Randolph et al. Vol. 196, No. 4, August 19, 2002. Pages 517–527.

Due to a typographical error, a $CD16^-$ and $CD16^+$ were switched in the middle of the third paragraph of page 521. The corrected paragraph appears below.

Differentiation and Migration Analysis after Depletion of $CD16^+$ Monocytes. In some depletion experiments, only the M-$DC8^+$ subpopulation of $CD14^+CD16^+$ monocytes was removed. Removal of M-$DC8^+$ cells only had no marked effect on the yield or phenotype of reverse-transmigrated cells (unpublished data), but when the depletion scheme eliminated all $CD16^+$ monocytes (CD16 depleted), 47 ± 22% ($P < 0.05$; four experiments) to 66 ± 13% ($P < 0.005$; four experiments) fewer reverse transmigrated cells were recovered from unstimulated and zymosan-stimulated cultures, respectively (Fig. 6 A). These results are in agreement with the CFSE experiments in Fig. 3. Even after thorough depletion of $CD16^+$ monocytes, many reverse-transmigrated cells recovered from cultures receiving only the $CD16^-$ monocytes expressed CD16 upon reverse transmigration, in contrast to the subendothelial monocyte-derived cells from the same cultures (Fig. 6 B). Thus, these cells appear to upregulate CD16 expression during reverse transmigration. When flow cytometry was conducted to analyze the maturation status of cells in the reverse-transmigrated fraction, the number of HLA-DR^+CD86^+ cells was 63 ± 14% (average of three experiments; $P < 0.005$) decreased per unit area of endothelial cell surface when blood $CD16^+$ monocytes were depleted from the starting population (Fig. 6 C; cells shown in each group were recovered from an equivalent area of endothelial surface). Moreover, the residual DCs recovered after depletion of $CD16^+$ monocytes expressed an order of magnitude less CD86 on the cell surface, indicating that these reverse-transmigrated, $CD16^-$ monocyte-derived cells were less mature than the reverse-transmigrated cells that develop in cultures that contained $CD16^+$ blood monocytes.