

# Increased intracellular pH is necessary for adult epithelial and embryonic stem cell differentiation

Bryne Ulmschneider,<sup>1,2</sup> Bree K. Grillo-Hill,<sup>3,4</sup> Marimar Benitez,<sup>1,2</sup> Dinara R. Azimova,<sup>1,2</sup> Diane L. Barber,<sup>3</sup> and Todd G. Nystul<sup>1,2</sup>

<sup>1</sup>Department of Anatomy, <sup>2</sup>Department of Obstetrics, Gynecology, and Reproductive Sciences, and <sup>3</sup>Department of Cell and Tissue Biology, University of California, San Francisco, San Francisco, CA 94143

<sup>4</sup>Department of Biological Sciences, San Jose State University, San Jose, CA 95192

Despite extensive knowledge about the transcriptional regulation of stem cell differentiation, less is known about the role of dynamic cytosolic cues. We report that an increase in intracellular pH (pHi) is necessary for the efficient differentiation of *Drosophila* adult follicle stem cells (FSCs) and mouse embryonic stem cells (mESCs). We show that pHi increases with differentiation from FSCs to prefollicle cells (pFCs) and follicle cells. Loss of the *Drosophila* Na<sup>+</sup>-H<sup>+</sup> exchanger *DNhe2* lowers pHi in differentiating cells, impairs pFC differentiation, disrupts germarium morphology, and decreases fecundity. In contrast, increasing pHi promotes excess pFC cell differentiation toward a polar/stalk cell fate through suppressing Hedgehog pathway activity. Increased pHi also occurs with mESC differentiation and, when prevented, attenuates spontaneous differentiation of naive cells, as determined by expression of microRNA clusters and stage-specific markers. Our findings reveal a previously unrecognized role of pHi dynamics for the differentiation of two distinct types of stem cell lineages, which opens new directions for understanding conserved regulatory mechanisms.

## Introduction

Cellular differentiation is a central feature of metazoan biology, driving tissue development, homeostasis, and repair. This process is often studied in the context of adult and embryonic stem cell (ESC) biology, where individual steps in the transition from a multipotent progenitor to a differentiated cell type can be carefully monitored. In both cell types, multiple regulatory mechanisms operate in concert to ensure that each step of differentiation occurs in a robust and precise manner. The role of developmental cues, transcription factors, and chromatin state in cellular differentiation has been the focus of intense investigation, but we know much less about the contributions of dynamic cytosolic signals.

In this study, we investigated how changes in intracellular pH (pHi) promote differentiation in the follicle stem cells (FSCs) of the *Drosophila melanogaster* ovary and mouse ESCs (mESCs). pHi dynamics are known to act as a cytosolic signal that contributes to the regulation of multiple cell processes, including cell cycle progression (Putney and Barber, 2003; Schreiber, 2005), membrane trafficking (Mukherjee et al., 2006; Brown et al., 2009; Kojima et al., 2012), and cell-substrate adhesion (Srivastava et al., 2008; Choi et al., 2013), and is dysregulated in some diseases, such as cancer (Webb et al., 2011; Parks et al., 2013) and neurodegenerative disorders (Harguindeguy et al., 2007; Wolfe et al., 2013). However, a role for

pHi dynamics in metazoan development remains understudied. Here, we show that pHi increases during the differentiation of FSCs and mESCs and is necessary for the efficient initial differentiation of both cell types. In addition, our data suggest a specific role for pHi dynamics in the regulation of Hedgehog (Hh) signaling in the FSC lineage.

## Results and discussion

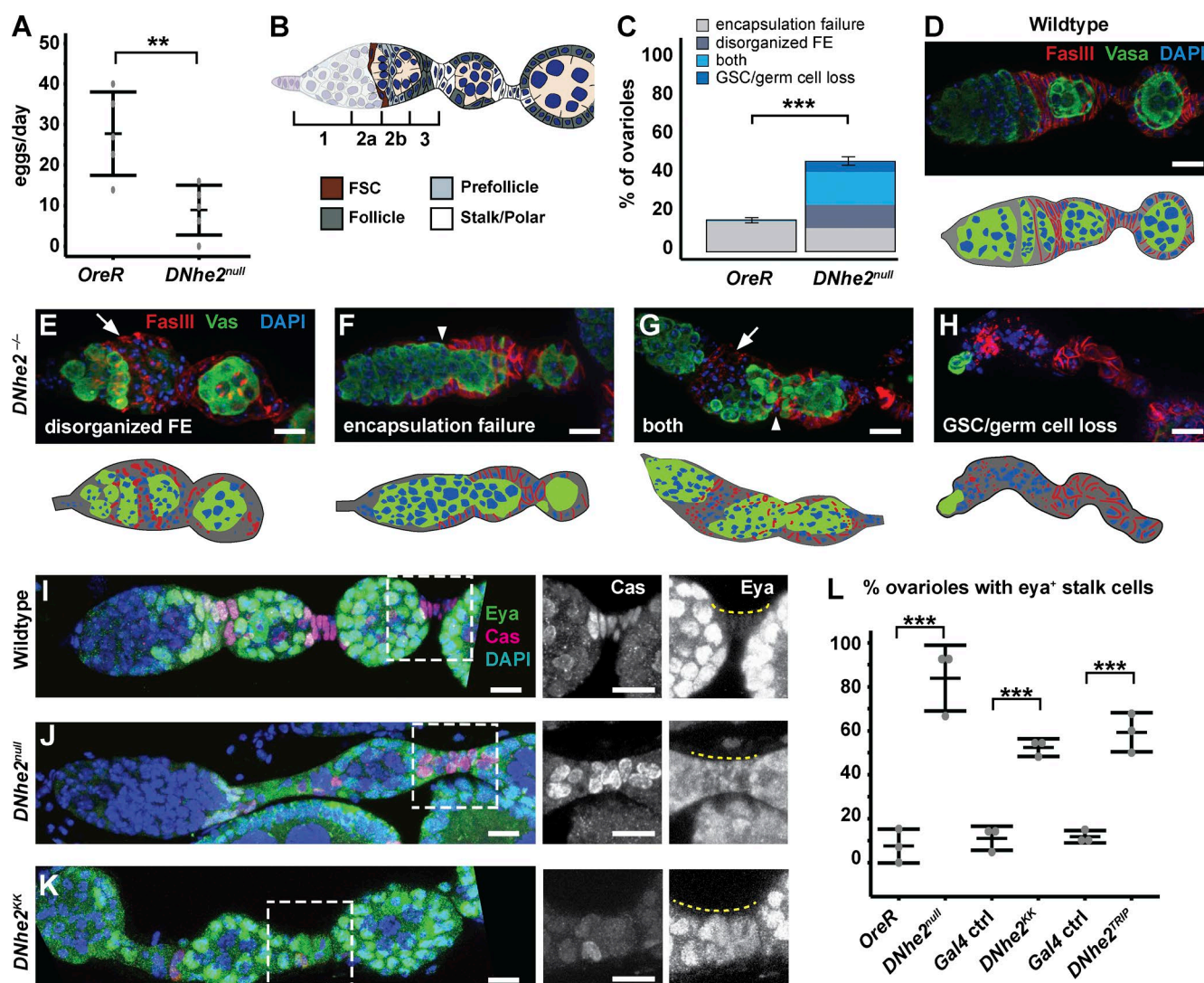
We previously reported that a null allele of *DNhe2*, an orthologue of the mammalian plasma membrane Na<sup>+</sup>-H<sup>+</sup> exchanger that catalyzes an influx of extracellular Na<sup>+</sup> and an efflux of intracellular H<sup>+</sup>, suppresses dysplasia with expression of activated oncogenes in *Drosophila* imaginal disks (Grillo-Hill et al., 2015). Through these studies, we noticed that flies homozygous for *DNhe2<sup>null</sup>* have reduced fertility. Thus, we performed an egg-laying assay and found that *DNhe2<sup>null</sup>* flies laid significantly fewer eggs per day compared with wild-type flies (Fig. 1 A). To investigate further, we searched for defects in oogenesis. The formation of new follicles during early oogenesis requires proper differentiation in the FSC lineage. This begins in the germarium (Fig. 1 B) with a pair of FSCs at the region 2a/2b border (Margolis and Spradling, 1995; Nystul and Spradling,

Correspondence to Todd G. Nystul: todd.nystul@ucsf.edu

Abbreviations used: Az, acetazolamide; EB, embryoid body; ESC, embryonic stem cell; FC, follicle cell; FSC, follicle stem cell; Hh, Hedgehog; LIF, leukemia inhibitory factor; mESC, mouse ESC; NRE, notch response element; pFC, prefollicle cell; pHi, intracellular pH.

© 2016 Ulmschneider et al. This article is distributed under the terms of an Attribution-Noncommercial-Share Alike-No Mirror Sites license for the first six months after the publication date (see <http://www.rupress.org/terms>). After six months it is available under a Creative Commons License (Attribution-Noncommercial-Share Alike 3.0 Unported license, as described at <http://creativecommons.org/licenses/by-nc-sa/3.0/>).



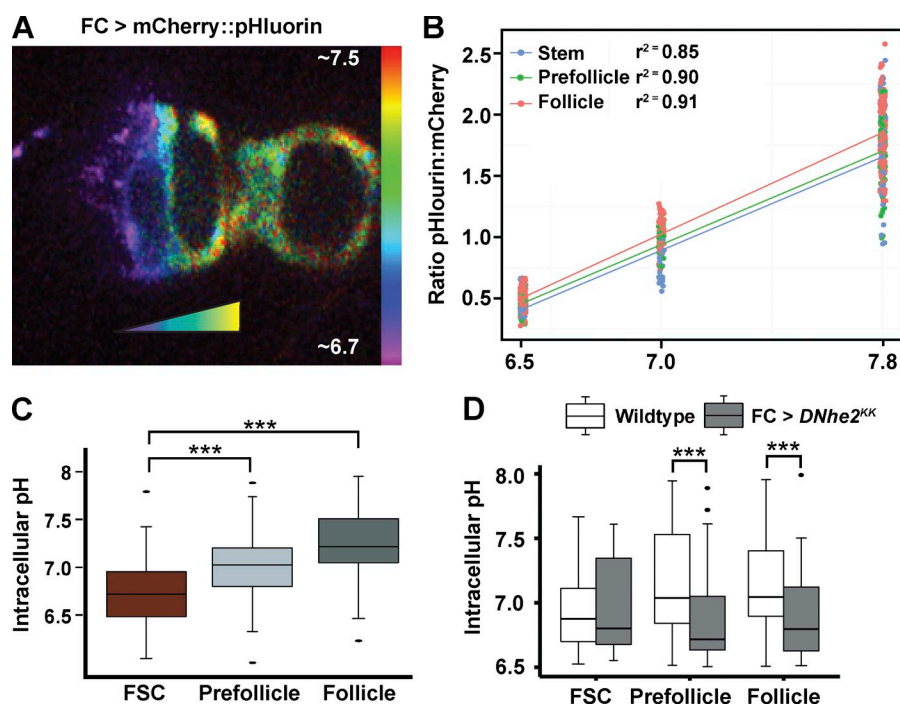


**Figure 1. *DNhe2* is necessary for differentiation in the FSC lineage.** (A) *DNhe2<sup>null</sup>* flies have significantly reduced egg laying. Graph depicts the mean number of eggs laid per female per day.  $n = 5$  independent replicates. \*\*,  $P < 0.01$ . (B) Diagram of the germarium showing the four regions, regions 1, 2a, 2b, and 3. Two FSCs (brown) are located in the middle of the germarium, at the region 2a/2b border. Cells that exit the FSC niche become pFCs (light gray) and then differentiate into main body FCs (dark gray), polar cells, or stalk cells (white). (C–H) *DNhe2<sup>null</sup>* ovarioles have morphological defects. (C) The frequency of each type of morphological defect.  $n = 3$  independent replicates;  $n > 50$  ovarioles. \*\*\*,  $P < 0.001$ . (D) Wild-type ovariole stained with FasIII to label FSCs, vasa to label germline, and DAPI. (E–H) Examples of ovarioles from *DNhe2<sup>null</sup>* flies. (E) A disorganized follicle epithelium (FE; arrow). (F) A failure of FCs to properly encapsulate the germline, resulting in fused cysts (arrowhead). (G) Both a disorganized follicle epithelium (arrow) and encapsulation defect (arrowhead). (H) A lack of germ cells and germline stem cells (GSCs). (I–L) Decreased *DNhe2* impairs stalk cell differentiation. Stalk cells (yellow dashed lines in I–K [right]) are Cas<sup>+</sup>, Eya<sup>-</sup> in wild-type ovarioles (I) but are Cas<sup>+</sup>, Eya<sup>+</sup> in *DNhe2<sup>null</sup>* (J) or *DNhe2<sup>KK</sup>* RNAi ovarioles (K). (L) Penetrance of Eya misexpression in stalks.  $n > 40$  ovarioles for all genotypes;  $n = 3$  independent replicates. \*\*\*,  $P < 0.001$ . Bars, 10  $\mu$ m. P-values were determined with a  $t$  test in B and a  $\chi^2$  test in C and L. Error bars represent SEM.

2007) that divide regularly to self-renew and produce prefollicle cell (pFC) daughters. Upon exiting the niche, a subset of pFCs begin to differentiate into polar and stalk cells (Larkin et al., 1996; Besse et al., 2002; Nystul and Spradling, 2010), which facilitate follicle budding, while the remaining pFCs differentiate into main body follicle cells (FCs) that surround the developing germline cyst. This well-defined lineage makes it possible to identify the stem cell and distinct stages of differentiation in vivo with single-cell resolution.

We observed clear morphological phenotypes in 45.2% of the ovarioles from *DNhe2<sup>null</sup>* flies (Fig. 1 C). In most cases, the pFCs either accumulated into disorganized clusters (Fig. 1 E) rather than forming into a single layered epithelium (Fig. 1 D); failed to completely encapsulate a germ cell cyst, resulting in

a fusion of two adjacent cysts (Fig. 1 F); or both (Fig. 1 G). In some ovarioles, there were few or no germ cell cysts, indicating a loss of germ cell production (Fig. 1 H). When stalks were present in *DNhe2<sup>null</sup>* ovarioles, stalk cells mostly remained rounded (Fig. 1 J) rather than acquiring their typical flattened shape (Fig. 1 I). In wild-type tissue, pFCs express both Castor (Cas) and Eyes absent (Eya), whereas stalk and polar cells are Cas<sup>+</sup>, Eya<sup>-</sup> (Fig. 1 I; Chang et al., 2013). However, in  $83 \pm 9\%$  ( $n = 42$ ) of *DNhe2<sup>null</sup>* ovarioles, cells in the first stalk downstream from the germarium remained Cas<sup>+</sup>, Eya<sup>+</sup> (Fig. 1 J) compared with  $9 \pm 4\%$  ( $n = 86$ ) of wild-type ovarioles (Figs. 1 L and S1, A and B). Knockdown of *DNhe2* in FSCs and early FCs using two separate RNAi lines driven by 109–30-Gal4 (FC-Gal4) produced phenotypes similar to *DNhe2<sup>null</sup>* flies, including the



**Figure 2. pH increases during pFC differentiation in a *DNhe2*-dependent manner.** (A) A germarium expressing *UAS-mCherry::pHluorin* in an FSC clone generated with the FLPout system. The pseudocoloring indicates the ratio of the fluorescence intensities of pHluorin to mCherry, which correlates with pH. Bar indicates pH estimates. (B) Three-point measurement of FSC clones expressing *mCherry::pHluorin* in freshly dissected ovarioles equilibrated to a pH of 6.5, 7.0, or 7.8, with nigericin buffers. The mean ratio of pHluorin to mCherry fluorescence increased linearly with increasing pH for each cell type ( $r^2 = 0.85$  for FSC,  $0.90$  for pFC, and  $0.91$  for FCs), confirming that the biosensor is responsive to pH changes as predicted when expressed in the follicle epithelium. (C) pH increases as cells transition from an FSC to an FC.  $n = 36$  germaria;  $n = 5$  independent replicates. (D) *Nhe2<sup>KK</sup>* RNAi driven in FCs with FC-Gal4 (gray boxes) causes a significant decrease in pH compared with control (white boxes) in pFC and FCs.  $n = 16$ – $17$  germaria;  $n = 3$  independent replicates. \*\*\*,  $P < 0.001$  ( $t$  test).

morphological phenotypes (Fig. S1, D–F) and *Cas<sup>+</sup>*, *Eya<sup>+</sup>* cells in the stalks (Fig. 1, K and L; and Fig. S1 C). However, the penetrance was lower, suggesting that there are some noncell autonomous effects on FC differentiation in *DNhe2<sup>null</sup>* flies or that knockdown with these RNAi lines is incomplete.

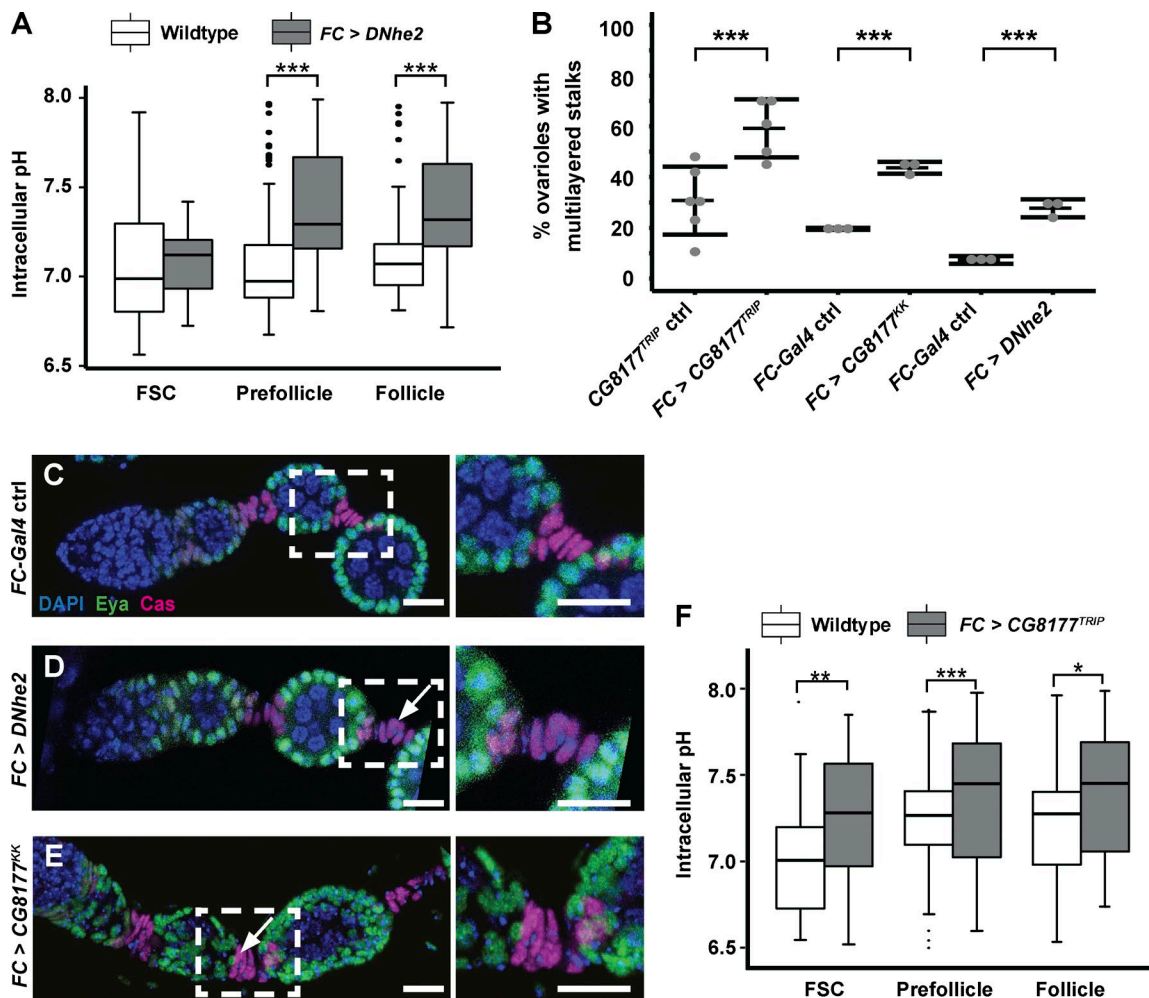
We previously reported that pH in eye epithelia is regulated by changes in *DNhe2* expression (Grillo-Hill et al., 2015) and hence investigated whether there are also pH changes in ovarioles, focusing on cells in the FSC lineage. We generated FSC clones expressing *mCherry::pHluorin* (Fig. 2 A), a genetically encoded pH biosensor (Koivusalo et al., 2010; Choi et al., 2013; Rossano et al., 2013; Grillo-Hill et al., 2015). The fluorescence intensity of pHluorin increases with increased pH within the physiological range (Miesenböck et al., 1998), whereas the fluorescence of mCherry is insensitive to physiological changes in pH and thus indicates biosensor abundance. Fluorescence ratios were converted to pH values by using the well-characterized nigericin calibration method (Fig. 2 B; Fig. S2, A–C; and Table S1; Meima et al., 2009; Choi et al., 2013; Grillo-Hill et al., 2014, 2015). We found significant differences in pH between each cell type in wild-type flies, with values of  $6.8 \pm 0.07$  for FSCs,  $7.0 \pm 0.06$  for pFCs, and  $7.3 \pm 0.05$  for FCs ( $P < 0.001$ ;  $n = 36$  germaria; Fig. 2 C and Table S2). Hence, pH is higher at progressively later stages of differentiation. RNAi knockdown of *DNhe2* significantly lowered the pH of pFCs and FCs by  $0.2$ – $0.25$  pH units (Fig. 2 D), indicating that the FC differentiation phenotypes we observed are associated with significant reductions in pH.

To determine how increased pH affects FC differentiation, we investigated the phenotypes caused by overexpression of *DNhe2*. Upon overexpression of *DNhe2*, pH in pFCs and FCs increased significantly ( $0.3$ – $0.35$  pH units,  $P < 0.001$ ; Fig. 3 A), and the stalks between budded follicles were multilayered in  $29 \pm 4\%$  ( $n = 113$ ; Fig. 3, B–D; and Fig. S2, D and E). To confirm that the stalk cell differentiation phenotype is caused by changes in pH, we performed an RNAi screen of 20 genes that are predicted to regulate pH homeostasis and identified

*CG8177*. *CG8177* is a putative  $\text{Cl}^-/\text{HCO}_3^-$  exchanger (an acid loader) with the highest similarity to SLC4A1 and SLC4A2. Knockdown of an acid loader is predicted to increase pH, and, indeed, we found that RNAi knockdown of *CG8177* using FC-Gal4 significantly increased pH by  $0.4$  pH units. In addition, *CG8177* knockdown significantly increased the frequency of multilayered stalks (Fig. 3, B, E, and F; and Fig. S2 F).

Polar and stalk cell differentiation are regulated by multiple signaling pathways, including Notch and Hh. Notch signaling is activated in a small subset of pFCs and is required specifically for polar cell differentiation, whereas Hh signaling is required in pFCs for differentiation toward the polar and stalk cell fates, but then must be down-regulated during the differentiation process for mature polar and stalk cells to form (Forbes et al., 1996a,b; Tworoger et al., 1999; Zhang and Kalderon, 2000; Besse et al., 2002). To test whether increased pH promotes polar and stalk cell differentiation by modulating the response to these differentiation cues, we examined the expression of Notch and Hh pathway activity reporters in pFCs overexpressing *DNhe2*. Overexpression of *DNhe2* did not affect the pattern or level of expression of the Notch pathway reporter, notch response element (NRE)–GFP (Fig. S2, G and H). However, we observed a significant decrease in the Hh pathway reporter, *Ptc-pelican-GFP*, in both early pFCs, just downstream from the FSC niche where Hh signaling is high, as well as in later pFCs, where Hh signaling is lower (Fig. 4, A–C). In addition, overexpression of *DNhe2* suppressed the stalk cell phenotype caused by overactivation of Hh signaling. Specifically, we found that knockdown of *ptc* by RNAi in FCs with FC-Gal4 caused the development of multilayered stalks in  $70 \pm 2\%$  ( $n = 467$ ) of ovarioles (Fig. 4, D and G), which is consistent with published results (Forbes et al., 1996a; Tworoger et al., 1999; Chang et al., 2013), whereas overexpression of *DNhe2* along with *ptc* RNAi reduced the frequency of this phenotype to  $30 \pm 5\%$  ( $n = 279$ ; Fig. 4, E and G). In contrast, overexpression of *DNhe2<sup>E358I</sup>*, an allele with a point mutation that prevents ion translocation (Denker et al., 2000; Grillo-Hill et al., 2015), did not increase





**Figure 3. Increasing pHi promotes excessive FC differentiation.** (A) Overexpression of *DNhe2* (gray boxes) increases pHi relative to controls (white boxes) in pFCs and FCs.  $n = 15\text{--}17$  germaria;  $n = 3$  independent replicates. (B) Penetrance of the stalk phenotypes with overexpression of *DNhe2* or RNAi knockdown of *CG8177*.  $n > 100$  for all genotypes;  $n > 3$  independent replicates. (C–E) Overexpression of *DNhe2* or knockdown of *CG8177* causes multilayered stalks. Stalk cells in a wild-type ovariole (C) form a single row, whereas overexpression of *DNhe2* (D) or knockdown of *CG8177* (E) causes the formation of multilayered stalks (white arrows). Insets are enlarged in C–E (right). (F) RNAi knockdown of *CG8177* (gray boxes) increases pHi relative to controls (white boxes).  $n = 17\text{--}19$  germaria;  $n = 3$  independent replicates. Bars, 10  $\mu\text{m}$ . \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ . P-values were determined with a  $t$  test in A and F and a  $\chi^2$  test in B.

pHi (Fig. S2 I) and did not suppress the *ptc* RNAi phenotype (Fig. 4 G). In addition, RNAi knockdown of *DNhe2* did not enhance the *ptc* RNAi phenotype (Fig. 4, F and G). These observations indicate that increased pHi attenuates wild-type and constitutively active Hh signaling.

To investigate how increased pHi attenuates Hh signaling, we examined the levels of a Smoothened::GFP (Smo::GFP) fusion protein expressed with FC-Gal4. We found that Smo::GFP levels were highest in region 2b, where pFC pHi is low (Fig. 2 A), and decreased progressively toward region 3, where pHi is higher (Fig. 4, H and K). In contrast, the level of an unrelated fusion protein, CD8::GFP, expressed with the same driver was uniform throughout the germarium (Fig. S2 J), indicating that the decrease in Smo::GFP is not caused by differences in FC-Gal4 expression levels or GFP stability in these regions. In addition, we found that overexpression of wild-type *DNhe2*, but not *DNhe2*<sup>E358I</sup>, significantly decreased Smo::GFP levels throughout the germarium (Fig. 4, I, J, and L). Collectively, our findings suggest a model in which an increase in pHi contributes to the patterning of pFC

differentiation, at least in part, by down-regulating Hh pathway activity (Fig. 4 M).

To determine the role of pHi dynamics in the differentiation of another stem cell type, we investigated mESCs (Murry and Keller, 2008; Young, 2011). Naive mESCs are maintained in a pluripotent state when cultured with leukemia inhibitory factor (LIF) and inhibitors of mitogen-activated protein kinase (MEK) and glycogen synthase kinase-3 $\beta$  (2i). Upon removal of LIF/2i, naive cells spontaneously differentiate to a primed state by 3–5 d (Ying et al., 2008). We measured pHi in cells loaded with the fluorescent H<sup>+</sup>-sensitive dye 2,7-bis(carboxyethyl)-5(6)-carboxyfluorescein (BCECF), calibrating ratios of pH-sensitive and pH-insensitive BCECF signals to pHi values with nigericin buffers as described previously (Meima et al., 2009; Grillo-Hill et al., 2015). Naive mESCs maintained a near-constant pHi between  $7.40 \pm 0.06$  and  $7.46 \pm 0.07$  over 6 d. However, in differentiating cells there was a transient increase in pHi to  $7.65 \pm 0.06$  at 48 h and  $7.57 \pm 0.05$  at 72 h ( $P < 0.001$ ; Fig. 5 A), with pHi returning to values similar to naive cells at 96 h and 6 d. Because a MEK inhibitor is included

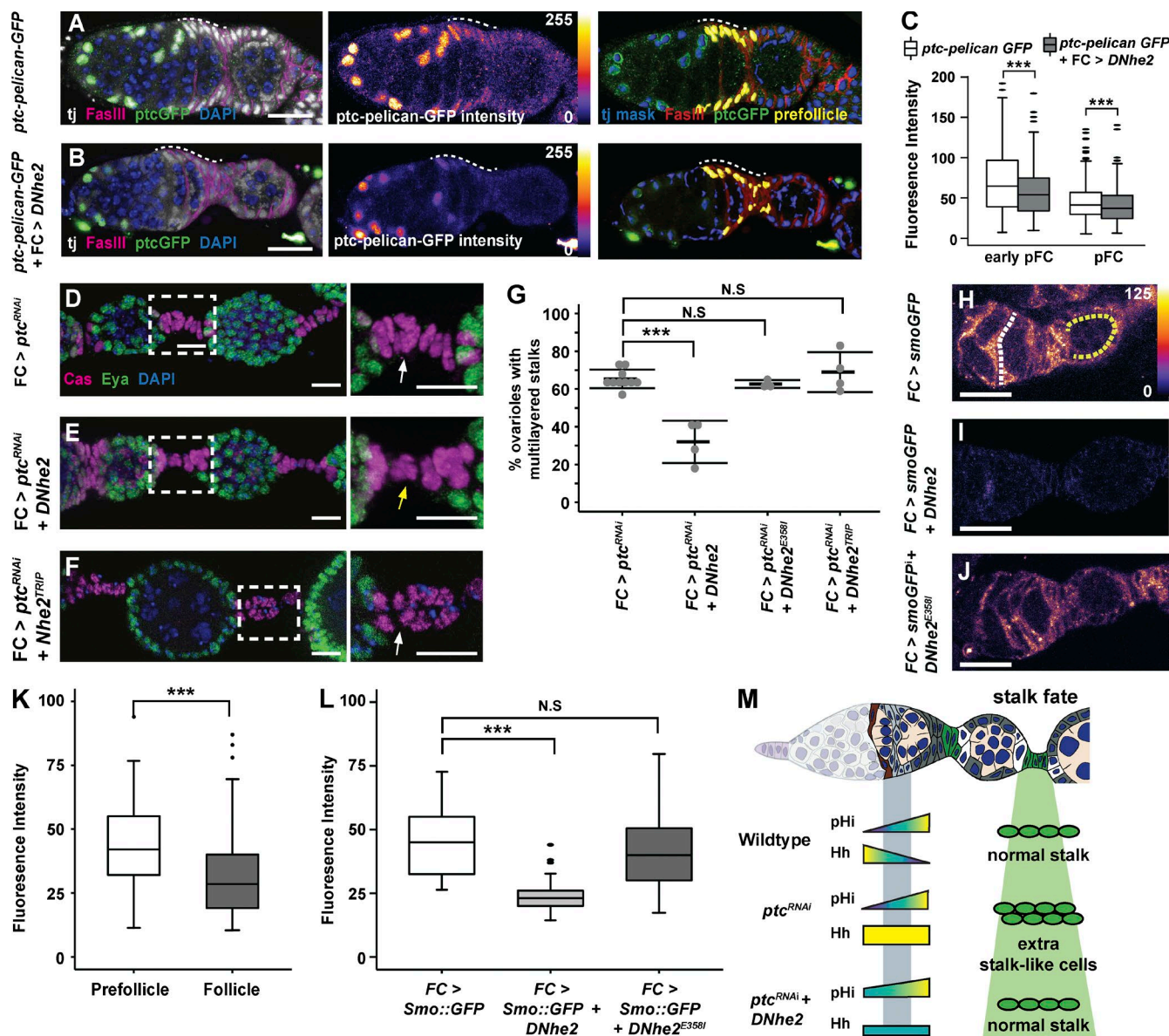
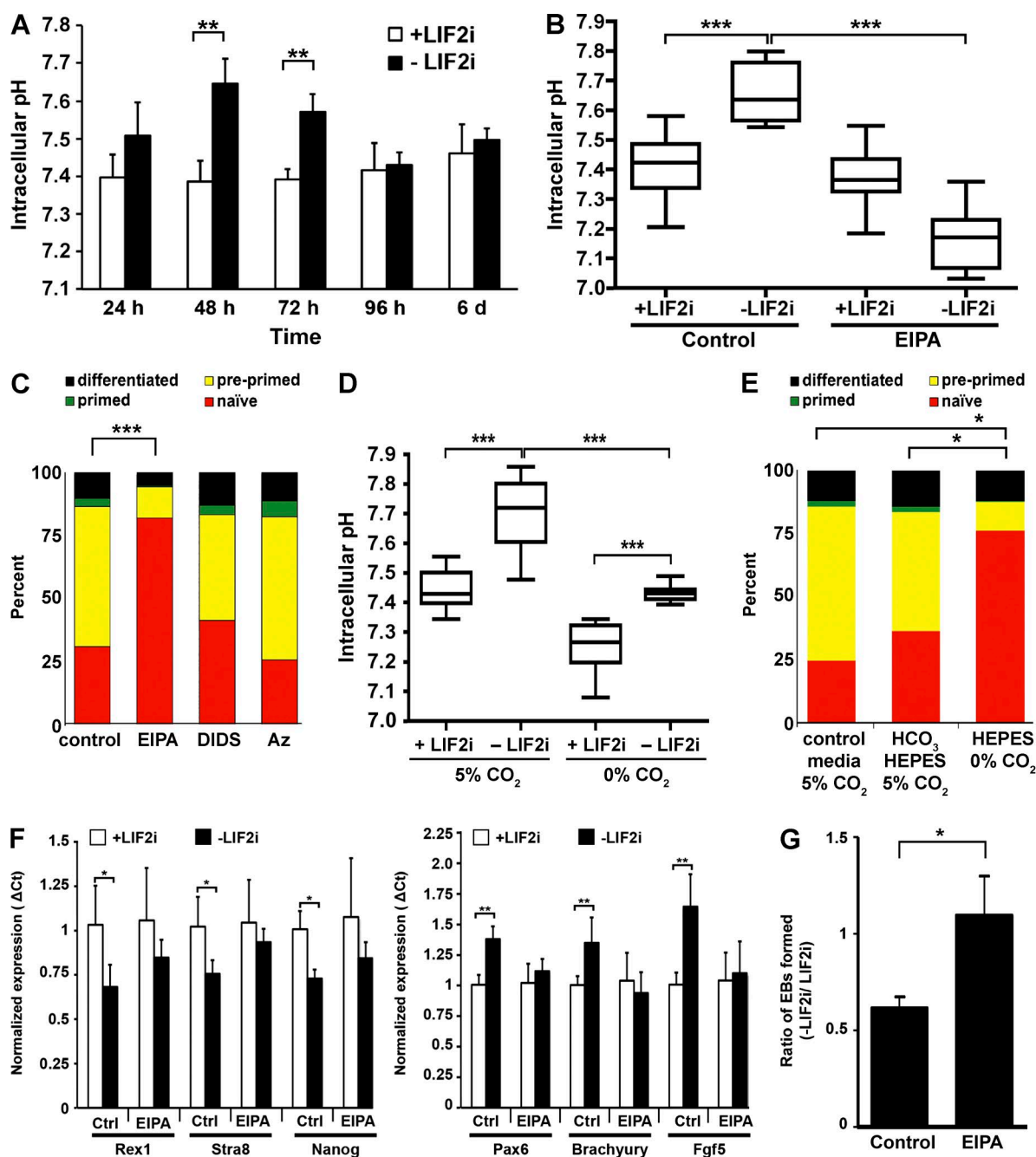


Figure 4. ***DNhe2* overexpression attenuates Hh signaling.** (A and B) Compared with control (A), overexpression of *DNhe2* decreases *Ptc-pelican-GFP* expression. Pseudocolor of GFP channel in A and B reflects fluorescence intensity. Nuclear volumes selected for intensity measurements (yellow) are shown (right). pFC regions are indicated with white dashed lines. (C) Quantification of *Ptc-pelican-GFP* fluorescence intensities in control germlaria (white boxes) with *DNhe2* overexpression (gray boxes) in early pFCs or pFCs.  $n > 450$  cells for all genotypes;  $n = 3$  independent replicates. (D–F) Multilayered stalk phenotype in FC > *ptc*<sup>RNAi</sup> (D) is less penetrant in FC > *ptc*<sup>RNAi</sup> + *DNhe2* (E), but not in FC > *ptc*<sup>RNAi</sup> + *DNhe2*<sup>TRIP</sup> RNAi (F). Insets enlarged in D–F (right). White and yellow arrows indicate stalks with double or single rows of cells, respectively. (G) The frequency of ovarioles with multilayered stalks in each genotype.  $n > 100$  ovarioles for all genotypes;  $n = 3$ –6 independent replicates. (H–J) In ovarioles expressing *Smo::GFP* only (H), the fluorescence intensity is higher in pFCs (yellow dotted line) than in FCs (white dotted line). Coexpression of wild-type *DNhe2* (I) but not *DNhe2*<sup>E358I</sup> (J) significantly reduces GFP fluorescence throughout the germlarium. (K) Quantifications of the fluorescence intensity in germlaria with FC-Gal4 driving *Smo::GFP* expression.  $n = 31$  germlaria. (L) Expression of *DNhe2* but not *DNhe2*<sup>E358I</sup> decreases the fluorescence intensity of *Smo::GFP*.  $n > 29$  germlaria for all genotypes. (M) Summary of the genetic interaction between RNAi knockdown of *ptc* and overexpression of *DNhe2*. In wild-type germlaria, pHi increases in pFCs as Hh pathway activity decreases, and a normal stalk forms. RNAi knockdown of *ptc* results in excessive Hh signaling and multilayered stalks. Overexpression of *DNhe2* increases pHi, attenuates Hh signaling, and suppresses the stalk cell phenotype caused by RNAi knockdown of *ptc*. Bars: (A, B, and H–J) 10  $\mu$ M; (D–F) 20  $\mu$ M. \*\*\*,  $P < 0.001$ . P-values were determined with a  $t$  test in C, K, and L and a  $\chi^2$  test in G.

in the LIF2i-containing medium for naive mESCs and MEK increases the activity of NHE1 and pHi (Malo et al., 2007), we adapted naive cells to LIF without 2i. In this condition, we observed a similar increase in pHi upon differentiation, indicating that the increased pHi does not merely reflect removal of MEK inhibition of NHE1 (Fig. S3 A).

To determine whether increased pHi is necessary for the spontaneous differentiation of naive mESCs, we first changed

pHi using the NHE1 selective pharmacological inhibitor 5-(*N*-ethyl-*N*-isopropyl) amiloride (EIPA; Putney et al., 2002). In naive cells maintained with LIF2i, pHi was not significantly different in the absence and presence of EIPA (Fig. 5 B). However, with EIPA, pHi did not increase with removal of LIF2i for 72 h and was significantly lower than in control naive cells, possibly reflecting loss of efflux of metabolic acids generated by increased glycolysis that occurs with differentiation (Ge et al., 2010). These



**Figure 5. Increased pHi during mESC differentiation is necessary for progression to primed fate.** (A) pHi measurements revealed no significant change over 6 d with LIF2i but significant increases at 48 and 72 h after removing LIF2i.  $n = 3$  cell preparations. (B) EIPA blocked the increase in pHi seen at 72 h after removal of LIF2i.  $n = 5$  cell preparations. (C) In the dual-reporter mESC line, naïve mESCs expressed *mir-290-mCherry*; preprimed cells expressed both *mir-290-mCherry* and *mir-302-eGFP*; primed cells expressed *mir-302-eGFP*; and further differentiated cells have silenced both reporters. After 72 h without LIF2i, 30.9% of control cells and 82.0% of EIPA-treated cells expressed *mir-290-mCherry* alone.  $n = 6$  cell preparations. (D) Cells maintained for 72 h in HEPES-buffered DMEM in the absence of CO<sub>2</sub> had lower pHi in the presence and absence of LIF2i compared with cells maintained at 5% CO<sub>2</sub>.  $n = 4$  cell preparations. (E) In the dual-reporter mESC line, more cells maintained in HEPES-buffered DMEM in the absence of CO<sub>2</sub> expressed in the naïve cell miRNA *mir-290-mCherry* (75.4%) compared with cells maintained at 5% CO<sub>2</sub> in either a predominantly HEPES-buffered DMEM (36.4%) or control DMEM (24.6%).  $n = 3$  cell preparations. (F) Naïve markers (Rex1, Stra8, and Nanog) were expressed at significantly lower levels, and differentiation markers (Pax6, T, and Fgf5) were expressed at significantly higher levels in control cells maintained without LIF2i for 72 h compared with naïve cells maintained with LIF2i. In contrast, in the presence of EIPA, naïve marker expression did not decrease, nor did differentiation marker expression increase upon removal of LIF2i. (G) Functional pluripotency, as measured by EB formation, is maintained in the presence of EIPA. The number of EBs formed after 72 h without LIF2i was normalized to the number of EBs formed in naïve mESCs for each experiment. Control mESCs form fewer EBs than naïve mESCs ( $61 \pm 5\%$ ). However, mESCs grown without LIF2i but with EIPA for 72 h form EBs as well as naïve mESCs ( $109 \pm 20\%$ ;  $P < 0.05$ ). Error bars are mean  $\pm$  standard deviation;  $n = 130$ –150 droplets;  $n = 3$  independent experiments. Box plots display mean  $\pm$  SEM. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$  ( $t$  test).



data suggest that NHE1 activity is necessary for the increased pHi in primed cells but not for steady-state pHi in naive mESCs.

We determined the differentiation state of mESCs in the absence and presence of EIPA using several approaches. We first used a dual-reporter mESC line expressing developmentally regulated miRNA clusters that are tagged with distinct fluorophores (Parchem et al., 2014). In this reporter line, naive mESCs express *mir-290-mCherry*, but not *mir-302-eGFP*; preprimed cells in intermediate stages express both reporters; primed cells express *mir-302-eGFP*, but not *mir-290-mCherry*; and both reporters are silenced in cells that have differentiated further. EIPA had no effect on the proportion of naive cells in self-renewing +LIF2i medium (Fig. S3 B). However, after 72 h without LIF2i, there was a significantly higher percentage (82.0%) of naive cells with EIPA compared with controls (30.9%;  $P < 0.001$ ; Fig. 5 C). After 5 d without LIF2i, there were also more *mir-290-mCherry*<sup>+</sup> cells in the presence of EIPA compared with controls (Fig. S3 C). We found no increase in cell death with EIPA treatment (Fig. S3 D), indicating that attenuated differentiation is not caused by selective elimination of preprimed and/or primed cells. In contrast to attenuated differentiation with EIPA, differentiation with the pharmacological inhibitors 4,4'-diisothiocyano-2,2'-stilbenedisulfonic acid (DIDS), which blocks activity of anion exchangers, or acetazolamide (Az), which blocks carbonic anhydrases, was not different than the control (Fig. 5 C). Moreover, despite the efficacy of both inhibitors in attenuating pHi recovery, when cells were rapidly switched from a HCO<sub>3</sub>-free Hepes buffer at 0% CO<sub>2</sub> to a Hepes-free buffer containing 25 mM NaHCO<sub>3</sub> superfused with 5% CO<sub>2</sub> (Fig. S3 E), neither inhibitor prevented the increase in pHi with differentiation (Fig. S3 F). Although NHE1 activity was previously shown to be necessary for differentiation of mESCs to cardiomyocytes (Li et al., 2009), to our knowledge, a role for increased pHi in the spontaneous differentiation of mESCs toward primed has not been reported.

To confirm that attenuated differentiation was caused by decreased pHi rather than by inhibiting a possible function of NHE1 independent of H<sup>+</sup> efflux, we used another approach to change pHi by maintaining cells in a predominantly Hepes-buffered medium at atmospheric 5% compared with 0% CO<sub>2</sub>. The pHi and differentiation of cells maintained at 5% CO<sub>2</sub> in DMEM containing 5 mM NaHCO<sub>3</sub> and 30 mM Hepes was not different than cells maintained at 5% CO<sub>2</sub> in control medium containing 44 mM NaHCO<sub>3</sub> and no Hepes (Fig. 5, D and E). In contrast, cells maintained at 0% CO<sub>2</sub> in a Hepes-buffered medium lacking NaHCO<sub>3</sub> had a lower pHi with LIF2i, and although pHi was increased after 72 h without LIF2i, it was significantly lower than controls without LIF2i and similar to controls with LIF2i (Fig. 5 D). Additionally, differentiation of dual-reporter cells maintained at 0% CO<sub>2</sub> was significantly attenuated, as indicated by the abundance of *mir-290-mCherry*<sup>+</sup> cells (Fig. 5 E). Collectively, these data indicate that an increase in pHi, likely to a pHi >7.5, is necessary for mESCs to differentiate toward the primed fate.

For a second method to determine the differentiation state of mESCs in the absence and presence of EIPA, we used quantitative RT-PCR to quantify expression of naive and primed markers. For control cells, we found that removal of LIF2i for 72 h decreased the expression of mESC markers *Rex1*, *Stra8*, and *Nanog* and increased the expression of primed cell markers *Fgf5*, *brachyury* (T), and *Pax6*, as expected (Parchem et al., 2014). However, with EIPA, there was no change in the

expression of these genes upon removal of LIF2i in cultures (Fig. 5 F), indicating that the increase in pHi is required for the expression of genes that drive differentiation. As a third metric for the differentiation state, we determined functional pluripotency in EIPA-treated cells cultured without LIF2i by testing for the formation of embryoid bodies (EBs) that form spontaneously when mESCs are grown in suspension (Kurosawa, 2007). We grew mESCs with or without LIF2i and EIPA for 72 h and then plated cells in suspension as described previously (Kurosawa, 2007). We found that control cells grown without LIF2i for 72 h formed EBs at a lower frequency than control cells grown with LIF2i ( $61 \pm 5\%$ ; Fig. 5 G). However, after 72 h without LIF2i, cells treated with EIPA formed EBs at a significantly higher frequency than control cells ( $109 \pm 20\%$ ;  $P < 0.05$ ), suggesting that mESCs treated with EIPA maintain functional pluripotency even in the absence of LIF2i.

Differentiation of adult and ESCs is regulated by distinct cues and is generally studied independently. Our study reveals that increased pHi contributes to the regulation of differentiation in both cell types. These findings open new directions to determine whether increased pHi is a conserved differentiation signal for other stem cell types and to resolve the molecular mechanisms mediating self-renewal and differentiation by pHi dynamics. As we described previously (Schönichen et al., 2013), regulation of cell processes by pHi dynamics is mediated by protonation of selective pH sensors, which acts as a posttranslational modification to change activity or ligand binding within the narrow physiological pH range. Our findings suggest that, in the FSC lineage, either Smo or a protein that regulates Smo stability may be a pH sensor. However, changes in pHi may also influence Hh signaling through effects on other cellular processes, such as vesicle trafficking, or by changing extracellular pH, neither of which can be assessed with our current tools. In addition, pHi dynamics could converge with Hh signaling and other cues (Simons et al., 2009) to regulate differentiation through a common effector such as the chromatin state, which is responsive to changes in pHi (McBrian et al., 2013). Although the pH sensors mediating mESC differentiation remain to be determined, several pH sensors we recently identified, including cofilin (Frantz et al., 2008), talin (Srivastava et al., 2008), and focal adhesion kinase (Choi et al., 2013), are required for mESC function or differentiation (Lee et al., 2011; Liu et al., 2011; Yun et al., 2012) and therefore may be components of the pH-responsive differentiation program. In summary, our findings show the critical importance of pHi dynamics in two distinct stem cell lineages, which advances our understanding of the cytosolic signals enabling stem cell differentiation.

## Materials and methods

### pHi measurements in *Drosophila*

FLPout clones were generated with a 15–30-min heat shock at 37°C and dissected 7–10 d after clone induction. In FSC clones, FSCs can be identified as the most anterior cell in the clone (Margolis and Spradling, 1995; Nystul and Spradling, 2007), pFCs are found just downstream from the FSCs, in region 2b, and more mature FCs are located further downstream, in region 3 of the germarium. For expressing UAS-mCherry::pHluorin with an FC-specific (FC) driver (109–30-Gal4), flies were dissected 7 d after eclosion. For pHi imaging, ovaries were dissected into bicarbonate buffer as described previously (Grillo-Hill et al., 2014, 2015) with 0.25 mg/ml concanavalin A conjugated to Alexa Fluor

647 and incubated for 15 min before imaging to visualize cell membranes (C21421; Thermo Fisher Scientific). Calibration curves were generated to calibrate mCherry::pHluorin ratios to pH values for each experiment and genotype. Although two-point calibration curves are commonly used, we tested whether the addition of a third point would improve pH estimates. However, the addition of a third point did not change the slope, Y intercept, or  $r^2$  value of the linear regression by >1% (Table S1). Thus, we used two-point calibration curves for all pH estimates. To generate the calibration curves, fresh ovaries were equilibrated into nigericin buffer, pH 6.5 or 7.5, for 15 min before imaging as described previously (Grillo-Hill et al., 2014, 2015) using 40  $\mu$ M nigericin (N1495; Invitrogen). 16-bit images in 1,024  $\times$  1,024 format were acquired with Leica Biosystems software on an SP5 or SP8 laser-scanning confocal microscope with a 40 $\times$  objective with a numerical aperture of 1.3 (Leica Biosystems), with gain, laser power, pinhole size, and other image acquisition settings consistent across matched control and experimental conditions. All imaging was done at room temperature. To obtain the fluorescence intensity measurement, we first subtracted background using a large region of interest in a black area of the image. Then, regions of interest were drawn around individual cells using the concanavalin A–Alexa Fluor 647 dye as a guide to outline individual cells. We measured one to two stem cells and three to five pFCs of bright and uniform intensity and selected the most highly expressed cells for measurement. Each region of interest was measured in one or two slices of 1  $\mu$ M. If two slices were used, fluorescence intensity was averaged over two slices. Calibration curves for each experiment, genotype, and cell type were generated, and linear regression analysis yielding  $r^2$  values of at least 0.70 were obtained and used to calculate pH values from ratios.

### **Drosophila tissue immunofluorescence staining**

Unless otherwise noted, ovaries were fixed and stained according to previously published immunostaining protocols (Castanieto et al., 2014). To reduce background in ovarioles stained for Cas and Eya, ovaries were blocked in 0.5% BSA (A7979; Sigma-Aldrich) in PBST (PBS + 0.2% Triton X-100) for at least 1 h before addition of both primary and secondary antibodies.

The following antibodies were used: from the Developmental Studies Hybridoma Bank, we purchased mouse anti-Eya (Eya-10H6, 1:50) and mouse anti-Fasciclin III (7G10, 1:100); guinea pig anti-traffic jam (1:1,000) was a gift from A. Spradling (Carnegie Institute of Washington, Baltimore, MD); from Thermo Fisher Scientific, we purchased rabbit anti-GFP (A11122, 1:1,000); from Santa Cruz Biotechnology, Inc., we purchased rabbit anti-vasa (1:1,000); and rabbit anti-castor (1:4,000 or 1:5,000) was a gift from W. Odenwald (National Institutes of Health, Bethesda, MD). The following secondary antibodies were used at 1:1,000: anti-mouse Alexa Fluor 488 and 555 (A11001 and A21422; Invitrogen), anti-rabbit Alexa Fluor 488 and 555 (A11008 and A21428; Invitrogen), and anti-guinea pig Alexa Fluor 633 (A21105; Invitrogen). Images were acquired on an SP5 or SP8 line-scanning confocal microscope for all experiments in which quantitative fluorescence comparisons were made and in experiments in which we scored for the presence of Eya. An M2 Axioimager microscope with an Apotome unit with a 63 $\times$  Apochromat objective with a numerical aperture of 1.4 (ZEISS) was used for images in which comparisons of morphological features were made. Images were cropped, rotated, contrast adjusted in Photoshop (Adobe), and saved as TIFF files.

### **Egg-laying assays**

Flies collected within 48 h were moved into egg-laying vessels. Egg-laying vessels consist of standard plastic polypropylene fly bottles turned

upside down onto standard fly food mixed with one to three drops of methylene blue dye with a small amount of wet yeast. One to three females per bottle were left overnight, up to 20 h, and then eggs were counted. The rate of eggs per fly per day was calculated as the hourly per fly egg-laying rate multiplied by 24 h.

### **Ptc-pelican-GFP quantitative analysis**

The middle section of a gerarium varying in thickness from 1 to 5  $\mu$ M stained with traffic jam, FasIII, GFP, and DAPI were imaged on an SP5 inverted confocal microscope. Imaris 3D analysis software (Bitplane) was used to render 3D surfaces over traffic jam expressing in all somatic cell nuclei. pFCs were then manually identified and selected based on the position and intensity of FasIII staining. Mean *Ptc-pelican-GFP* intensity per nuclear volume was measured with the Imaris software. Three independent experiments all had similar distributions, means, and standard deviations, so data were aggregated.

### **Smo::GFP quantitative analysis**

The middle section (one to five slices 1  $\mu$ M thick) of a gerarium stained with GFP and FasIII was imaged on an SP8 inverted confocal microscope. Two different types of quantitative analyses were performed. First, for control (109–30-gal4 > Smo::GFP), we quantified the background-subtracted fluorescence intensity for pFCs and FCs. For each of these regions, two or three lines were drawn through the brightest portion of the region, and the mean intensity of the line was measured. For the second quantification, we used the FasIII channel to mask the GFP channel and then took a measurement of the mean fluorescence intensity within each geraria to get a measurement of the total fluorescence intensity in every geraria.

### **Plotting and statistics for Drosophila data**

We used RStudio statistical analysis software to generate plots. For the boxplots displaying the pH data and the quantitative fluorescence measurements, the box represents the interquartile range, which is the middle 50% of the data. The black line is the median, and the whiskers show the range of the data, with outliers of 1.5 $\times$  the interquartile range plotted as dots. For the dot plots, the lines indicate the median and the standard deviation. We believe that displaying our data thus gives the reader the most accurate picture of the full variation within our data. Unless otherwise noted, we used RStudio to perform statistical comparisons and generate plots.

### **Stocks**

Stocks were generated with standard crossing schemes, maintained on standard molasses food, and cultured at 25°C unless otherwise noted, and all stocks were acquired from the Bloomington Drosophila Stock Center unless otherwise noted. For mCherry::pHluorin clones using the FLPout system,  $y[1] w[*]; P\{w[+mC]=GAL4-Act5C(FRT.CD2).P\}S$  (Bloomington ID 4780). For *ptcRNAi* experiments, *ptc<sup>RNAi</sup>* is a transgenic RNAi line from the Transgenic RNAi Project collection ( $y[1] v[1]; P\{y[+t7.7] v[+t1.8]=TRiP.JF03223\} attP2$ ; Bloomington ID 28795). All experiments using FC> refers to  $y^1 w^*$ ;  $P\{GawB\}109-30 /CyO$  (Bloomington ID 7023). For *DNhe2* knockdown via RNAi, *DNhe2<sup>KK102518</sup>* is a transgenic line containing  $P\{KK102518\}$  (Vienna Drosophila Resource Center, FlyBase ID FBti0117476). *DNhe2<sup>HMC03243</sup>* is a transgenic line from the Transgenic RNAi Project collection ( $y^1 v^1; P\{TRiP.HMC03243\} attP2$ ; Bloomington ID 51491). For *DNhe2* and UAS-mCherry::pHluorin, *UAS-DNhe2*, *UAS-DNhe2<sup>E3581</sup>*, *DNhe2<sup>null</sup>*, and *UAS-mCherry::pHluorin* were as described by Grillo-Hill et al. (2015). For CG8177 knockdown via RNAi, CG8177<sup>TRIP</sup> refers to  $y[1] v[1]; P\{y[+t7.7] v[+t1.8]=TRiP.HMC03399\} attP40$  (Bloomington ID 51827). CG8177<sup>KK</sup> refers to



P{KK100095}VIE-260B (Bloomington ID v109594, obtained from the Vienna Drosophila Resource Center). For Smo::GFP analysis, y[1] w[\*]; P{w[+mC]=UAS-smo.GFP}2 (Bloomington ID 44624). For *Ptc-pelican-GFP* analysis, yw\*; *Ptc-pelican-GFP* (a gift from T. Kornberg, University of California, San Francisco, San Francisco, CA). NRE-GFP refers to w[1118]; P{w[+m\*]=NRE-EGFP.S}5A (Bloomington ID 30727). CD8 GFP refers to yw; UAS-CD8::GFP/cyo (a gift from B. Ohlstein, Columbia University, New York, NY).

### mESC culture

Wild-type v6.5 and dual-reporter mESCs were a gift from R. Blelloch (University of California, San Francisco, San Francisco, CA). Cells were maintained as reported previously (Parchem et al., 2014). For differentiation assays, cells were plated in DMEM (BRL 0569; Gibco) containing LIF (ESGRO; EMD Millipore) and inhibitors for MEK (PD0325901, 04-0006-10; Stemgent) and glycogen synthase kinase-3 $\beta$  (CHIR99021, 04-0004-10; Stemgent), termed LIF2i. After plating for 24 h, spontaneous differentiation was initiated by washing cells three times in PBS and then maintaining cells for the indicated times in media without LIF2i. FACS analysis of dual-reporter mESCs was performed at the University of California, San Francisco, Flow Cytometry Core on an LSR II flow cytometer (BD), and analysis was performed using FACSDiva software (BD).

To make EBs, cells were plated and differentiated as described previously (Kurosawa, 2007) with or without EIPA for 72 h and then plated into hanging droplets ( $10^4$  cells per droplet) in media without LIF2i on the lids of 10-cm Petri dishes. Hanging droplets were cultured for 72 h. Droplets were washed in media without LIF2i to low-adhesion 6-well cell culture dishes, and the number of EBs was counted the next day. Data are reported as the percentage of EBs formed, calculated by dividing the number of EBs formed by cells cultured without LIF2i by the number of EBs formed in control cells maintained in media with LIF2i. EB data reported are from three independent cell preparations with the following numbers of droplets per condition: control +LIF2i (150), control -LIF2i (150), EIPA +LIF2i (132), and EIPA -LIF2i (141). Data were analyzed, and statistical analysis was performed in Excel (Microsoft) using a paired *t* test.

### pHi measurements of mESCs

pHi was measured as described previously (Meima et al., 2009; Grillo-Hill et al., 2015). In brief, cells maintained in 24-well plates were loaded with 1  $\mu$ M fluorescent H<sup>+</sup>-sensitive dye BCECF in an HCO<sub>3</sub><sup>-</sup>-containing buffer (110 mM NaCl, 5 mM KCl, 10 mM glucose, 25 mM NaHCO<sub>3</sub>, 1 mM MgSO<sub>4</sub>, 1 mM KPO<sub>4</sub>, and 2 mM CaCl<sub>2</sub>, pH 7.4) and washed, and ratios of BCECF fluorescence at Ex490/Em530 and Ex440/Em530 were acquired every 15 s for 5 min using a SpectraMax M5 plate reader (Molecular Dynamics). The fluorescence ratios were converted to pHi by calibrating each well with 10  $\mu$ M nigericin (Invitrogen) in 105 mM KCl. Pharmacological inhibitors were added 24 h after plating and included EIPA (ALX-550-266-M005; final concentration of 10  $\mu$ M; Enzo), DIDS (309795; final concentration of 1  $\mu$ M; EMD Millipore), and Az (A6011; final concentration of 200  $\mu$ M; Sigma-Aldrich). The efficacy of DIDS and Az was confirmed by their ability to inhibit pHi recovery after rapidly switching cells from a nominally HCO<sub>3</sub><sup>-</sup>-free Hepes buffer at 0% CO<sub>2</sub> to a Hepes-free buffer containing 25 mM NaHCO<sub>3</sub> superfused with 5% CO<sub>2</sub>, as previously described (Hulikova et al., 2014). For experiments with altered CO<sub>2</sub> and NaHCO<sub>3</sub> medium, cells were plated and maintained in control DMEM for 24 h, washed, and then maintained for 72 h in the absence or presence of LIF2i in NaHCO<sub>3</sub>-free DMEM (D5030; Sigma-Aldrich) supplemented with 4.5 g/liter glucose, 30 mM Hepes, pH 7.4,

0.11 mg/liter Na-pyruvate, 1 $\times$  Glutamax (35050-061; Gibco), 1 $\times$  nonessential amino acids, 2 mM glutamine, 100 mg/liter penicillin/streptomycin, and 15% fetal bovine serum. Medium for cells maintained at 5% CO<sub>2</sub> but not for cells maintained at 0% CO<sub>2</sub> was also supplemented with 5 mM NaHCO<sub>3</sub>.

### Quantitative RT-PCR experiments

RNA was extracted from mESCs maintained in the absence or presence of LIF2i for 72 h using TRIzol (Ambion) according to the manufacturer's protocol with the following modifications: 800  $\mu$ l TRIzol was added to cells, 2  $\mu$ l glycoblue (AM9515; Ambion) was added to visualize the RNA pellet, and the pellet was rinsed in 75% EtOH. RNA purity was assessed on a Nanodrop spectrometer. cDNA was synthesized using a Superscript III kit (18080093; Invitrogen) from 2 mg RNA and diluted 1:1 before being added to the PCR reaction. Primers were as described in Parchem et al. (2014). Quantitative PCR was performed with SYBR Greener (quantitative PCR SuperMix for ABI prism; 11760; Invitrogen) according to the manufacturer's protocol on a Real-Time PCR ViiA7 instrument (Applied Biosystems). Quantitative PCR analysis was done in the open source RStudio software using the ReadqPCR and NormqPCR (Perkins et al., 2012). Open source packages are available on the Bioconductor website. For each gene, three to five independent cell preparations were used.

### Online supplemental material

Fig. S1 shows additional images of the Eya channel for images stained with Cas and Eya. The images were background subtracted in ImageJ and then displayed as either pseudocolored images to reflect fluorescence intensity or as grayscale. Additional morphological features of *FC > DNhe2<sup>KK</sup>* stained with FasIII and Vasa are also shown. Fig. S2 shows example images of calibration curves, example calibrations for quantitative pH imaging, and images of the Notch reporter, NRE-GFP, as well as quantitative pH measurements for *FC > DNhe2<sup>E358I</sup>*. Fig. S3 shows additional aspects of the interaction between changes in pHi and mESC differentiation, including the lack of an effect of EIPA on cell survival, dual-reporter expression indicating EIPA maintains more cells in a naive state, and increased pHi in differentiating cells being independent of relieving MEK inhibition. Table S1 shows the differences in the slope and intercept of the linear regression model using two or three calibration points. Table S2 shows the summary of the pHi estimates for the wild-type controls in all experiments.

### Acknowledgments

We thank Sumitra Tatapudy for assistance with image acquisition of mCherry::pHluorin, Bradley Webb for assistance with quantitative pHi imaging analysis, and Torsten Wittmann and members of the Wittmann, Barber, and Nystul laboratories for helpful discussions. We also thank Robert Blelloch, Tom Kornberg, and Ophir Klein for suggestions on the manuscript.

This work was funded by the California Institute of Regenerative Medicine fellowship TG2-01153 to B. Ulmschneider and National Institutes of Health grants GM097158 to T.G. Nystul, GM47413 to D.L. Barber, and GM116384 to D.L. Barber and T.G. Nystul.

The authors declare no competing financial interests.

Submitted: 7 June 2016

Revised: 18 August 2016

Accepted: 5 October 2016

## References

- Besse, F., D. Busson, and A.-M. Pret. 2002. Fused-dependent Hedgehog signal transduction is required for somatic cell differentiation during *Drosophila* egg chamber formation. *Development*. 129:4111–4124.
- Brown, D., T.G. Paunescu, S. Breton, and V. Marshansky. 2009. Regulation of the V-ATPase in kidney epithelial cells: dual role in acid–base homeostasis and vesicle trafficking. *J. Exp. Biol.* 212:1762–1772. <http://dx.doi.org/10.1242/jeb.028803>
- Castanieto, A., M.J. Johnston, and T.G. Nystul. 2014. EGFR signaling promotes self-renewal through the establishment of cell polarity in *Drosophila* follicle stem cells. *eLife*. 3. <http://dx.doi.org/10.7554/eLife.04437>
- Chang, Y.-C., A.C.-C. Jang, C.-H. Lin, and D.J. Montell. 2013. Castor is required for Hedgehog-dependent cell-fate specification and follicle stem cell maintenance in *Drosophila* oogenesis. *Proc. Natl. Acad. Sci. USA*. 110:E1734–E1742. <http://dx.doi.org/10.1073/pnas.1300725110>
- Choi, C.-H., B.A. Webb, M.S. Chimenti, M.P. Jacobson, and D.L. Barber. 2013. pH sensing by FAK-His58 regulates focal adhesion remodeling. *J. Cell Biol.* 202:849–859. <http://dx.doi.org/10.1083/jcb.201302131>
- Denker, S.P., D.C. Huang, J. Orlowski, H. Furthmayr, and D.L. Barber. 2000. Direct binding of the Na–H exchanger NHE1 to ERM proteins regulates the cortical cytoskeleton and cell shape independently of H<sup>+</sup> translocation. *Mol. Cell*. 6:1425–1436. [http://dx.doi.org/10.1016/S1097-2765\(00\)00139-8](http://dx.doi.org/10.1016/S1097-2765(00)00139-8)
- Forbes, A.J., H. Lin, P.W. Ingham, and A.C. Spradling. 1996a. hedgehog is required for the proliferation and specification of ovarian somatic cells prior to egg chamber formation in *Drosophila*. *Development*. 122:1125–1135.
- Forbes, A.J., A.C. Spradling, P.W. Ingham, and H. Lin. 1996b. The role of segment polarity genes during early oogenesis in *Drosophila*. *Development*. 122:3283–3294.
- Frantz, C., G. Barreiro, L. Dominguez, X. Chen, R. Eddy, J. Condeelis, M.J.S. Kelly, M.P. Jacobson, and D.L. Barber. 2008. Cofilin is a pH sensor for actin free barbed end formation: role of phosphoinositide binding. *J. Cell Biol.* 183:865–879. <http://dx.doi.org/10.1083/jcb.200804161>
- Ge, L., W. Meng, H. Zhou, and N. Bhowmick. 2010. Could stroma contribute to field cancerization? *Med. Hypotheses*. 75:26–31. <http://dx.doi.org/10.1016/j.mehy.2010.01.019>
- Grillo-Hill, B.K., B.A. Webb, and D.L. Barber. 2014. Ratiometric imaging of pH probes. *Methods Cell Biol.* 123:429–448. <http://dx.doi.org/10.1016/B978-0-12-420138-5.00023-9>
- Grillo-Hill, B.K., C. Choi, M. Jimenez-Vidal, and D.L. Barber. 2015. Increased H<sup>+</sup> efflux is sufficient to induce dysplasia and necessary for viability with oncogene expression. *eLife*. 4. <http://dx.doi.org/10.7554/eLife.03270>
- Harguindey, S., S.J. Reshkin, G. Orive, J.L. Arranz, and E. Anitua. 2007. Growth and trophic factors, pH and the Na<sup>+</sup>/H<sup>+</sup> exchanger in Alzheimer's disease, other neurodegenerative diseases and cancer: new therapeutic possibilities and potential dangers. *Curr. Alzheimer Res.* 4:53–65. <http://dx.doi.org/10.2174/156720507779939841>
- Hulikova, A., N. Aveyard, A.L. Harris, R.D. Vaughan-Jones, and P. Swietach. 2014. Intracellular carbonic anhydrase activity sensitizes cancer cell pH signaling to dynamic changes in CO<sub>2</sub> partial pressure. *J. Biol. Chem.* 289:25418–25430. <http://dx.doi.org/10.1074/jbc.M114.547844>
- Koivusalo, M., C. Welch, H. Hayashi, C.C. Scott, M. Kim, T. Alexander, N. Touret, K.M. Hahn, and S. Grinstein. 2010. Amiloride inhibits macropinocytosis by lowering submembranous pH and preventing Rac1 and Cdc42 signaling. *J. Cell Biol.* 188:547–563. (published erratum appears in *J. Cell Biol.* 2010. 189:385) <http://dx.doi.org/10.1083/jcb.200908086>
- Kojima, A., J.Y. Toshima, C. Kanno, C. Kawata, and J. Toshima. 2012. Localization and functional requirement of yeast Na<sup>+</sup>/H<sup>+</sup> exchanger, Nhx1p, in the endocytic and protein recycling pathway. *Biochim. Biophys. Acta*. 1823:534–543. <http://dx.doi.org/10.1016/j.bbamer.2011.12.004>
- Kurosawa, H. 2007. Methods for inducing embryoid body formation: in vitro differentiation system of embryonic stem cells. *J. Biosci. Bioeng.* 103:389–398. <http://dx.doi.org/10.1263/jbb.103.389>
- Larkin, M.K., K. Holder, C. Yost, E. Giniger, and H. Ruohola-Baker. 1996. Expression of constitutively active Notch arrests follicle cells at a precursor stage during *Drosophila* oogenesis and disrupts the anterior–posterior axis of the oocyte. *Development*. 122:3639–3650.
- Lee, S.H., Y.J. Lee, and H.J. Han. 2011. Role of hypoxia-induced fibronectin–integrin  $\beta$ 1 expression in embryonic stem cell proliferation and migration: involvement of PI3K/Akt and FAK. *J. Cell. Physiol.* 226:484–493. <http://dx.doi.org/10.1002/jcp.22358>
- Li, X., P. Karki, L. Lei, H. Wang, and L. Fliegel. 2009. Na<sup>+</sup>/H<sup>+</sup> exchanger isoform 1 facilitates cardiomyocyte embryonic stem cell differentiation. *Am. J. Physiol. Heart Circ. Physiol.* 296:H159–H170. <http://dx.doi.org/10.1152/ajpheart.00375.2008>
- Liu, J., X. He, Y. Qi, X. Tian, S.J. Monkley, D.R. Critchley, S.A. Corbett, S.F. Lowry, A.M. Graham, and S. Li. 2011. Talin1 regulates integrin turnover to promote embryonic epithelial morphogenesis. *Mol. Cell. Biol.* 31:3366–3377. <http://dx.doi.org/10.1128/MCB.01403-10>
- Malo, M.E., L. Li, and L. Fliegel. 2007. Mitogen-activated protein kinase–dependent activation of the Na<sup>+</sup>/H<sup>+</sup> exchanger is mediated through phosphorylation of amino acids Ser770 and Ser771. *J. Biol. Chem.* 282:6292–6299. <http://dx.doi.org/10.1074/jbc.M611073200>
- Margolis, J., and A. Spradling. 1995. Identification and behavior of epithelial stem cells in the *Drosophila* ovary. *Development*. 121:3797–3807.
- McBrian, M.A., I.S. Behbahan, R. Ferrari, T. Su, T.-W. Huang, K. Li, C.S. Hong, H.R. Christofk, M. Vogelauer, D.B. Seligson, and S.K. Kurdastani. 2013. Histone acetylation regulates intracellular pH. *Mol. Cell*. 49:310–321. <http://dx.doi.org/10.1016/j.molcel.2012.10.025>
- Meima, M.E., B.A. Webb, H.E. Witkowska, and D.L. Barber. 2009. The sodium–hydrogen exchanger NHE1 is an Akt substrate necessary for actin filament reorganization by growth factors. *J. Biol. Chem.* 284:26666–26675. <http://dx.doi.org/10.1074/jbc.M109.019448>
- Miesenböck, G., D.A. De Angelis, and J.E. Rothman. 1998. Visualizing secretion and synaptic transmission with pH-sensitive green fluorescent proteins. *Nature*. 394:192–195. <http://dx.doi.org/10.1038/28190>
- Mukherjee, S., L. Kallay, C.L. Brett, and R. Rao. 2006. Mutational analysis of the intramembranous H10 loop of yeast Nhx1 reveals a critical role in ion homeostasis and vesicle trafficking. *Biochem. J.* 398:97–105. <http://dx.doi.org/10.1042/BJ20060388>
- Murry, C.E., and G. Keller. 2008. Differentiation of embryonic stem cells to clinically relevant populations: lessons from embryonic development. *Cell*. 132:661–680. <http://dx.doi.org/10.1016/j.cell.2008.02.008>
- Nystul, T., and A. Spradling. 2007. An epithelial niche in the *Drosophila* ovary undergoes long-range stem cell replacement. *Cell Stem Cell*. 1:277–285. <http://dx.doi.org/10.1016/j.stem.2007.07.009>
- Nystul, T., and A. Spradling. 2010. Regulation of epithelial stem cell replacement and follicle formation in the *Drosophila* ovary. *Genetics*. 184:503–515. <http://dx.doi.org/10.1534/genetics.109.109538>
- Parchem, R.J., J. Ye, R.L. Judson, M.F. LaRussa, R. Krishnakumar, A. Blelloch, M.C. Oldham, and R. Blelloch. 2014. Two miRNA clusters reveal alternative paths in late-stage reprogramming. *Cell Stem Cell*. 14:617–631. <http://dx.doi.org/10.1016/j.stem.2014.01.021>
- Parks, S.K., J. Chiche, and J. Pouyssegur. 2013. Disrupting proton dynamics and energy metabolism for cancer therapy. *Nat. Rev. Cancer*. 13:611–623. <http://dx.doi.org/10.1038/nrc3579>
- Perkins, J.R., J.M. Dawes, S.B. McMahon, D.L.H. Bennett, C. Orengo, and M. Kohl. 2012. ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*. 13. <http://dx.doi.org/10.1186/1471-2164-13-296>
- Putney, L.K., and D.L. Barber. 2003. Na–H exchange-dependent increase in intracellular pH times G<sub>i</sub>/M entry and transition. *J. Biol. Chem.* 278:44645–44649. <http://dx.doi.org/10.1074/jbc.M308099200>
- Putney, L.K., S.P. Denker, and D.L. Barber. 2002. The changing face of the Na<sup>+</sup>/H<sup>+</sup> exchanger, NHE1: structure, regulation, and cellular actions. *Annu. Rev. Pharmacol. Toxicol.* 42:527–552. <http://dx.doi.org/10.1146/annurev.pharmtox.42.092001.143801>
- Rossano, A.J., A.K. Chouhan, and G.T. Macleod. 2013. Genetically encoded pH-indicators reveal activity-dependent cytosolic acidification of *Drosophila* motor nerve termini in vivo. *J. Physiol.* 591:1691–1706. <http://dx.doi.org/10.1113/jphysiol.2012.248377>
- Schönichen, A., B.A. Webb, M.P. Jacobson, and D.L. Barber. 2013. Considering protonation as a posttranslational modification regulating protein structure and function. *Annu. Rev. Biophys.* 42:289–314. <http://dx.doi.org/10.1146/annurev-biophys-050511-102349>
- Schreiber, R. 2005. Ca<sup>2+</sup> signaling, intracellular pH and cell volume in cell proliferation. *J. Membr. Biol.* 205:129–137. <http://dx.doi.org/10.1007/s00232-005-0778-z>
- Simons, M., W.J. Gault, D. Gotthardt, R. Rohatgi, T.J. Klein, Y. Shao, H.-J. Lee, A.-L. Wu, Y. Fang, L.M. Satlin, et al. 2009. Electrochemical cues regulate assembly of the Frizzled/Dishevelled complex at the plasma membrane during planar epithelial polarization. *Nat. Cell Biol.* 11:286–294. <http://dx.doi.org/10.1038/ncb1836>
- Srivastava, J., G. Barreiro, S. Groscurth, A.R. Gingras, B.T. Goult, D.R. Critchley, M.J.S. Kelly, M.P. Jacobson, and D.L. Barber. 2008. Structural model and functional significance of pH-dependent talin–actin binding for focal adhesion remodeling. *Proc. Natl. Acad. Sci. USA*. 105:14436–14441. <http://dx.doi.org/10.1073/pnas.0805163105>
- Tworoger, M., M.K. Larkin, Z. Bryant, and H. Ruohola-Baker. 1999. Mosaic analysis in the *Drosophila* ovary reveals a common hedgehog-inducible precursor stage for stalk and polar cells. *Genetics*. 151:739–748.

- Webb, B.A., M. Chimenti, M.P. Jacobson, and D.L. Barber. 2011. Dysregulated pH: a perfect storm for cancer progression. *Nat. Rev. Cancer*. 11:671–677. <http://dx.doi.org/10.1038/nrc3110>
- Wolfe, D.M., J.-H. Lee, A. Kumar, S. Lee, S.J. Orenstein, and R.A. Nixon. 2013. Autophagy failure in Alzheimer's disease and the role of defective lysosomal acidification. *Eur. J. Neurosci*. 37:1949–1961. <http://dx.doi.org/10.1111/ejn.12169>
- Ying, Q.-L., J. Wray, J. Nichols, L. Battle-Morera, B. Doble, J. Woodgett, P. Cohen, and A. Smith. 2008. The ground state of embryonic stem cell self-renewal. *Nature*. 453:519–523. <http://dx.doi.org/10.1038/nature06968>
- Young, R.A. 2011. Control of the embryonic stem cell state. *Cell*. 144:940–954. <http://dx.doi.org/10.1016/j.cell.2011.01.032>
- Yun, S.P., J.M. Ryu, M.O. Kim, J.H. Park, and H.J. Han. 2012. Rapid actions of plasma membrane estrogen receptors regulate motility of mouse embryonic stem cells through a profilin-1/cofilin-1-directed kinase signaling pathway. *Mol. Endocrinol*. 26:1291–1303. <http://dx.doi.org/10.1210/me.2012-1002>
- Zhang, Y., and D. Kalderon. 2000. Regulation of cell proliferation and patterning in *Drosophila* oogenesis by Hedgehog signaling. *Development*. 127:2165–2176.