

IN VITRO MACROPHAGE MANIFESTATION OF  
CORTISONE-INDUCED DECREASE IN  
RESISTANCE TO MOUSE HEPATITIS VIRUS\*

BY C. E. TAYLOR, W. Y. WEISER,‡ AND F. B. BANG

*From the Department of Pathobiology, The Johns Hopkins University, School of Hygiene and Public Health, Baltimore, Maryland 21205*

It has been known since cortisone was first used that acute infections are often more severe when this drug is administered and that a number of immune responses are suppressed by this drug (1). We show here that peritoneal exudate cells (PEC) taken from hydrocortisone (HC)-treated, normally resistant C<sub>3</sub>H mice are 1,000 times more susceptible to challenge with mouse hepatitis virus (MHV) in vitro than are PEC from untreated mice. Furthermore, resistant macrophages from normal C<sub>3</sub>H mice succumb to viral destruction when cultured with spleen cells from cortisone-treated mice of the same strain but not with spleen cells of normal mice of the same strain.

**Materials and Methods**

*Animals.* 4-6-wk-old inbred C<sub>3</sub>H and C<sub>3</sub>H<sub>88</sub> mice were used. The C<sub>3</sub>H mice are resistant to mouse hepatitis virus (MHV-2) and have been maintained by continuous inbreeding in this laboratory since 1955. The C<sub>3</sub>H<sub>88</sub> strain of mice, into which the gene for susceptibility was introduced, was developed by crossing susceptible PRI mice with C<sub>3</sub>H mice and back-crossing the hybrids with the C<sub>3</sub>H. The offspring were selected by testing the susceptibility of macrophages from individual mice and then breeding from those that had susceptible cells. The congenic strain derived from this is just as susceptible as the PRI mouse (2).

*Virus.* The MHV-2 strain of mouse hepatitis virus originally obtained from Dr. J. Nelson, Rockefeller Institute, Princeton, N. J. in 1955, has been maintained in this laboratory by passage through 1-mo-old PRI mice.

*Culture of PEC.* Methods of harvesting and culturing peritoneal macrophages have been previously described in detail (3). PEC containing >90% macrophages were resuspended in Chang's medium and then dispensed into 12- × 100-mm Wassermann tubes. All tubes were incubated at 37°C in a roller drum.

*HC Treatment.* Mice were injected i.p. with 3 ml of thioglycollate medium on day 0, followed 12 h later by 2.5 mg HC (Solu-Cortef; Upjohn Co., Kalamazoo, Mich.). Two more injections of 2.5 mg HC were given 12 and 36 h after the first dose. PEC were harvested on day 3 (i.e., 24 h after the last dose). Control mice received phosphate-buffered saline instead of HC.

*Spleen Cells from Mice with Cortisone Treatment.* Mice were given 2.5 mg of cortisone on day 0; on day 3, they were killed and spleen cells were released by gentle teasing.  $0.5 \times 10^8$  spleen cells were cultured together with normal C<sub>3</sub>H macrophages for 24 h. For control, equal numbers of syngeneic spleen cells from untreated mice were used.

*Preparation of Supernatant Fluid from concanavalin A (Con A)-treated Spleen Cells.* Details of the preparation of supernatant fluid from Con A-treated spleen cells are provided elsewhere (4). Briefly, spleen cells from Con A-treated mice were cultured for 3 d in the presence of 2 µg/ml of Con A. Supernatant fluid was obtained by centrifuging the cell suspension at 1,000 rpm for 30 min.

\* Supported by grant 1 RO1 AI-16036 from the National Institutes of Health.

‡ Present address: Department of Medicine, Harvard Medical School, Boston, Mass. 02115.

## Results

*Effect of Adding Spleen Cells from Cortisone-treated Mice.* Previous work from our laboratory has suggested that susceptibility of macrophages may be regulated by various lymphokines. Therefore, cells from cortisone-treated mice were combined in vitro with normally resistant C<sub>3</sub>H macrophages and the susceptibility of these was tested. C<sub>3</sub>H mice were given 2.5 mg cortisone on day 0, and the spleens were removed 3 d later.  $0.5 \times 10^6$  spleen cells were then added to each culture of normal C<sub>3</sub>H macrophages. Control macrophage cultures received equal numbers of untreated spleen cells from syngeneic mice. Various dilutions of virus [MHV(PRI)] were inoculated into both groups of cultures. All cultures were kept for 6 d at 37°C. The results are summarized in Table I. Genetically resistant macrophages were destroyed by day 3 at 100-fold greater dilutions when they were cocultivated with spleen cells from cortisone-treated spleen cells but not the normal spleen cells. Spleen cells that had been killed by heating to 56°C for 30 min or by freezing and thawing did not alter the resistance of C<sub>3</sub>H macrophages.

*Direct Addition of HC to Peritoneal Adherent Cell Populations.* 10, 50, 100, 150, or 350 µg of HC was added to normal C<sub>3</sub>H macrophages in culture and incubated for a period of 24 h before viral infection. The mean lethal dose (LD<sub>50</sub>) of the virus in these macrophages was found to be  $10^{2.5}$  for control macrophages (i.e., no HC),  $10^{3.0}$ ,  $10^{3.0}$ ,  $10^{3.5}$ , and  $10^{2.8}$  when 10, 50, 100, and 150 µg of HC was given, respectively. None of the cultures inoculated with these amounts of HC alone showed destruction, but 350 µg of HC was toxic to the macrophages.

*In Vivo Induction of In Vitro Susceptibility of C<sub>3</sub>H Macrophages to MHV(PRI) Virus.* Earlier work had failed to induce large changes in susceptibility of macrophage cultures even though it was always clear that the mouse was made highly susceptible with cortisone. Table II, however, shows that macrophages from HC-treated resistant mice (which had been treated three times) were 400–1,000 times more susceptible to the virus than those from untreated mice. When the mice were given only one injection of HC, the macrophages were not susceptible. Viral growth was demonstrated primarily by cell destruction, which occurred as early as 2 d, but giant-cell formation also occurred which eventually ended in loss of contact with the culture tube. Virus yield was determined in critical cases. The 1,000-fold increase in susceptibility of the macrophages was routinely observed if three doses of HC ranging from 50 µg to 5 mg were given to the mice, but when 10 mg was used, the cells were only 10-fold more susceptible.

Shif and Bang (3) demonstrated that when high multiplicities of MHV(PRI) were

TABLE I  
*Effect of Adding Spleen Cells from Cortisone-treated Mice to Cultures of Syngeneic Macrophages*

Virus	Source of spleen cells*	
	Cortisone-treated mice	Untreated syngeneic mice
$10^{-1}$	6/6	6/6
$10^{-2}$	6/6	4/6
$10^{-3}$	6/6	0/6
$10^{-4}$	5/6	0/6
$10^{-6}$	0/6	0/6

\*  $0.5 \times 10^6$  spleen cells were added to each macrophage culture.

TABLE II  
Susceptibility of C<sub>3</sub>H Macrophages to MHV(PRI) After In Vivo Administration of HC

C <sub>3</sub> H* control	1 mg HC	2.5 mg HC	5 mg HC	10 mg HC	2.5 mg‡ HC	C <sub>3</sub> H <sub>SS</sub> * control
<i>log<sub>10</sub> LD<sub>50</sub>/0.1 ml</i>						
2.4	4.6	5.4	4.8	3.5	2.6	7.7

Macrophage cultures were infected with serial 10-fold dilutions of MHV(PRI). Readings were taken 5 d postinfection. Figures represent dilution of virus producing destruction.

\* Control mice received saline instead of HC.

‡ Single dose.

TABLE III  
Effect of Various Steroids on Resistance of Thioglycollate-elicited C<sub>3</sub>H Macrophages\*

C <sub>3</sub> H control	Dexamethasone	Prednisolone	Progesterone	Testosterone	HC
<i>log<sub>10</sub> LD<sub>50</sub>/0.1 ml</i>					
2.4	4.8	5.4	1.3	1.5	4.8

Each group of C<sub>3</sub>H mice (six per group) received three intraperitoneal doses (1 mg each) of the steroid after thioglycollate stimulation, and PEC were harvested 24 h after the last dose.

\* All macrophages in the experiment were seeded in 16-mm 24-well tissue culture cluster plates and incubated at 37°C with 5% CO<sub>2</sub> and humidified air.

given to C<sub>3</sub>H macrophages, a mutant virus, MHV(C<sub>3</sub>H), emerged that grew to high titer in both resistant and susceptible macrophages. Following the above findings, macrophage cultures from HC-treated mice receiving 10<sup>4</sup> LD<sub>50</sub> of MHV(PRI), and which were destroyed by the virus, were assayed for virus yield on C<sub>3</sub>H and C<sub>3</sub>H<sub>SS</sub> macrophages on days 5 and 7 postinfection. Virus harvested from macrophages of HC-treated mice on day 5 postinfection titered 10,000 times higher in C<sub>3</sub>H<sub>SS</sub> macrophages than in C<sub>3</sub>H macrophages. By day 7 postinfection, the difference had decreased to 2 logs, presumably because of late growth of the mutant. Because of this, most of the subsequent experiments were limited to observations within 5 d of inoculation.

The effect of cortisone is apparently glucocorticoid-specific. Prednisolone and dexamethasone were administered in the same way as HC: three successive injections, intraperitoneally, in vivo. Two steroids (progesterone and testosterone) were similarly administered. The results showed that the active glucocorticoids increased susceptibility of the C<sub>3</sub>H macrophages, whereas the other steroids did not (Table III).

*Growth Curve of MHV(PRI) in Macrophages from C<sub>3</sub>H<sub>SS</sub>, C<sub>3</sub>H, and HC-treated Mice.* Virus (1.5 × 10<sup>4</sup> tissue culture infective doses/2 × 10<sup>6</sup> cells) was adsorbed for 60 min at 37°C in a roller drum. The fluid was then replaced by culture medium. At various intervals, individual tubes were harvested, frozen, then titered for total cellular and supernatant virus.

As can be seen from Fig. 1, there was a sharp increase in virus titer in C<sub>3</sub>H<sub>SS</sub> macrophages during the first 10 h. There was no rapid initial rise in virus titer in either normal or HC-treated C<sub>3</sub>H macrophages. However, after about 11 h, the growth rate in the macrophages from HC-treated mice increased rapidly. At 21 h postinfection, virus replication in C<sub>3</sub>H resistant macrophages started to decelerate, whereas in macrophages from HC-treated C<sub>3</sub>H mice, it continued to increase. Although virus growth in the latter attained almost the same final yield as in C<sub>3</sub>H<sub>SS</sub>, it required ~20 h longer for this to occur.

In vivo, however, C<sub>3</sub>H mice challenged with virus 3 d after cortisone (2.5 mg s.c.) die in just as short, or shorter, time period than susceptible C<sub>3</sub>H<sub>SS</sub> mice (mean survival time: 1.9 d for C<sub>3</sub>H; 2.5 d for C<sub>3</sub>H<sub>SS</sub>). Earlier, Willenborg (7) had found that C<sub>3</sub>H

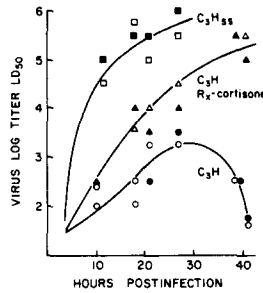


FIG. 1. Growth of MHV(PRI) in macrophages from C<sub>3</sub>H<sub>55</sub>, cortisone-treated (R<sub>c</sub>-cortisone) C<sub>3</sub>H mice and untreated C<sub>3</sub>H mice. Two independent experiments are shown for C<sub>3</sub>H<sub>55</sub> (■, □), cortisone-treated C<sub>3</sub>H (▲, △), and untreated C<sub>3</sub>H (●, ○). LD<sub>50</sub> were determined according to the method of Reed and Muench (6).

mice challenged with virus 1 h after cortisone administration take longer to die than genetically susceptible PRI mice. Evidently, the time of cortisone administration may be a factor in the induction of susceptibility to the virus.

*Reversal of HC-induced Modulation by Addition of Supernate from Con A-treated Spleen Cells.* It has been previously shown in our laboratory that genetically susceptible mice, as well as their macrophages, gain resistance when the mice are treated with Con A, or when the macrophages are treated with the supernate from Con A-treated spleen cells (4). It was therefore of interest to determine whether Con A would be antagonistic to cortisone.

Therefore, 0.1 ml of supernate fluid from Con A-treated spleen cells (4) and also from normal spleen cells was added to macrophages from HC-treated mice and incubated for 14 h before inoculation of virus. Table IV shows the complete reversal of the HC effect by supernatant fluid from Con A-treated spleen cells but only a partial reversal by supernate fluid from normal spleen cells.

The addition of normal spleen cells did not make the macrophages from cortisone-treated mice resistant again.  $0.5 \times 10^6$  spleen cells/ml, harvested from normal C<sub>3</sub>H mice, were added to macrophages from HC-treated mice and incubated for 24 h before viral infection. There was no significant difference between macrophages that received the spleen cells and those that did not (Table IV).

### Discussion

Although it has been known almost from the start of research on cortisone that this powerful anti-inflammatory drug depresses various immunologic mechanisms (1), the way in which it alters susceptibility to specific agents is not clear. The correspondence between in vivo and in vitro susceptibility of macrophages to mouse hepatitis made this model an attractive one to study. Cortisone-induced increase in susceptibility of mice to mouse hepatitis virus has also been observed by a number of other investigators as summarized by Vella and Starr (8). Administration of cortisone to the genetically resistant C<sub>3</sub>H mice (9), as well as to outbred mice, rendered them susceptible to viral [MHV(PRI)] infection. The LD<sub>50</sub> of MHV(PRI) virus was  $10^{8.5}$  in PRI mice,  $10^{6.5}$  in cortisone-treated C<sub>3</sub>H mice, and  $<10^{1.0}$  in untreated C<sub>3</sub>H mice (9). The slight or equivocal change in macrophage resistance, which had been demonstrated in vitro (7, 9), was in no way comparable to the marked effects on the intact animal. Gallily et al. (9), using a 1:100 dilution of stock virus ( $10^6$  LD<sub>50</sub>), had found that 85–100% of

TABLE IV  
*Effect of Adding Normal Spleen Cells or Supernatant Fluid from Con A-treated C<sub>3</sub>H Spleen Cells to Macrophages from HC-treated Mice (HC Macrophages)*

C <sub>3</sub> H control macrophages	HC macrophages	HC macrophages and untreated spleen cells	HC macrophages and Con A supernate	HC macrophages* and normal spleen cell supernate
2.3	5.7	5.5	1.5	3.5

$0.5 \times 10^6$  spleen cells or 0.1 ml of Con A-activated spleen supernate was added 24 and 14 h before viral infection, respectively.

\* Normal spleen cell supernate was obtained from a 96-h culture of normal C<sub>3</sub>H spleen cells.

cortisone-treated cultures were killed by virus as compared to ~20% of cultures given virus alone. Willenborg (7), using a single dose of 2.5 mg cortisone, had found that the LD<sub>50</sub> of MHV(PRI) virus in macrophage cultures from cortisone-treated C<sub>3</sub>H mice was 10<sup>3.4</sup>, and in cultures from untreated mice, it was 10<sup>3.2</sup>. In our studies, it was apparently crucial to administer three doses of HC to the mice to achieve sufficient suppression of viral resistance for in vitro expression of the effect. Presumably, a 1,000-fold increase of susceptibility of macrophages may explain most, if not all, of the 10<sup>5.5</sup> difference in titer in normal and cortisone-treated mice.

The mechanism whereby HC or cortisone changes the resistance of the all-important macrophage remains unclear. Addition of spleen cells from cortisone-treated mice enhanced the susceptibility of C<sub>3</sub>H macrophages 100-fold. Furthermore, cultures from HC-treated mice remained susceptible up to 72 h in culture, but fully regained resistance by 1 wk. The implication is that resistance of C<sub>3</sub>H macrophages is altered by some product of lymphoid cell activity induced by the steroid. That two other steroids, testosterone and progesterone, failed to cause the development of susceptible macrophages, whereas dexamethasone and prednisolone did, again suggests a correspondence of the in vitro results with the susceptibility of the whole animal.

The addition of whole spleen cells from normal resistant mice to macrophages from HC-treated mice for a period of 24 h before viral infection did not restore resistance. However, adding 0.1 ml of supernate fluid from Con A-stimulated spleen cells reversed the HC effect. This suggests that resistance is dependent, in part at least, upon a lymphokine released from T cells. Because Con A stimulates a variety of lymphokines, such as migration-inhibitory factor, interferon, soluble immune response suppressor, and T cell growth factor, it will be necessary to identify which specific lymphokine is responsible for the effect.

The growth-curve experiment reveals that during the first 12 h of observation, virus replication was equal in both sets of C<sub>3</sub>H macrophages, but that after this period, virus growth started to decelerate in the C<sub>3</sub>H control macrophages, although it continued to increase in macrophages from HC-treated mice. This suggests that resistance might be a result of the capacity of the C<sub>3</sub>H macrophage to produce an anti-viral substance during early growth which restricts viral growth.

The work of Gillis et al. (10) on how cortisone is able to suppress immune responses begins to explain the well known ability of cortisone to suppress antibody and T cell response. Suppression of the production of a T cell growth factor by this corticosteroid is then followed by a failure of the T cell compartment to expand after either specific antigenic stimulus or administration of Con A. That Con A and cortisone act

antagonistically in their studies (10) is very relevant to the similar blocking effect of one by the other in our system. However, because our studies seem to demonstrate that the cortisone-treated cells cause a change in the macrophage rather than prevent the release of a factor from lymphocytes, the analogy needs to be explored further.

### Summary

Genetically resistant C<sub>3</sub>H mice routinely yielded macrophages that were resistant when grown in 90% horse serum. These mice also routinely yielded macrophages that were susceptible to the same virus, MHV(PRI), in vitro after the mice had been treated with three intraperitoneal doses of hydrocortisone. Dexamethasone and prednisolone when similarly administered also increased the susceptibility of C<sub>3</sub>H macrophages taken from the treated animal, but progesterone and testosterone did not. In addition, spleen cells from mice treated with cortisone made the resistant C<sub>3</sub>H macrophages 100 times more susceptible in vitro.

Increased in vitro susceptibility induced in this way by hydrocortisone was reversed by exposure to supernatant fluid removed from cultures of concanavalin A-treated spleen cells.

*Received for publication 10 November 1980.*

### References

1. Parrillo, J. E., and A. S. Fauci. 1979. Mechanisms of glucocorticoid action on immune processes. *Annu. Rev. Pharmacol. Toxicol.* **19**:179.
2. Weiser, W., I. Vellisto, and F. B. Bang. 1976. Congenic strains of mice susceptible and resistant to mouse hepatitis virus. *Proc. Soc. Exp. Biol. Med.* **152**:499.
3. Shif, I., and F. B. Bang. 1970. In vitro interaction of mouse hepatitis virus and macrophages from genetically resistant mice. II. Biological characterization of a variant virus MHV(C<sub>3</sub>H) isolated from stocks of MHV(PRI). *J. Exp. Med.* **131**:851.
4. Weiser, W. Y., and F. B. Bang. 1977. Blocking of in vitro and in vivo susceptibility to mouse hepatitis virus. *J. Exp. Med.* **146**:1467.
5. Werb, Z. 1978. Biochemical actions of glucocorticoids on macrophages in culture. Specific inhibition of elastase, collagenase, and plasminogen activator secretion and effects on other metabolic functions. *J. Exp. Med.* **147**:1695.
6. Reed, L. D., and H. Muench. 1938. A simple method of estimating fifty percent end-points. *Am. J. Hyg.* **27**:493.
7. Willenborg, D. O. 1971. The effect of chemical and physical agents on the genetic resistance of mice to mouse hepatitis virus MHV(PRI). Doctor of Science Thesis. The Johns Hopkins University, School of Hygiene and Public Health, Baltimore, Md. 1.
8. Vella, P. P., and T. J. Starr. 1965. Effect of x-irradiation and cortisone on mouse hepatitis virus infection in germ-free mice. *J. Infect. Dis.* **115**:271.
9. Gallily, R., A. Warwick, and F. B. Bang. 1964. Effect of cortisone on genetic resistance to mouse hepatitis virus in vivo and in vitro. *Proc. Natl. Acad. Sci. U. S. A.* **51**:1158.
10. Gillis, S., G. R. Crabtree, and K. A. Smith. 1979. Glucocorticoid-induced inhibition of T cell growth factor production. I. The effect on mitogen-induced lymphocyte proliferation. *J. Immunol.* **123**:1624.