photometry, we compared the fluorescence of cells we would select for those experiments to the calibrated FSc and FSy beads. Note that beads were calibrated against CFP or YFP in aqueous solution, but fusion to other proteins and the cellular environment can alter the quantum efficiency. We did not correct for such changes. Pooled across all transfections, the cells were on average 22% as bright as an FSc bead in the CFP channel (Fig. 3 A) and 88% as bright as an FSy bead in the YFP channel (Fig. 3 B), corresponding to roughly 10^7 CFP and 10^7 YFP molecules per typical cell. Our estimated numbers show internal consistency in that for the receptor construct that had stoichiometric amounts of YFP and CFP covalently linked, the calculated fluorophore ratio was 1.1 ± 0.1 ($n = 7$ cells).

PH probes are CFP-tagged or YFP-tagged PH domains from PLC δ 1 that bind PIP $_2$. In confocal images, the fluorescence of cells transfected with PH probes is brightest at the plasma membrane. When PIP $_2$ is depleted, PH probes leave the membrane and the cytosolic fluorescence

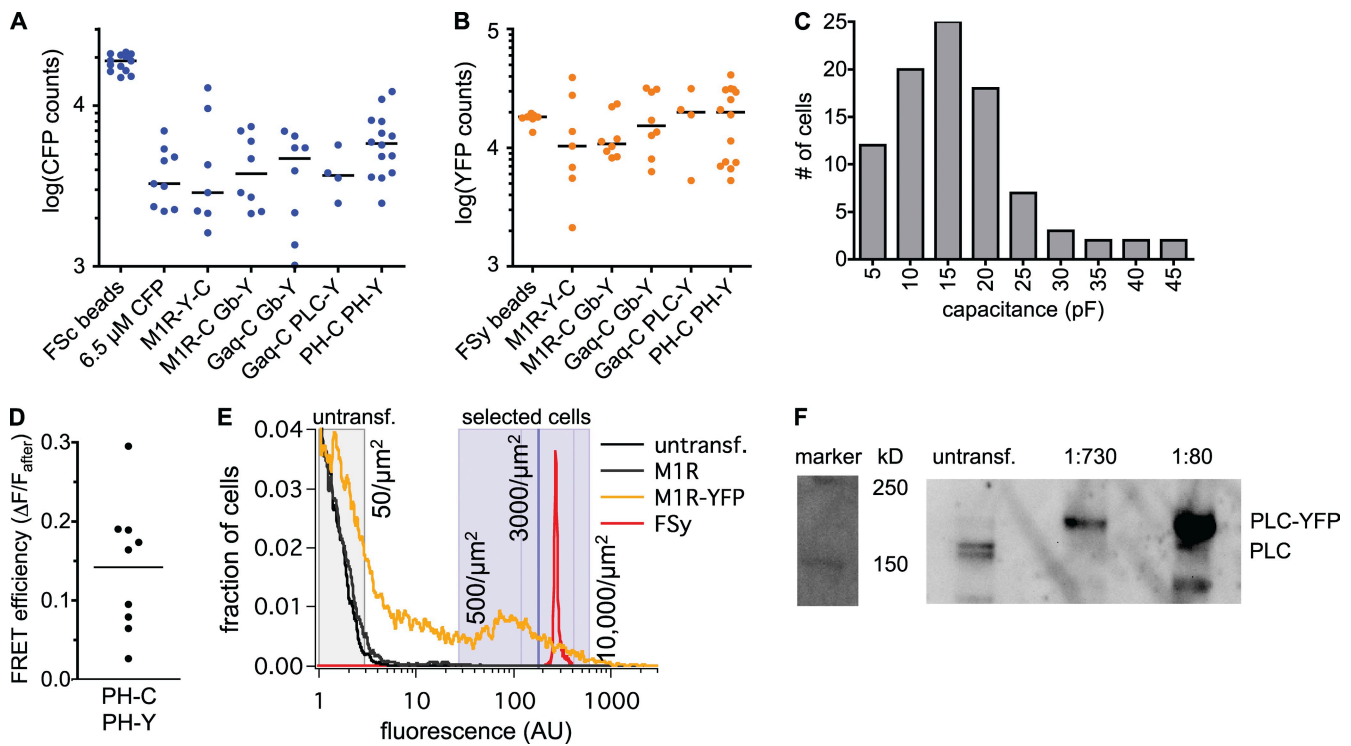


Figure 3. Determining expression levels. (A) Fluorescence counts from epifluorescence microscopy with 440 nm excitation and 480 nm emission (CFP channel) of: FSc beads (FSc), cells equilibrated with a 6.5- μM CFP solution by whole cell patch clamp (6.5 μM CFP), and selected cells transfected with various FRET constructs after bleaching of YFP. Constructs: intramolecular FRET probe for M₁R activation (M₁R-Y-C); M₁R-CFP, G β -YFP, G α , and G γ (M₁R-C Gb-Y); G α_q -CFP, G β -YFP, M₁R, and G γ (Gaq-C Gb-Y); G α_q -CFP, PLC-YFP, M₁R, G β , and G γ (Gaq-C PLC-Y); PH-PLC δ 1-CFP, PH-PLC δ 1-YFP, and M₁R (PH-C PH-Y). (B) Epifluorescence counts with 500 nm excitation and 535 nm emission (YFP channel) for FSc beads (FSc) and the same transfected cells as in A before the bleaching of YFP. (C) Distribution of cell capacitance from whole cell patch clamp recordings (93 cells). (D) FRET efficiency estimated from donor dequenching by acceptor photobleaching. The calculation uses the fractional increase (ΔF) in CFP fluorescence after the bleaching of YFP and the final fluorescence F_{after} . (E) Flow cytometry of untransfected cells (black), of cells transfected with M₁R, G α , G β , and G γ (gray), or M₁R-YFP, G α , G β , and G γ (yellow), and of fluorescent beads (FSy; red). Excitation was 488 nm, and emission was 530/30 nm. Fluorescence intensity histograms were normalized to the maximum, which was in the first bin (offscale). Shaded gray area contains 99% of untransfected cells. Shaded blue area is the brightness range of cells selected for experiments (see B), as estimated from comparison to FSc beads. Dark blue line represents the median intensity of these cells, and lighter blue lines represent the 25th and 75th percentiles. Three independent experiments gave similar histograms. (F) Western blot of lysates from untransfected cells and from cells transfected with PLC-YFP at 1:730 and 1:80 dilutions. Primary antibody against PLC- β 1. Two bands in untransfected cells around the 150-kD marker correspond to PLC- β 1a (top; 138 kD) and PLC- β 1b (bottom; 133 kD, nuclear localization). PLC-YFP is expected to run 27 kD higher. Endogenous PLC is comparable in intensity to the 1:730 dilution of transfected cells. Similar results were found in four independent experiments.

increases two- to threefold (van der Wal et al., 2001; Horowitz et al., 2005; Winks et al., 2005). This suggests that 50–60% of the PH probe molecules are at the plasma membrane at rest. As the appearance of cells transfected with the other fluorescently tagged proteins (M₁R, G α_q , G β , and PLC) was similar to cells transfected with PH probes, and in the absence of a better estimate, 50% plasma membrane localization was assumed for all of these proteins. The median surface area of our cells was 1,500 μm^2 judging from the membrane capacitance (median, 15 pF; Fig. 3 C). Collectively, these numbers give a surface density of 3,200 expressed PH probes per μm^2 (50% of the 10^7 molecules per 1,500 μm^2 cell). Because we selected cells using the same brightness criteria for each of the transfected probes we studied, we now assume that the average density of any of them was

around 3,200 μm^{-2} in the subset of transfected cells we studied in Jensen et al. (2009).

As an independent check on these numbers, we equilibrated untransfected cells with a solution of purified CFP in the patch pipette chosen to give a similar brightness. The matching test solution contained 6.5 μM CFP (Fig. 3 A). Lacking a PH domain, these CFP molecules would all remain in the cytoplasm, but if 50% of them had translocated to the plasma membrane, they would have reached a surface density of 3,250 μm^{-2} , as estimated above, assuming a surface to volume ratio of 0.6 μm^{-1} . This factor is used in all places where we compare surface densities with cytoplasmic concentrations for our cells.

In principle, FRET efficiency can be used to estimate intermolecular distances. If donors and acceptors were

placed randomly at the plasma membrane, FRET efficiency would depend only on the density of acceptor molecules within the expected range of mean densities (Fung and Stryer, 1978). The approximately linear theoretical relationship has an initial slope of 8.3% FRET efficiency per 1,000 acceptors per μm^2 . (These numbers assume CFP/YFP fluorophores. A graph of the theoretical relationship is shown in Fig. S3 B of our companion paper by Falkenburger et al., 2010). In the PH-CFP- and PH-YFP-transfected cells we selected, the median resting FRET efficiency as determined by acceptor photobleaching was $14 \pm 3\%$ (Fig. 3 D). This corresponds to 1,700 acceptor PH-YFP molecules per μm^2 , smaller than the previous values but in a similar range.

From the small fraction of bright cells in the epifluorescence microscope, we had initially supposed that the efficiency of our transfections was low (Jensen et al., 2009). However, experiments with flow cytometry on the entire population of cells showed that many more cells were expressing fluorescent proteins but at levels well below our selection criteria. Comparison with FSy beads calibrated the cytometer readings. Untransfected cells and cells transfected with untagged M_1R s had low background fluorescence equivalent to $<1.7 \times 10^5$ YFP per cell or 50 YFP proteins per μm^2 (Fig. 3 E). Transfection with M_1R -YFP increased the fluorescence of most cells, but the values ranged over three orders of magnitude. In summary, the brightness range of cells selected for FRET experiments corresponded to a range of 500–10,000 fluorescent proteins per μm^2 .

Based on the brightness distribution in flow cytometry, we can calculate the mean density of fluorescent

proteins if we picked cells at random in a dish of transfected cells. The arithmetic mean of the brightness of all cells was 28 arbitrary fluorescence units with M_1R -YFP and 61 for PLC-YFP in transfections together with unlabeled G proteins (median of three independent experiments). This corresponds to 500 molecules μm^{-2} for M_1R -YFP and 1,000 for PLC-YFP.

Endogenous levels of PLC are much lower

To obtain an estimate of endogenous levels of PLC, we compared PLC levels in untransfected cells with PLC-YFP-transfected cells by Western blot (Fig. 3 F). Protein lysates from PLC-YFP-transfected cells had to be diluted 600–700 times to produce immunostaining that was similar in intensity to endogenous PLC. Whereas a random cell in a dish transfected with PLC-YFP expresses on average 1,000 μm^{-2} PLCs at the plasma membrane (from flow cytometry; plus an equal number in the cytosol), a native tsA cell might express only around 3 μm^{-2} . Thus, when we transfect with fluorescent PLC, our selection criteria pick cells with $\gg 100$ -fold increased expression of PLC $\beta 1$, which explains why PLC transfection greatly speeds muscarinic suppression of KCNQ current compared with control cells (Jensen et al., 2009). Note that the Western blot determined only PLC $\beta 1$, and there may be other PLC subtypes also contributing to the G_q -coupled inhibition of KCNQ2/3 current. In an alternative approach below, we suggest from modeling that assuming 10 μm^{-2} PLC in native cells would be appropriate to describe the Oxo-M concentration–response relationship measured for inhibition of KCNQ2/3 current in cells transfected with only

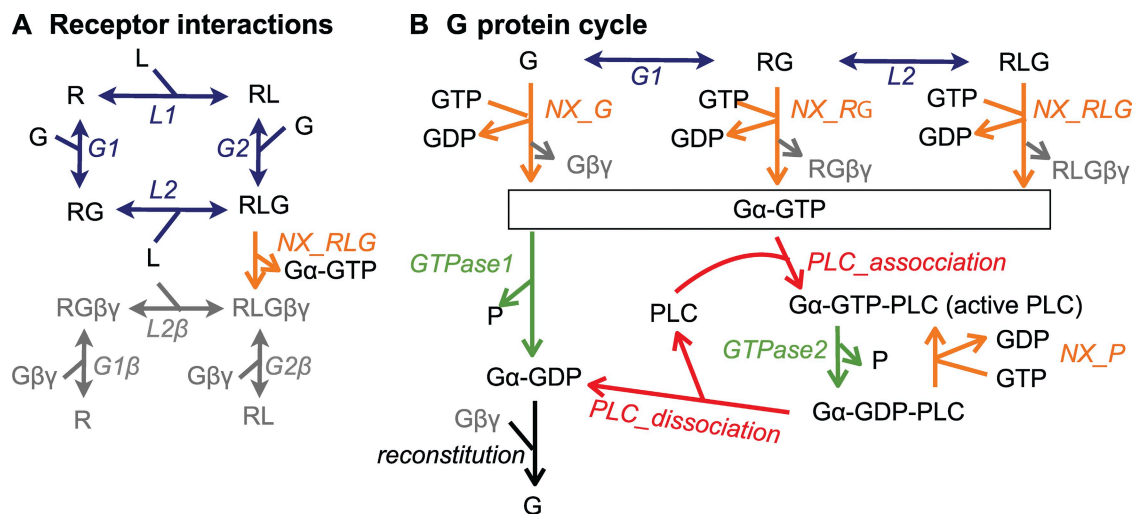


Figure 4. Outline of the kinetic model. Model species are depicted with Roman letters, and reactions are in italics. For reversible reactions (double arrows), forward reactions are in the direction of association. Note that most species and some reactions occur in several places in the schema. (A) Binding of agonist ligand (L) and G protein heterotrimer (G) to the M_1R (R) are implemented as an allosteric (ternary complex) model (blue), where ligand binding increases the affinity for G proteins and G protein binding increases the affinity for ligand. It is assumed that after NX, $G\alpha_q$ -GTP dissociates from the receptor, leaving behind $G\beta\gamma$ (gray). (B) G protein activation by NX (includes G protein dissociation; orange) and G protein deactivation by GTPase activity (green). Activated G proteins ($G\alpha$ -GTP) deactivate slowly through their own GTPase activity ($GTPase1$) or much faster ($GTPase2$) when bound to PLC.

untagged M₁R and KCNQ2/3 channel subunits (Jensen et al., 2009).

Modeling muscarinic signaling to G proteins and PLC

We then could develop a kinetic model of the entire M₁R signaling cascade. The model is outlined in Fig. 4. The signaling cascade is confined to the plasma membrane, and proteins are assumed to be at uniform density and freely diffusing. Tables II–IV list initial conditions, differential equations, and rate constants. The first part is an allosteric model for the binding of ligand (L) and G proteins (G) to the receptor (R) (Fig. 4 A). L and G bind to R to make the ternary complex RLG, which catalyzes formation of dissociated, active G α_q -GTP (Fig. 4, A and B). This active moiety then binds to and activates PLC until the GTPase activity of G α_q restores the inactive G α_q -GDP (Fig. 4 B). The model is constrained by time courses and concentration–response curves of previously published FRET data (Jensen et al., 2009). Modeling hydrolysis of PIP₂ by active PLC, PIP₂ synthesis, and PIP₂ effects on KCNQ2/3 current is described in our companion paper (Falkenburger et al., 2010).

Our model of ligand binding to M₁R can be called a ternary complex model, but it does not distinguish between “activated” and “resting” forms of the receptor because our FRET data (Jensen et al., 2009) do not provide information about this distinction. In this regard it contrasts with cubic ternary complex models where the distinction is made (Weiss et al., 1996; Kinzer-Ursem and Linderman, 2007). Our experimental data also cannot discriminate NX from G protein dissociation; therefore, we lumped these steps in a single, first-order reaction. Some studies suggest that certain receptors and G proteins are preassembled even without agonist (for review see Hein and Bünemann, 2009). The observed agonist-

induced increase in M₁R/G β FRET could then reflect a conformational change in such preassembled complexes rather than de novo binding of G proteins to the receptor. Similarly, the decrease in G α /G β FRET could reflect a conformational change in the G protein heterotrimer rather than its dissociation. We did find it possible to construct an alternative model that uses conformational change instead of G protein binding and dissociation that reproduces our experimental results equally well. We have chosen the implementation of Fig. 4 as it relates to previous (classical) models and reflects a simple interpretation of our FRET findings. For the remainder of this paper, we continue with this binding/dissociation framework for clarity. However, our data do not allow us to favor this implementation over that of a preassembled complex.

Further, the model does not include desensitization of receptors (phosphorylation and/or internalization), which is more prominent in the termination of activity of some other GPCRs (Maeda et al., 2003; Gainetdinov et al., 2004). It also does not consider the dependence of PLC on cytosolic calcium, which constitutes a highly nonlinear positive feedback loop that is not well constrained by our experiments to date.

The initial conditions are taken from the measurements and estimates of protein amounts given in the previous sections. In addition to initial conditions, the model has five free parameters for binding of ligand

TABLE II
Initial conditions

Species	Density	Source
	μm^{-2}	
R (endogenous)	1	From ratio of endogenous R to G
R (M ₁ R-YFP)	3,000	From fluorescence
R (M ₁ R, selected for FRET)	3,000	To reproduce G α /G β FRET onset
R (M ₁ R, current measurements)	500	Arithmetic mean of all cells (flow cytometry)
G protein (endogenous)	40	To fit concentration-response curve of current
G protein (overexpressed)	3,000	From fluorescence
PLC β 1 (endogenous)	3	From western blot
PLC β 1 (overexpressed)	3,000	From fluorescence
PLC (total endogenous)	10	To fit concentration-response curve of current
PH domains (membrane)	3,000	From fluorescence, consistent with FRET

TABLE III
Differential equations

Reaction	Rate equation
Blue ^a	
L1	$k_f_{L1} * R * L - k_r_{L1} * RL$
L2	$k_f_{L2} * L * RG - k_r_{L2} * RLG$
G1	$k_f_{G1} * G * R - k_r_{G1} * RG$
G2	$k_f_{G2} * G * RL - k_r_{G2} * RLG$
Grey ^a	
G1 β	$k_f_{G1} * G\beta\gamma * R - k_r_{G1} * RG\beta\gamma$
G2 β	$k_f_{G2} * G\beta\gamma * RL - k_r_{G2} * RLG\beta\gamma$
L2 β	$k_f_{L2} * L * RG\beta\gamma - k_r_{L2} * RLG\beta\gamma$
Orange ^a	
NX_RLG	$k_{NX_RLG} * RLG$
NX_G	$k_{NX_G} * G$
NX_RG	$k_{NX_G} * RG$
NX_P	$k_{NX_P} * G\alpha\text{-GDP-PLC}$
Green ^a	
GTPase1	$k_{GTPase1} * G\alpha\text{-GTP}$
GTPase2	$k_{GTPase2} * G\alpha\text{-GTP-PLC}$
Red ^a	
PLCassoc	$k_{PLCassoc} * G\alpha\text{-GTP} * PLC$
PLCdiss	$k_{PLCdiss} * G\alpha\text{-GDP-PLC}$
Black ^a	
G-reconstitution	$k_{reconst} * G\beta * G\alpha\text{-GDP}$

Units of fluxes (reaction rates) are molecules $\mu\text{m}^{-2} \text{s}^{-1}$.

^aColor of arrow in Fig. 4.

TABLE IV
Rate constants

Parameter	Value	Units	Comment
α	100		Allosteric constant (cooperativity factor); Cortés and Palacios (1986); Vannucchi and Goldman-Rakic (1991)
KL1	2	μM	KL2 * α , similar to EC50 for M ₁ R-Y-C FRET
KL2	0.02	μM	From EC50 for Oxo-M on M ₁ R/G β FRET
KG1	25,500	μm^{-2}	α * KG2
KG2	255	μm^{-2}	kr_G2/kf_G2
kf_L1	2.8	$\mu\text{M}^{-1} \text{s}^{-1}$	kr_L1/KL1
kr_L1	5.5	s^{-1}	From M ₁ R-Y-C FRET recovery
kf_L2	2.8	$\mu\text{M}^{-1} \text{s}^{-1}$	As kf_L1
kr_L2	5.5×10^{-2}	s^{-1}	kf_L1 * KL2
kf_G1	2.7×10^{-4}	$\mu\text{m}^2 \text{s}^{-1}$	Not well-constrained
kr_G1	6.8	s^{-1}	kf_G1 * KG1
kf_G2	2.7×10^{-3}	$\mu\text{m}^2 \text{s}^{-1}$	From M ₁ R/G β FRET onset
kr_G2	0.68	s^{-1}	From M ₁ R/G β FRET recovery
k_NX_RLG	0.65	s^{-1}	From G α /G β FRET onset, fits G α /PLC FRET onset
k_NX_G	1.5×10^{-5}	s^{-1}	From Turcotte model, d1 ^a
k_NX_P	4.7	s^{-1}	From Turcotte model, d4 ^b
k_GTPase1	0.026	s^{-1}	From G α /G β FRET recovery
k_GTPase2	15.0	s^{-1}	From Turcotte model, p4 ^c
k_PLCassoc	1	$\mu\text{m}^2 \text{s}^{-1}$	Not rate-limiting
k_PLCdiss	0.71	s^{-1}	From G α -PLC FRET recovery
k_reconst	1	$\mu\text{m}^2 \text{s}^{-1}$	Reconstitution of G $\alpha\beta\gamma$ trimers, affects timing if <1.0

^ad1 refers to the dissociation of GDP from G α alone in Turcotte et al. (2008).

^bd4 refers to the dissociation of GDP from G α bound to RL and PLC.

^cp4 refers to GTP hydrolysis bound to RL and PLC.

and G proteins to receptor and eight parameters for the G protein cycle. These rate constants were chosen manually to reproduce the experimentally observed time courses (Jensen et al., 2009) for individual steps of the M₁R signaling cascade. Most rate constants are strongly constrained by at least one of the time courses (see comments in Table IV). The number of “hidden” steps in the model is therefore small. Some rate constants for NX and G α_q GTPase activity were also informed by biochemical measurements (Turcotte et al., 2008). Rate constants are compared with literature values in the Discussion.

The model can reproduce our FRET results. Fig. 5 (A and B) summarizes the observed onset and recovery time courses of the first four steps of muscarinic signaling after agonist application, as measured by FRET (measured symbols from Jensen et al., 2009). The steps are agonist binding to receptor, G protein binding to receptor, G α -G $\beta\gamma$ dissociation, and G α binding to PLC. Smooth curves from the model are drawn on logarithmic axes. In comparing model results to the FRET time courses, initial conditions differed between traces to accommodate the different transfections required to measure FRET as listed in the legend.

We must consider how the choice of M₁R density (initial condition of R) was made. For any overexpressed proteins that are fluorescently labeled and selected for,

we assumed a density of 3,000 μm^{-2} ; however, to measure G α /G β FRET we had transfected an unlabeled M₁R. Its mean density should be between 500 and 3,000 μm^{-2} , depending on whether its expression level is uncorrelated or identical to that of the fluorescently tagged G proteins. It is not unreasonable to assume some correlation between expression levels. For example, we find that when cells are transfected with GFP and unlabeled KCNQ2/3 channel subunits, most GFP⁺ cells also show KCNQ2/3 current. At the same time, we find under such conditions that a cell’s brightness correlates poorly with the current amplitude (unpublished data). The time course of dissociation of G α from G $\beta\gamma$ observed with FRET was better approximated with 3,000 μm^{-2} receptors (solid line in Fig. 5 A) than with 1,000 (dotted line in Fig. 5 A) or 500 μm^{-2} (not depicted). The model reproduced the time course of G α /PLC FRET reasonably using the same receptor density. The predicted onset of G protein dissociation has two components (dotted line in Fig. 5 A) if the number of receptors is much lower than the number of G proteins (i.e., not all G proteins are activated at once) and the deactivation of G proteins is much slower than their activation (see Fig. 5 B).

Fig. 6 shows predicted FRET time courses on an absolute scale and with a linear time axis (solid traces). In addition to time courses, we reproduced

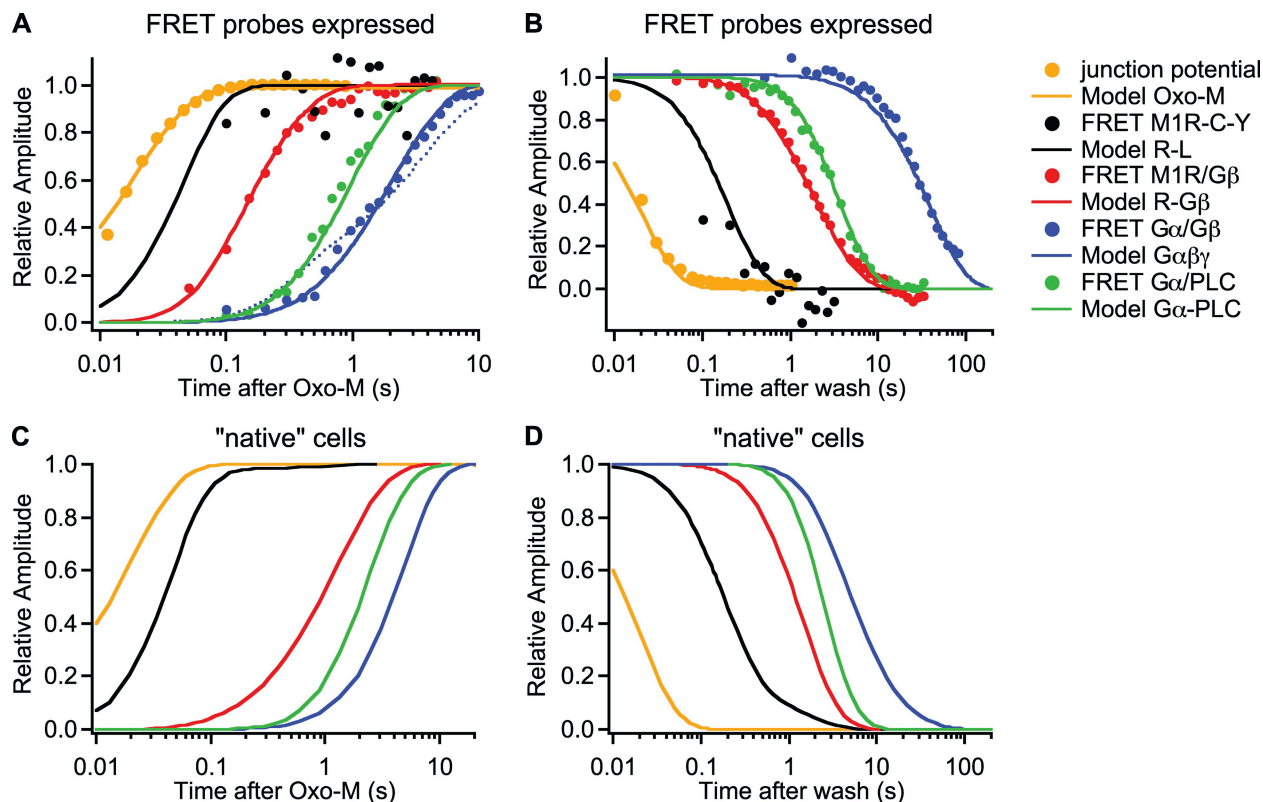


Figure 5. The model reproduces many individual steps of M₁R signaling. Rate constants for the kinetic model in Fig. 4 were chosen to mimic time courses of signaling steps previously measured by FRET (Jensen et al., 2009). Curves are model outputs, and circles are binned averages of FRET data from several experiments. Solution exchange was measured as the change in junction potential as 160 mM KCl is washed on. The exchange data are offset by 10 ms to the left. A and C show normalized onset time courses induced by the application of “saturating” M₁R agonist (10 μ M Oxo-M). The vertical axis is like a reaction coordinate from the baseline level (0.0) to the final level (1.0). A logarithmic time axis is used to display fast and slow events. B and D show recovery time courses. R-L is the sum of all species with R and L (RL, RLG, and RLG β); R-G β is the sum of all species with R and G β (RG, RLG, RLG β , and RG β); G α β γ is the sum of all species with undissociated G proteins (G, RG, and RLG); and G α -PLC is the sum of G α -GTP-PLC and G α -GDP-PLC. Note that the changes of G α /G β FRET and G α β γ model output are actually negative-going but become positive-going in the coordinates used. For the model outputs in A and B, amounts of receptors, G proteins, and PLC (in μm^{-2}) were chosen to reflect the conditions with overexpressed FRET probes: R, 3,000; G proteins, 0 (R-L) or 3,000 (R-G β , G α β γ , and G α -PLC); PLC, 10 (R-L, R-G β , and G α β γ) or 3,000 (G α -PLC). For model outputs in C and D, amounts were chosen as estimated for native (untransfected) cells: R, 1; G proteins, 40; PLC, 10 μm^{-2} . The dotted line in A shows the predicted onset of G α /G β FRET assuming 1,000 μm^{-2} receptors. Note the biphasic time course, which is unlike the measured data.

concentration–response data from the FRET measurements of Jensen et al. (2009). Fig. 7 overlays experimental Oxo-M concentration–response curves (symbols) with model curves (solid traces) for G protein binding to receptor (Fig. 7 A), G protein dissociation (Fig. 7 B), and G α_q binding to PLC (Fig. 7 C). Again, the match is generally reasonable.

As the initial conditions differed, the various FRET time courses could not be compared easily. To permit comparison, we used the model informed by the FRET measurements to calculate time courses for identical initial conditions. We used amounts of endogenous receptors, G proteins, and PLC estimated below (“native cells”; see Figs. 5, C and D, and 7 D). This also allowed us to ask how the FRET time courses we had measured with high amounts of receptors, G proteins, or PLC (Fig. 6, solid lines) would com-

pare with those with endogenous amounts (Fig. 6, dotted traces).

Endogenous amounts of PLC and G proteins were estimated by modeling the KCNQ2/3 current inhibition by M₁R activation seen in cells transfected only with unlabeled M₁R, KCNQ2/3 channel subunits and soluble GFP (Jensen et al., 2009; no explicit selection criteria concerning GFP brightness). We assumed 500 μm^{-2} receptors in these experiments (the arithmetic mean of the population) and 5,000 μm^{-2} PIP₂ molecules at rest (see Falkenburger et al., 2010). The observed steady-state block of KCNQ2/3 current by 0.1 and 10 μ M Oxo-M was 50 and 90% (Jensen et al., 2009). If KCNQ2/3 current was simply linear with PIP₂, steady-state PIP₂ densities would have to be 2,500 μm^{-2} with 0.1 μ M and 500 μm^{-2} with 10 μ M Oxo-M. PIP₂ metabolism and PIP₂ effects on KCNQ2/3 current were in fact modeled as

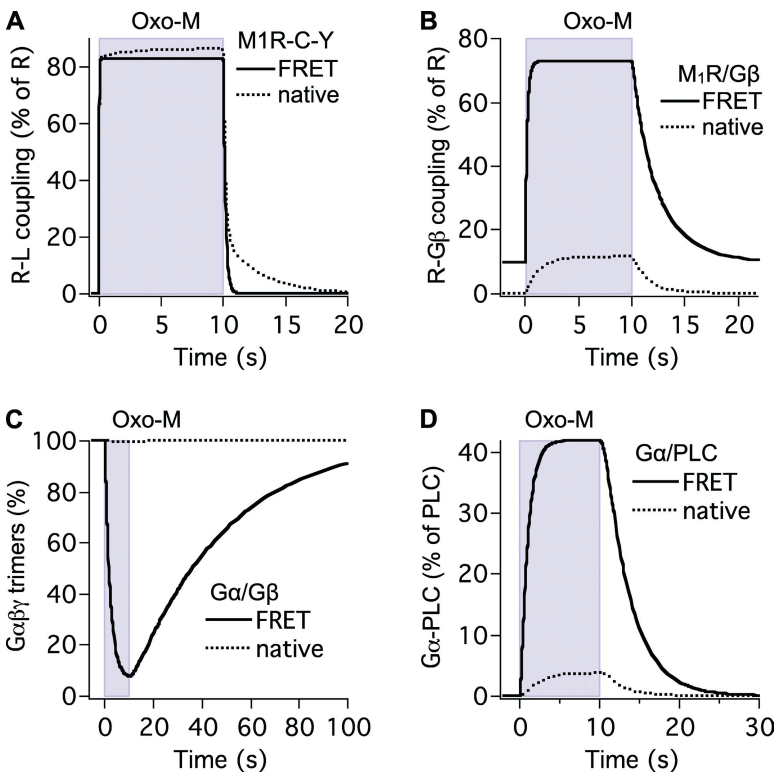


Figure 6. Comparison of model predictions for cells overexpressing FRET probes and for native cells. Solid traces are the time courses from Fig. 5 (A and B) on an absolute y axis and a linear time axis, and dotted traces are from Fig. 5 (C and D). Naming conventions are as in Fig. 5. Initial conditions for solid traces (in μm^{-2}): R, 3,000; G proteins, 0 (A) or 3,000 (B-D); PLC, 10 (A-C) or 3,000 (D). Initial conditions for dotted traces (in μm^{-2}): R, 1; G proteins, 40; PLC, 10.

described in our companion paper (Falkenburger et al., 2010), with roughly similar results (Fig. 8 A). Achieving these steady-state PIP_2 densities (2,500 and $500 \mu\text{m}^{-2}$) in the face of ongoing constant PIP_2 synthesis requires a fivefold higher activity of active PLC with $10 \mu\text{M}$ as compared with $0.1 \mu\text{M}$ Oxo-M. When we tried different assumed amounts of PLC (Fig. 8 B) and G proteins

(Fig. 8 C), we found that the combination of 10 PLC molecules μm^{-2} and 40 G proteins μm^{-2} produces fivefold more active PLC with $10 \mu\text{M}$ as compared with $0.1 \mu\text{M}$ Oxo-M (2.5 and $0.5 \text{ active PLC } \mu\text{m}^{-2}$). The assumed density of PLC molecules is only slightly higher than that obtained above by Western blot ($3 \mu\text{m}^{-2}$). An endogenous level of $40 \mu\text{m}^{-2}$ G proteins compares well to

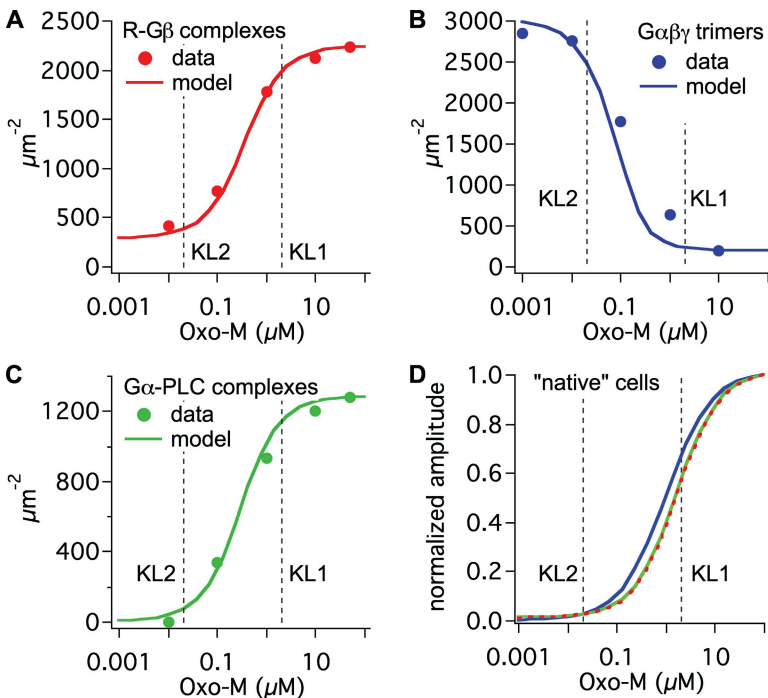


Figure 7. The model reproduces Oxo-M concentration-response curves. Maximum formation of R-Gβ complexes (A), maximum depletion of $\text{G}\alpha\beta\gamma$ trimers (B), and maximum formation of $\text{G}\alpha$ -PLC complexes (C) predicted by the model for different concentrations of Oxo-M (lines) were compared with averaged and normalized Oxo-M concentration data (symbols; from Jensen et al., 2009) for $\text{M}_1\text{R}/\text{G}\beta$ FRET (A), $\text{G}\alpha/\text{G}\beta$ FRET (B), and $\text{G}\alpha/\text{PLC}$ FRET (C). (D) Superimposed, normalized concentration-response curves predicted for maximum formation of R-Gβ complexes (red), maximum depletion of $\text{G}\alpha\beta\gamma$ trimers (blue; inverted), and maximum formation of $\text{G}\alpha$ -PLC complexes (green) predicted by the model for native cells. Initial conditions (in μm^{-2}): R, 1; G proteins, 40; PLC, 10. Vertical lines indicate dissociation constants KL1 and KL2 for receptors without and with G proteins bound.

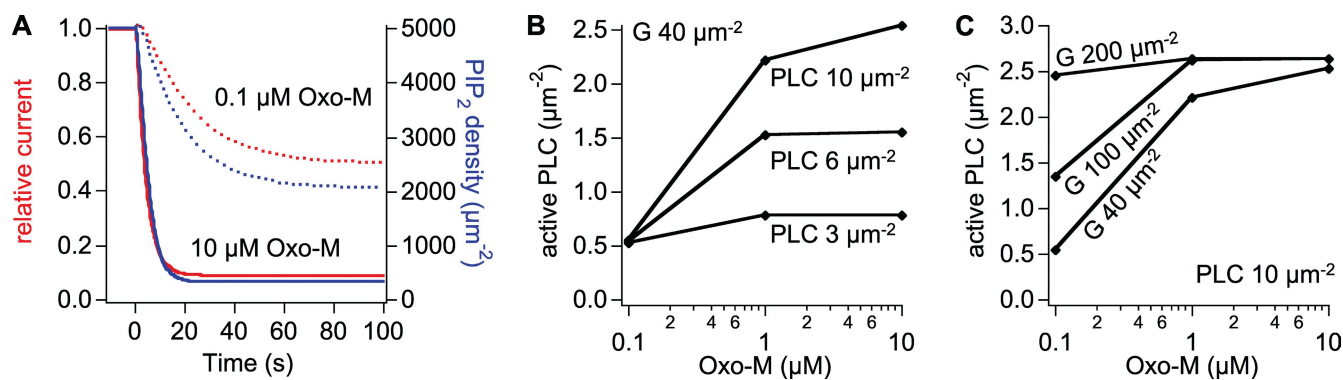


Figure 8. Estimating endogenous densities of PLC and G proteins. (A) Predicted effect of 10 and 0.1 μM Oxo-M on KCNQ current and plasma membrane PIP_2 . For reactions and parameters involving PIP_2 metabolism and KCNQ current see our companion paper (Falkenburger et al., 2010). (B and C) Predicted densities of active PLC for different initial amounts of PLC and G proteins. Unless otherwise noted, amounts were (in μm^{-2}): R, 500; G proteins, 40; PLC, 10.

66 and $466 \mu\text{m}^{-2}$ in other systems (10^5 and 7×10^5 per cell; Mahama and Linderman, 1994; Alt et al., 2001). The ratio of G proteins to receptors has previously been given as 20–50 (Luetje et al., 1987; Neubig, 1994; Alt et al., 2001). Although tsA201 cells do not express endogenous M_1 Rs, we accordingly approximated a pseudo-endogenous situation in the model with 1 receptor μm^{-2} and 40 G proteins μm^{-2} .

Comparing predicted time courses with the extrapolated “native” initial conditions revealed several interesting properties that are explored in more detail in the second half of the Discussion. (1) Modeling native tsA-like cells (Fig. 5 C) suggests a substantial delay between the binding of ligand and the binding of G proteins to the receptor. Such a “gap” is less apparent with overexpression of receptors and G proteins for FRET (Fig. 5 A). (2) G protein dissociation comes to steady state only after $G\alpha$ /PLC binding does. (3) The very slow reassociation of G proteins seen by FRET (Fig. 5 B, blue trace) is likely a consequence of the G protein overexpression, as it goes away in simulations with native conditions (Fig. 5 D). (4) Ligand binding in modeled native cells includes a slow component. This was not seen with the intramolecular FRET probe for ligand binding (M_1 R-YC) that does not couple to G proteins. (5) Ligand unbinding in modeled native cells is much slower than that seen with the overexpressed M_1 R-YC FRET probe (Fig. 6 A). (6) Modeled native cells have only 0.3% of receptors bound to $G\beta$ at rest, rising to 10% coupling during agonist. The fraction of G protein-bound receptors is much larger in cells transfected for M_1 R/ $G\beta$ FRET (Fig. 6 B). (7) Only a very small fraction of G proteins is dissociated (to $G\alpha$ and $G\beta\gamma$) by agonist in modeled native cells, whereas most of the G proteins are dissociated by agonist in cells transfected for $G\alpha$ / $G\beta$ FRET (Fig. 6 C). (8) Fewer than 5% of the PLC enzymes are activated by agonist in modeled native cells, more in cells transfected for $G\alpha$ /PLC FRET (Fig. 6 D). (9) For native cells, the model predicts similar Oxo-M EC_{50}

values for three measured interactions: receptor–G protein, G protein dissociation, and $G\alpha$ -PLC (Fig. 7 D). The EC_{50} values are also similar to KL1, the model’s dissociation constant for ligand binding to receptors without G proteins.

DISCUSSION

We have constructed a kinetic description of M_1 R signaling to PLC based on measurements of protein amounts and on time courses of individual signaling steps obtained from our earlier FRET measurements (Jensen et al., 2009). This model combines several concepts. (1) An allosteric approach provides for cooperative binding of ligand and G proteins to the receptor. Predicted time courses and EC_{50} values were similar to those measured by FRET. (2) Important rate constants for NX and GTPase activities were taken from a model of G protein signaling that had been refined by in vitro measurement of GTPase activity (Turcotte et al., 2008). Those measurements complement ours because FRET does not inform well about GTPase activities but does constrain rate constants for protein–protein interaction (binding of receptor and PLC to G proteins; see Fig. 3 in Turcotte et al., 2008). We first compare numerical values we have chosen with those reported for other GPCR systems in the literature. Then we discuss the implications of our model for GPCR signaling.

Protein amounts

When performing transient transfection, we often think of transfected and untransfected cells as two distinct populations of cells. Flow cytometry showed, however, that there is a continuous spectrum of transfection with fluorescence levels spanning three orders of magnitude, starting in a range that overlaps with untransfected cells (Fig. 3 E). Others made similar observations (e.g., He and Hristova, 2008). A local maximum in our fluorescence intensity histogram (Fig. 3 E) falls within the

range of selected cells and is close to our estimate of 3,000 molecules per μm^2 . One can characterize this subset of cells by a median (or modal) expression level. However, the expression level averaged over the entire population falls pretty much between the two modes of the distribution (which includes the obviously transfected cells on the right and cells that are indistinguishable from untransfected cells on the left). Thus, comparing transient expression levels in individual cells to measurements averaged from the entire population, such as radioligand binding or Western blot, is not straightforward.

Several independent estimates of protein expression levels agree that in the cells we selected, overexpressed proteins have densities around 3,000 μm^{-2} at the plasma membrane. (1) Comparisons of the fluorescence intensity of cells versus calibrated fluorescent beads suggest $\sim 3,000$ fluorescent proteins per μm^2 . (2) Filling untransfected cells with CFP by whole cell patch clamp yields a similar estimate. (3) The observed FRET efficiency of 14% for PH probes suggests a density of acceptors of 1,700 μm^{-2} . These values are in a range similar to the estimated density of expressed PH probes in N1E-115 cells (4,617 μm^{-2} ; Xu et al., 2003), but they are an order of magnitude higher than the copy numbers in identified synapses of scaffolding proteins fused to EGFP expressed in primary hippocampal neurons by adenoviral infection (Fig. 5 in Sugiyama et al., 2005). This is not unexpected as tsA201 cells were constructed for high expression of genes driven by the cytomegalovirus promoter we used.

As cells were selected based on their measured fluorescence, it is not surprising that the amounts of fluorescent proteins were similar across different cells and constructs. Our original selection of cells for the FRET experiments had been guided (1) on the lower end, by the sensitivity of our photometry apparatus to get adequate signal to noise ratio; (2) on the upper end, by the experience that very bright cells were often not as healthy or contained clusters of fluorescent proteins that do not participate in the response to agonist application; and (3) by an attempt to select a “typical” cell, i.e., one with a commonly observed level of expression. The resulting 3,000 proteins per μm^2 is a high number when compared with our estimate of endogenous PLC (3–10 μm^{-2}) and is comparable to our model estimate for free PIP_2 lipids (5,000 μm^{-2}). It is one to three orders of magnitude higher than typical ion channel densities (Hille, 2001). In cases where overexpressed proteins are supposed to report activities of native cells, it would be desirable to work with lower expression levels in the future. This would require some combination of more dilute transfection, stronger excitation light, better light collection and detection, longer integration of optical signals, and brighter and more bleach-resistant fluorophores.

We have quantified fluorescently tagged signaling molecules (receptors, G proteins, and PLC) mainly based on their fluorescence. Due to slow maturation of fluorescent proteins and cleavage, the number of fluorescent molecules need not be identical to the number of biologically active molecules. For PLC-YFP, we have evidence for $\sim 10\%$ cleavage products (Fig. 3 F), which is small compared with the overall variance of fluorescence intensity between individual selected cells (Fig. 3 A). In addition, the biological activity of PLC to speed up GTPase activity was similar between our estimates, calibrated with fluorescence intensity, and *in vivo* measurements where PLC amounts were determined biochemically (Turcotte et al., 2008).

The native cells in our model primarily represent the tsA201 cells we used for our experiments. Estimates of PLC and G proteins were based on Western blots for untransfected tsA201 cells and on KCNQ2/3 current reduction. Amounts are likely quite different in primary cells where signaling by G_q -coupled receptors is important. For example, a (locally) higher density of G proteins is one possible explanation for the higher fraction of high-affinity binding sites for Oxo-M in the brain as compared with tsA201 cells (see below).

Rate constants for ligand binding and unbinding, G protein activation, and deactivation

We now consider the rationale for the values selected for equilibrium and rate constants. The model uses equilibrium dissociation constants KL1 and KL2 for ligand binding to receptors (without and with G proteins, respectively) to mimic our concentration–response curves measured with FRET (Fig. 6, A–C) (Jensen et al., 2009). KL1 and KL2 are in the range of reported values for low- and high-affinity Oxo-M-binding sites in the brain (0.1 and 10 μM , Cortés and Palacios, 1986; 0.024 and 0.944 μM , Potter et al., 1988). The allosteric constant (also termed the cooperativity factor) is $\text{KL1}/\text{KL2} = 100$ (as in Cortés and Palacios, 1986; Vannucchi and Goldman-Rakic, 1991). The low-affinity equilibrium dissociation constant $\text{KL1} = 2 \mu\text{M}$ for ligand binding to receptor without G proteins is similar to our experimental K_D for Oxo-M binding to the intramolecular receptor FRET probe (M_1 R-Y-C), which does not couple to G proteins (4 μM ; Jensen et al., 2009).

The unbinding rate of Oxo-M from G protein-free receptor ($k_{r_L1} = 5.5 \text{ s}^{-1}$; $\tau_{\text{off}} = 182 \text{ ms}$) was set to match the decay of M_1 R-Y-C FRET. From KL1 and k_{r_L1} , we then calculated the ligand association rate constant k_{f_L1} as $2.8 \mu\text{M}^{-1}\text{s}^{-1}$. This is similar to the binding of the radioactive muscarinic antagonist quinuclidinyl benzilate to cultured heart cells ($3 \mu\text{M}^{-1}\text{s}^{-1}$; Galper and Smith, 1978), radioactive α_2 adrenergic agonist UK14304 to platelet membranes ($5 \mu\text{M}^{-1}\text{s}^{-1}$; Neubig et al., 1988), fluorescently tagged neurokinin to EGFP-NK2R ($5 \mu\text{M}^{-1}\text{s}^{-1}$; Palanche et al., 2001), and phenylisopropyl

adenosine to A₁-adenosine receptors (1 μM⁻¹s⁻¹; Lohse et al., 1984). It is faster than the estimated binding of the α₂ adrenergic antagonist yohimbine to platelet membranes (0.25 μM⁻¹s⁻¹; Neubig et al., 1988), and a little slower than the estimated association of *N*-formyl peptides to neutrophils (10 μM⁻¹s⁻¹, Waller et al., 2004; 20 μM⁻¹s⁻¹, Sklar, 1987). The resulting onset time constant for M₁R-Y-C FRET (40 ms) is in a range similar to that reported for ligand binding to α_{2A} or β₁ adrenergic receptors (40–60 ms; Vilardaga et al., 2003; Hein et al., 2005; Hoffmann et al., 2005; Rochais et al., 2007), but faster than for parathyroid receptors (1 s; Vilardaga et al., 2003). The forward rate constants for ligand binding to receptors are often fast and relatively insensitive to the ligand structure, whereas the off rate constants vary and give different ligands different affinities. For example, a series of *N*-formyl ligands differed more in their predicted unbinding than in their binding rate constant (Waller et al., 2004). Therefore, we set $k_{f_L2} = k_{f_L1}$. FRET within M₁R-Y-C in fact reports the conformational change after ligand binding, not the actual binding itself. The fact that this event falls in the same time range as ligand binding, as reported by fluorescent or radioactive ligands, suggests that the conformational change after ligand binding is not a slow step.

The slower rate constant for unbinding of ligand from G protein-bound receptor ($k_{r_L2} = 0.055 \text{ s}^{-1}$; $\tau_{\text{off}} = 18 \text{ s}$) derives from k_{f_L2} and $K_{L2} = 20 \text{ nM}$ (see Table II). This rate constant is similar to that determined for unbinding of radioactive Oxo-M from G protein-bound M₂ receptors (0.01 s⁻¹, Hirschberg and Schimerlik, 1994; $K_d = 1.3 \text{ nM}$). As the unbinding reaction reflects affinity, it is slower than the unbinding of IgG from FcR (2 s⁻¹, Lieto et al., 2003; $K_d = 2.4 \text{ μM}$), but faster than the unbinding of quinuclidinyl benzilate (0.005 s⁻¹, Galper and Smith, 1978; $K_d = 1.8 \text{ nM}$), UK14304 (0.0065 s⁻¹, Neubig et al., 1988; $K_d = 1 \text{ nM}$), yohimbine (0.001 s⁻¹, Neubig et al., 1988; $K_d = 4.8 \text{ nM}$), *N*-formyl peptides (0.001 s⁻¹, Waller et al., 2004; $K_{dS} = 0.04, 0.4, \text{ and } 4.6 \text{ nM}$), or erythropoietin (0.005 s⁻¹, Jarsch et al., 2008; $K_d = 2.9 \text{ nM}$) from their receptors.

Rate constants for receptor binding and unbinding of G proteins were determined from the time course of M₁R/Gβ FRET. (Again, in an alternative interpretation, this transition could represent a conformational change rather than de novo binding.) Because Oxo-M binds faster to the receptor than G proteins do, and 99.4% of receptors have ligand bound in the presence of 10 μM Oxo-M, the onset time course of M₁R/Gβ FRET (0.2 s) primarily constrained the rate constant k_{f_G2} (as opposed to k_{f_G1}). Similarly, the recovery time course of M₁R-Gβ FRET (3.7 s) mainly constrained the rate constant k_{r_G2} . K_{G2} was set to k_{r_G2}/k_{f_G2} , and K_{G1} was set to $\alpha * K_{G2}$. As the reaction *G1* does not contribute much to our measured time courses, *G1* rate constants are not further constrained by our measurements.

To avoid extreme values, we made them similar to the *G2* rate constants.

The choice of model rate constants for NX and G protein GTPase activity (Fig. 4 B) was guided both by our prior FRET measurements and by values from biochemical measurements of GTPase activity (Turcotte et al., 2008). These two sources dovetail nicely. Our rate of NX at ligand-bound receptor (NX_RLG), informed by the onset of Gα/Gβ FRET, was 0.65 s⁻¹, which compares to 0.4 s⁻¹ in Biddlecome et al. (1996) and to 0.28 s⁻¹ for the rate-limiting step in Turcotte et al. (2008). The rate constant for the slow endogenous GTPase activity of Gα_q (0.03 s⁻¹), chosen to reproduce recovery of Gα_q/Gβ FRET, was similar to values obtained in vitro (0.01 s⁻¹, Mukhopadhyay and Ross, 1999; 0.018 s⁻¹ for step p1 and 0.0063 s⁻¹ for p3 in Turcotte et al., 2008; 0.013 for p1 and p3 in Bornheimer et al., 2004, where steps p1 and p3 refer to the GTPase activity of Gα_q in the absence and presence of receptor ligand).

Interpretation of the model: binding of ligand and G proteins to receptors

Our model suggests that ligand binding occurs mainly through reaction *L1* (R + L). Binding of G proteins to the receptor occurs mainly through *G2* (RL + G). It is much slower than the binding of ligand (compare k_{f_L1} and k_{f_G2} in Table IV). The slow G protein binding is responsible for the majority of the delay between ligand binding and later steps (black trace to red trace in Fig. 5 C). The binding of G proteins to receptors was accelerated by overexpression of receptors and G proteins, making this delay less apparent with FRET experiments (Fig. 5 A). Slow G protein binding also is responsible for a slower component in ligand binding (dotted trace in Fig. 6 A) that has been described with radioligands (e.g., Neubig et al., 1988; Lohse et al., 2008) but was not observed with our intramolecular receptor FRET probe (M₁R-Y-C), presumably because it does not interact with G proteins.

With 10 μM Oxo-M and “endogenous” amounts of receptor, G proteins, and PLC, our model predicts the following distribution of receptors: 74% RL, 15% R alone, and 11% RLG and RLGβ combined. The unbinding of ligand then occurs mainly from RL through the reversal of *L1*, producing the fast initial decrease in R-L complexes (Fig. 6 A). Ligand unbinding from RLG complexes occurs through the reversal of first *G2* and then *L1* (k_{r_L2} is slower than k_{r_G2}), making slow unbinding of G proteins from RLG (k_{r_G2}) rate limiting for a portion of ligand unbinding. This results in the slower component of ligand unbinding in native cells (Fig. 6 A, dotted trace). The overall speed of ligand unbinding thus depends on the balance of RL versus RLG. It is faster with endogenous amounts (mainly RL) and slower with overexpression of G proteins (more RLG).

One long-standing question about the signaling of receptors and G proteins is to what extent G proteins are complexed with receptors at rest (“preassembly”). Previous measures of preassembly by FRET have yielded mixed results (see Lohse et al., 2008; Hein and Bünemann, 2009). In our own data, the nonzero resting $M_1R/G\beta$ FRET might suggest preassembly. However, expression levels of M_1R -CFP and $G\beta$ -YFP were similar to those for PH-CFP and PH-YFP (Fig. 3, A and B), which already achieve 14% FRET efficiency (Fig. 3 D) simply from crowding at the membrane. We therefore have to assume that a substantial proportion of $M_1R/G\beta$ resting FRET also arises from crowding after overexpression and not from specific binding to one another. FRET efficiency for $M_1R/G\beta$ was not much higher than for PH probes, so that we too have no clear evidence for preassembly.

The time course of $M_1R/G\beta$ FRET changes contains additional information about steady-state receptor G protein binding that is independent from resting FRET levels. Our model suggests that with overexpression of receptor and G proteins, 10% of receptors have G proteins bound at rest, a number that rises to 73% in the presence of saturating Oxo-M (Fig. 6 B). The fraction of receptor–G protein complexes in the presence of Oxo-M in the model is determined by the equilibrium dissociation constant KG_2 , which is well constrained by the time course of $M_1R/G\beta$ FRET as outlined earlier. The extent of receptor–G protein binding at rest is determined by $KG_1 = \alpha * KG_2$, where α is an allosteric constant (Table IV). The value for α was taken from the literature. The prediction of the model, that only a small fraction (10%) of G proteins are coupled to receptors at rest when both are overexpressed, is in line with observations by others on the mobility of transfected fluorescently tagged G proteins, assayed by fluorescence recovery after photobleaching. For example, Qin et al. (2008) reported that immobilizing α_2 adrenergic receptors in HEK293 cells did not lower measured G protein mobility.

In the model, the fraction of receptors with G proteins bound at rest depends on the density of G proteins (but not on the density of receptors). In cells transfected with receptors only, the much weaker inhibition of KCNQ current by 0.1 μ M Oxo-M as compared with 10 μ M is best reproduced with a low number of G proteins (40 μ m⁻²; Fig. 8 C), i.e., many fewer G proteins than receptors. In that case, very few receptors can be G protein bound, and ligand would see only low-affinity binding sites (receptors without G protein). This is consistent with our radioligand binding studies of M_1R (Fig. S1 in Jensen et al., 2009), where we did not see evidence for high-affinity binding sites in cells transfected with M_1R or M_1R -YFP. Then, the EC50 values for the formation of receptor–G protein complexes, for G protein dissociation, and for $G\alpha$ -PLC binding (Fig. 7) would

also reflect low affinity, KL1. In contrast, with overexpression of G proteins for FRET measurements, high-affinity binding sites become more relevant and the EC50 values fall between KL1 and KL2.

Interpretation of the model: G protein activation and deactivation

Our FRET measurements showed a substantial gap between the change in $M_1R/G\beta$ FRET (red in Fig. 5 A) and the change in $G\alpha/G\beta$ FRET (blue in Fig. 5 A). In the model, this delay results from two factors. (1) The underlying reactions are sequential so G proteins have to be bound to receptors before meaningful NX and G protein dissociation can take place. (2) In agreement with the literature (Biddlecome et al., 1996; Turcotte et al., 2008), our time constant for the second reaction (NX) is >1 s, whereas that for the first reaction (G–R interaction) is 20-fold shorter.

Less intuitive is why $G\alpha$ binding to PLC (green in Fig. 5) reaches steady state before the preceding G protein dissociation does (blue in Fig. 5), both in native and transfected cells. The answer is that when displayed on an absolute scale, G protein dissociation and $G\alpha$ binding to PLC progress together, but even after PLC binding is saturated, dissociation of more G protein continues. Therefore, the time to steady state is longer for G protein dissociation than for PLC binding.

In our model, the NX activity and the GTPase activity are both ≥ 500 times faster for PLC-bound $G\alpha_q$ than for PLC-free $G\alpha_q$, as in the Turcotte et al. (2008) model. Compare, for example, NX rate constant NX_P with NX_G and GTPase rate constant $GTPase_2$ with $GTPase_1$ in Table IV. The chosen values make PLC simultaneously a strong guanine NX factor (GEF) (Turcotte et al., 2008) and a strong GTPase accelerating protein (GAP) (Biddlecome et al., 1996).

Our model of native cells reveals that in the steady state during Oxo-M, NX at RLG (NX_{RLG}) is compensated by an equal production of new RLG through reaction G2. Hence, the reactions NX_{RLG} , G2 β , and G2 create the cycle: $RLG \rightarrow RLG\beta\gamma \rightarrow RL \rightarrow RLG$, etc. (see Fig. 4 A). Of the $G\alpha$ -GTP formed, 99% binds to PLC, which accelerates GTP hydrolysis (GAP activity). However, 87% of $G\alpha$ -GDP-PLC formed then becomes $G\alpha$ -GTP-PLC again due to the fast NX at PLC (GEF activity). The activated PLC pool is thus not static but in a steady state of continual GTP-GDP NX and hydrolysis. In the model for native cells with agonist, $\sim 75\%$ of $G\alpha$ -PLC bears GDP, and just 25% bears GTP. The benefit of such dynamic GTP consumption is that it allows PLC to be turned off quickly when agonist is removed.

As a consequence of the rapid recycling of G proteins and the relatively low number of receptors in native cells, only a small fraction of their G proteins become dissociated into $G\alpha$ and $G\beta\gamma$ subunits (Fig. 6 C), consistent with the observation that receptors generally do

not have to compete for G proteins (e.g., Neubig, 1994). The situation changes markedly with overexpression of the $G\alpha$ and $G\beta$ FRET probes. The amount of G proteins then greatly surpasses endogenous PLC, and G protein deactivation must rely on the slow endogenous GTPase activity of $G\alpha$ until most of the G proteins are deactivated. Thus, deactivation is slow (time constant of 30 s; Figs. 5 B and 6 C), and a larger proportion (93%) of G proteins accumulates in the active dissociated form waiting to deactivate. Under such conditions, we would expect competition for G proteins to generate crosstalk between GPCRs.

To measure $G\alpha$ /PLC FRET, we coexpressed PLC and G proteins. Under such conditions, the model predicts that deactivation of G proteins is sped up and that the deactivation of PLC occurs at almost the same time (blue and green in Fig. 5 D). This is consistent with the faster recovery of $G\alpha$ /PLC FRET (time constant of 3.6 s) as compared with $G\alpha$ / $G\beta$ FRET. The GAP effect of PLC on termination of signaling was thus evident in our FRET experiments (Jensen et al., 2009).

In our ligand-gated GPCR system, the binding of G protein to activated receptor takes several hundred milliseconds when both are overexpressed, and longer with native levels of G proteins. This contrasts with photodetection by rhodopsin and the G protein transducin. The vertebrate photoreceptor is optimized for rapid response and is the best kinetically described of all GPCR signaling systems. In amphibian rod disk membranes, the endogenous densities of rhodopsin (Rh) and transducin (G_T) molecules are 25,000 and 2,500 μm^{-2} , respectively (Pugh and Lamb, 1993). The activated form of rhodopsin Rh* is formed within a couple of milliseconds of light absorption, and, at room temperature, one Rh* initiates activation and subunit dissociation of transducins at 4,000 s^{-1} , $\sim 60\%$ of the estimated collision rate in the disk membrane. Thus, in addition to having higher endogenous concentrations of the signaling proteins in the rod, the intrinsic rate constant for receptor activation of G proteins is ~ 500 times larger than for the system we studied (Hamer et al., 2005). Even if there might be preformed Rh- G_T complexes in the dark, they must break and reform several times per millisecond to allow the amplification steps of photoactivation. As with the PLC system, the effector (phosphodiesterase) is the GAP for transducin GTP, shortening its lifetime to 2 s. Finally, in the rod, the deactivation of Rh* requires actions of GPCR kinase and arrestin rather than dissociation of the ligand. Nevertheless, these deactivation steps take <1 s in the rod.

Conclusion

We have constructed a quantitative description of M_1R signaling that is based on FRET time courses and protein amounts. We find that the transient transfection of our probes generates a very broad range of protein

expression in different cells, and that choosing readily visible fluorescent cells selects cells with enormous overexpression of the proteins. Nevertheless, it is possible to work with such data to make a model that describes transfected and untransfected cells well, both in the time domain and in their steady-state concentration dependence on agonist. Indeed, the enormous overexpression allows approximating the density of signaling molecules as the number of fluorescent molecules. The model helps to interpret our FRET measurements further, showing clearly, for example, in living cells the GAP and GEF action of PLC on G_q . Several components of the model are not specific for M_1R and should be applicable to thinking about other signaling cascades.

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Note added in proof. After this article was in review, a relevant modeling paper appeared (Callender et al. 2009. *J. Theor. Biol.* doi:10.1016/j.jtbi.2009.10.030). If we assume that RAW 264.7 cells have the same 1,500- μm^2 membrane area, the surface densities per square micrometer deduced in that paper for untransfected cells are equivalent to PIP₂ 6,000 G_q G proteins 20 and P2Y₆ purinergic receptors 1.3.

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