

Golgi targeting of *Drosophila melanogaster* β 4GalNAcTB requires a DHHC protein family-related protein as a pilot

Anita Johswich,¹ Benjamin Kraft,¹ Manfred Wuhrer,² Monika Berger,¹ André M. Deelder,² Cornelis H. Hokke,² Rita Gerardy-Schahn,¹ and Hans Bakker¹

¹Department of Cellular Chemistry, Hannover Medical School, 30625 Hannover, Germany

²Department of Parasitology, Leiden University Medical Center, 2300 RC Leiden, Netherlands

D*rosophila melanogaster* β 4GalNAcTB mutant flies revealed that this particular *N*-acetylgalactosaminyltransferase is predominant in the formation of lacdiNAc (GalNAc β 1,4GlcNAc)-modified glycolipids, but enzymatic activity could not be confirmed for the cloned enzyme. Using a heterologous expression cloning approach, we isolated β 4GalNAcTB together with β 4GalNAcTB pilot (GABPI), a multimembrane-spanning protein related to Asp-His-His-Cys (DHHC) proteins but lacking the DHHC consensus sequence. In the absence of GABPI, inactive β 4GalNAcTB is trapped in the endoplasmic reticulum (ER). Coexpression of β 4GalNAcTB and GABPI gen-

erates the active enzyme that is localized together with GABPI in the Golgi. GABPI associates with β 4GalNAcTB and, when expressed with an ER retention signal, holds active β 4GalNAcTB in the ER. Importantly, treatment of isolated membrane vesicles with Triton X-100 disturbs β 4GalNAcTB activity. This phenomenon occurs with multimembrane-spanning glycosyltransferases but is normally not a property of glycosyltransferases with one membrane anchor. In summary, our data provide evidence that GABPI is required for ER export and activity of β 4GalNAcTB.

Introduction

Glycosylation in the secretory pathway is a complex process in which hundreds of glycosyltransferases are involved (Taniguchi et al., 2002). Many glycosyltransferases appear in gene families specified mainly by the nature of the nucleotide sugar donor (Coutinho et al., 2003). Within a given family, individual glycosyltransferases differ regarding the recognized acceptor structures. To understand and modulate cellular glycosylation pathways, it is important to know how this substrate specificity is generated (de Graffenried and Bertozzi, 2004).

Important in this respect is the observation that acceptor specificity in many glycosyltransferases is not restricted to recognition of one or a few specifically linked monosaccharides. Some protein-specific glycosyltransferases obtain additional selectivity by recognizing specific peptide motifs in the acceptor. A classic example is the *N*-acetylgalactosaminyltransferase

(GalNAcT), which modifies glycoprotein hormones with high selectivity (Smith and Baenziger, 1988).

Some glycosyltransferases require other proteins that are not part of the acceptor structure for their specific activity. β 1,4-galactosyltransferase (β 4GalT) acts on terminally positioned *N*-acetylglucosamine (GlcNAc) residues conjugated to proteins or lipids. Its specificity changes if it builds a complex with α -lactalbumin. In the complex, free glucose is used as an acceptor, and lactose is formed (Brew et al., 1968). In the case of core 1 β 3-galactosyltransferase (C1 β 3GalT), a molecular chaperone called Cosmc, with specificity for this single client, is required for folding and transportation to the Golgi (Ju and Cummings, 2002, 2005; Ju et al., 2008). Also, for O-mannosylation, two proteins, POMT1 and POMT2, are required (Manya et al., 2004). However, in this case, a two-protein enzymatic complex is proposed. The same is true in heparin sulfate biosynthesis in

Correspondence to Hans Bakker: bakker.hans@mh-hannover.de

Abbreviations used in this paper: Cer, ceramide; dsRNA, double-stranded RNA; GABPI, β 4GalNAcTB pilot; Gal, galactose; GalNAc, *N*-acetylgalactosamine; GalNAcT, *N*-acetylgalactosaminyltransferase; GlcNAc, *N*-acetylglucosamine; MALDI, matrix-assisted laser desorption/ionization; MS, mass spectrometry; pNP, *p*-nitrophenyl; TOF, time of flight.

© 2009 Johswich et al. This article is distributed under the terms of an Attribution-Noncommercial-Share Alike-No Mirror Sites license for the first six months after the publication date [see <http://www.jcb.org/misc/terms.shtml>]. After six months it is available under a Creative Commons License [Attribution-Noncommercial-Share Alike 3.0 Unported license, as described at <http://creativecommons.org/licenses/by-nc-sa/3.0/>].

which two different exostosins are required for efficient biosynthesis (McCormick et al., 2000).

For several glycosyltransferases involved in glycolipid biosynthesis, data indicate that factors other than the enzyme and the acceptor substrate play a role. This is the case for β 4GalT-V and -VI, which are homologues of the β 4GalT mentioned in the previous paragraph and of *Drosophila melanogaster* β 4GalNAcTB (the subject of this study). Under in vitro conditions, β 4GalT-V and -VI transfer galactose (Gal) into β 1-4 linkage to terminally expressed GlcNAc residues on glycoproteins (van Die et al., 1999; Guo et al., 2001). However, their involvement in the biosynthesis of lactosyl ceramide (Cer) by Gal transfer onto glucosyl Cer has been demonstrated as well (Nomura et al., 1998; Sato et al., 2000; Kolmakova and Chatterjee, 2005). At least in the case of galactosyltransferase V, this latter activity depends on the enzyme's anchorage in the membrane (van Die et al., 1999; Sato et al., 2000). Other enzymes involved in glycolipid biosynthesis have been shown to exhibit very low (de Vries et al., 1995; Zhu et al., 1998; Togayachi et al., 2001) or no (Steffensen et al., 2000; Schwientek et al., 2002) activity if expressed as soluble proteins. In general, very little is known about how lipid acceptors are recognized by glycosyltransferases. However, it has been suggested that a membrane-bound activator protein is required to present glycolipid acceptors to the modifying glycosyltransferases (Ramakrishnan et al., 2002). This hypothesis is substantiated by analogy to the lysosomal sphingolipid degradation machinery in which the sphingolipid activator protein presents the glycolipid substrates to glycosidases (Kolter and Sandhoff, 2005).

In this study, we describe a novel mechanism of glycosyltransferase maturation and functionalization for the glycolipid-specific β 4GalNAcTB from *Drosophila*. This enzyme, which has been described as an inactive homologue of β 4GalNAcTA in a previous study (Haines and Irvine, 2005), is a member of the invertebrate branch of the β 4GalT family involved in the biosynthesis of the lacdiNAc (GalNAc β 1,4GlcNAc) epitope (Kawar et al., 2002; Vadaie et al., 2002; Haines and Irvine, 2005; Stolz et al., 2008). Because β 4GalNAcT had not been cloned when this study was started, we searched for the corresponding activity using expression cloning (Bakker et al., 1997, 2005; Münster et al., 1998). In a heterologous approach, a cDNA library from *Drosophila* was used for expression in CHO cells, whereas formation of the lacdiNAc epitope was monitored with a specific monoclonal antibody (van Remoortere et al., 2000). As will be demonstrated in this study, the expression of two cDNA clones was required to install the functionally active enzyme.

Results

Expression cloning of a *Drosophila* β 4GalNAcT

Other than in many invertebrates, the lacdiNAc element has been identified on only a few glycoconjugates in mammals (Sato et al., 2003). We ascertained that CHO cells are negative for lacdiNAc. Considering that in mammalian cells terminal GlcNAc residues are recognized by several galactosyltransferases, we additionally hypothesized that signals in the complementation cloning approach could be improved by the use of CHO Lec8

cells. They lack the Golgi UDP-Gal transporter and, consequently, show drastically reduced incorporation of Gal in glycans (Deutscher and Hirschberg, 1986). A cDNA library was constructed from *Drosophila*, subdivided into pools, and, in an established sibling selection procedure (Bakker et al., 1997), used to search for clones that rendered cells positive for lacdiNAc. Cell surface lacdiNAc expression was monitored with antibody 259-2A1, which was originally raised from *Schistosoma mansoni*-infected mice (van Remoortere et al., 2000). In this procedure, it became obvious that two cDNA clones were required for the expression of the lacdiNAc epitope. Although clone one was a member of the β 1,4GalT family (van Die et al., 1997), the second clone (flybase CG17257) encoded a type III membrane protein that was related to a gene family referred to as Asp-His-His-Cys (DHHC) proteins (Mitchell et al., 2006). This protein was termed β 4GalNAcTB pilot (GABPI) to describe its crucial role in generating a functionally active β 4GalNAcTB as shown in the following experiments. The identified β 4GalNAcT was identical to the inactive β 4GalNAcTB recently cloned by Haines and Irvine (2005) in a homology-based approach. In agreement with this study, we found no in vivo activity for full-length β 4GalNAcTB expressed in mammalian cells. It is remarkable that expression cloning identified β 4GalNAcTB and not the homologous β 4GalNAcTA, which was shown in an earlier study (Haines and Irvine, 2005) to be an active enzyme. To resolve the controversial finding, β 4GalNAcTA was cloned by PCR and expressed in comparison with β 4GalNAcTB–GABPI in CHO and HEK293 cells. These experiments demonstrated that the cell surface lacdiNAc expression detected with antibody 259-2A1 as a result of β 4GalNAcTA was much lower than the lacdiNAc formation after the combined expression of β 4GalNAcTB and GABPI. Thus, the data demonstrate the existence of two functionally active β 4GalNAcTs (β 4GalNAcTA and β 4GalNAcTB) in *Drosophila* of which β 4GalNAcTB needs the cooperation of GABPI. Because CHO cells demonstrated a low tolerance to expression of the *Drosophila* *N*-acetylgalactosamine (GalNAc) transferases and HEK293 cells turned out to be a more suitable expression system, subsequent experiments were performed exclusively in HEK293 cells.

β 4GalNAcTB specifically modifies glycolipids

Despite elaborated analyses of *Drosophila* glycoproteins (North et al., 2006), the lacdiNAc structure has so far only been found as a modification of glycolipids (Seppo et al., 2000). With both cloned enzymes at hand, we evaluated the question of acceptor specificity. HEK293 cells were transfected with β 4GalNAcTA, β 4GalNAcTB, or the combination β 4GalNAcTB–GABPI and analyzed for the presence of lipid- and protein-bound lacdiNAc using TLC followed by immunoverlay (Fig. 1 A) and Western blotting (Fig. 1 B), respectively. In both systems, *Drosophila* S2 cells, which are naturally positive for the antibody epitope, and HEK293 cells transfected with *Caenorhabditis elegans* β 4GalNAcT (Kawar et al., 2002) were used as controls. Although expression of the *C. elegans* enzyme confirmed the availability of β 4GalNAcT acceptors on proteins, the absence of specific signals in both HEK293 cells transfected with the β 4GalNAcTs

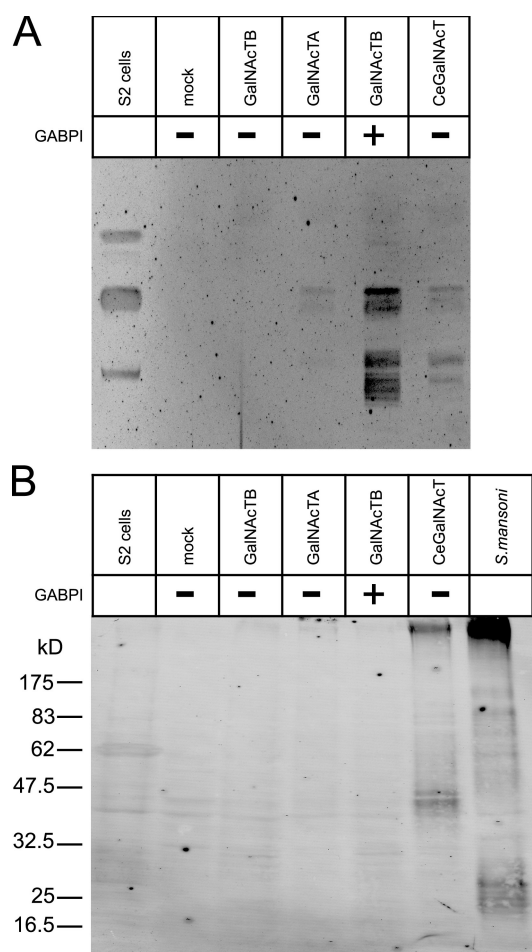


Figure 1. *Drosophila*-derived $\beta 4$ GalNAcTs are specific for glycolipids. (A) Glycolipid extracts from S2 cells and HEK293 cells after transfection with empty vector (mock) and GalNAcTs were separated on TLC and immunoblotted with antibody 259-2A1. (B) Protein extracts from cells as described in A were analyzed by Western blotting with 259-2A1. A protein extract from *S. mansoni* eggs was loaded as a control.

and in S2 cells confirmed the earlier observations in flies. In contrast, immunostaining of the lipid extracts resulted in positive signals for S2 cells as well as for HEK293 cells transfected with the $\beta 4$ GalNAcTB–GABPI pair. Expression of $\beta 4$ GalNAcTB alone was not sufficient to produce a signal, whereas faint signals were reliably obtained with $\beta 4$ GalNAcTA. It is important to mention that lipid specificity is preserved, although the glycolipid acceptor structures are different in *Drosophila* and HEK293 cells.

In vitro activity of $\beta 4$ GalNAcTB–GABPI is detergent sensitive

The observations that $\beta 4$ GalNAcTB is inactive if it is separately expressed in a heterologous cell system or if it is tested as a recombinant soluble protein (Haines and Irvine, 2005) encouraged further analyses to examine at which step $\beta 4$ GalNAcTB and GABPI interact with each other in the biosynthesis. In the first experiment, it was established that a soluble secreted construct of $\beta 4$ GalNAcTB was still inactive when coexpressed with GABPI. Subsequently, we wondered whether the two proteins expressed in separate cells have the capacity to form an

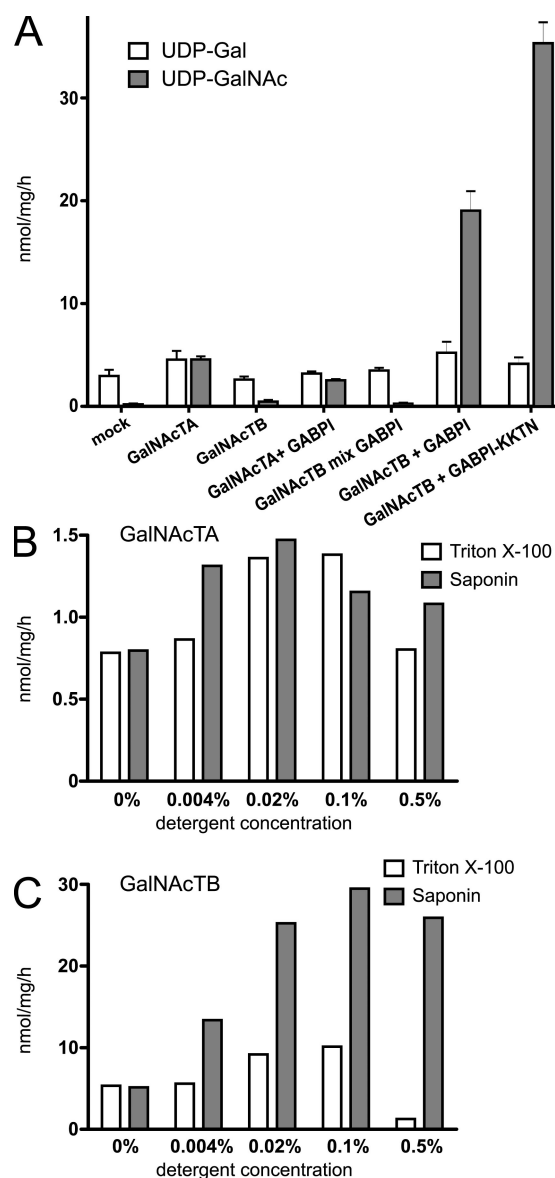


Figure 2. Activity of $\beta 4$ GalNAcTB is disrupted by Triton X-100. (A) Microsomal preparations of HEK293 transfected with $\beta 4$ GalNAcTs and GABPI as indicated were assayed for activity with GlcNAc-pNP as an acceptor and [3 H]UDP-GalNAc or [3 H]UDP-Gal (endogenous activity as an internal control) as donor substrates. Each value represents the mean of three independent vesicle preparations with standard deviation. GalNAcTB mix GABPI indicates that the proteins were expressed separately but mixed afterward for assays. (B and C) Microsomal fractions of HEK293 cells transfected with $\beta 4$ GalNAcTA (B) or the combination $\beta 4$ GalNAcTB–GABPI (C) were treated with Triton X-100 and saponin in various concentrations and assayed for GalNAcT activity as in A.

active enzyme. Microsomal fractions of HEK293 cells transfected with either $\beta 4$ GalNAcTB or GABPI were isolated, mildly treated with detergent (saponin 0.01%), and functionally tested in mixtures. The assay system used to follow $\beta 4$ GalNAcT activity was adapted from an established assay (Palcic et al., 1988). In this assay system, [3 H]UDP-GalNAc is the donor, and GlcNAc-*p*-nitrophenyl (pNP) is the acceptor substrate. No GalNAc transfer was measured in mixed vesicles (Fig. 2A), whereas controls with microsomes of $\beta 4$ GalNAcTA– or $\beta 4$ GalNAcTB–GABPI-transfected HEK293 cells were active after

identical treatment with detergent. In accordance with lacdiNAc formation in intact cells, β 4GalNAcTB was not active when expressed alone, but it showed higher activity than β 4GalNAcTA when expressed in combination with GABPI.

In contrast to β 4GalNAcTA, the β 4GalNAcTB activity strongly depended on the detergent used. Only background levels were measured if membranes were treated with Triton X-100 (Fig. 2 B) or NP-40 (not depicted) at 0.5%, which is routinely used in glycosyltransferase assays. The milder detergent saponin increased activity over a wide concentration range (Fig. 2 B). The rather low activity measured in the absence of detergent was probably a result of limited transport of the substrates over the vesicle membranes. As saponin is known to perforate and not disrupt membranes (Schulz, 1990), these data suggest that the maintenance of protein complexes in intact membrane patches is required for β 4GalNAcTB activity.

ER export of β 4GalNAcTB requires GABPI

The data presented so far for the interaction between β 4GalNAcTB and GABPI are highly reminiscent of the interactions between the human C1 β GalT generating the T antigen (core 1 O-glycan Gal β 1-3GalNAc α 1-Ser/Thr) and its client-specific molecular chaperone, Cosmc (Ju and Cummings, 2002). Cosmc supports functional folding of C1 β GalT in the ER but then dissociates and releases C1 β GalT (Ju et al., 2002b; Ju et al., 2008). Therefore, the following experiments addressed the subcellular localization of GABPI and β 4GalNAcTs. Flag- β 4GalNAcTA and Myc-GABPI were separately expressed in HEK293 cells and, after selection of stable clones, were detected by indirect immunofluorescence. Flag- β 4GalNAcTA and Myc-GABPI colocalized with the Golgi marker α -mannosidase II (Fig. 3, A and C). Only the signal generated by Flag- β 4GalNAcTB overlapped with the ER marker calnexin (Fig. 3 B). However, when GABPI was cotransfected (Fig. 3, D–F), the immunofluorescence images showed a clear shift of β 4GalNAcTB to the Golgi. Moreover, as shown in Fig. 3 F, GABPI and β 4GalNAcTB colocalized in this compartment. This experiment demonstrated that ER export of β 4GalNAcTB needs piloting by GABPI, which by itself is an autonomous protein fully equipped with the information required for folding and transport to the Golgi.

As GABPI moves with β 4GalNAcTB, the question was raised whether both proteins remain associated in the Golgi. To answer this question, GABPI was tagged with a C-terminal KKTN dilysine signal (Zerangue et al., 2001), which retains proteins in the ER. GABPI was indeed successfully localized in the ER using this approach (Fig. 3 G). More importantly, β 4GalNAcTB was also retained in the ER in cells expressing KKTN-tagged GABPI (Fig. 3 H). In vitro enzymatic activity of β 4GalNAcTB was about two times as high as the nonretained construct (Fig. 2 A, right bars), and cell surface lacdiNAc was also detectable in these cells. Although the latter probably required cycling to the Golgi of at least part of the enzyme, ER retention might have allowed a higher protein expression level that was enzymatically active in vitro. Together, these data demonstrated that the DHHC family-related protein has, in contrast to Cosmc, functions that go beyond those of a client-specific chaperone.

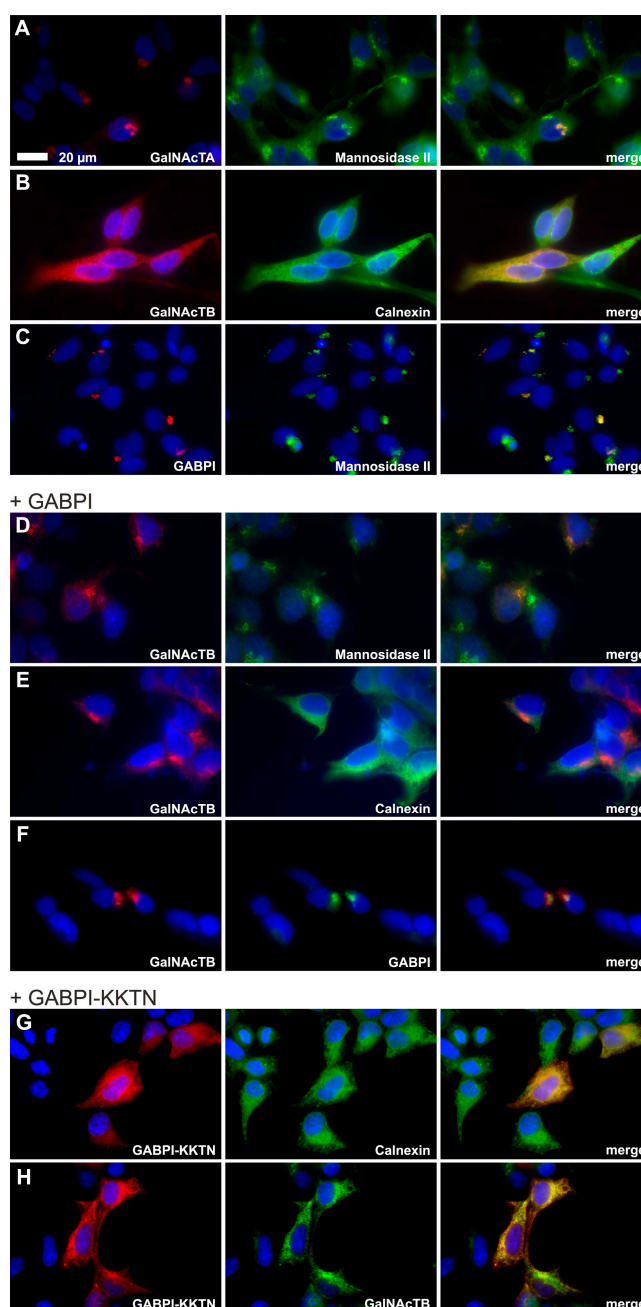


Figure 3. The Golgi resident protein GABPI pilots β 4GalNAcTB to the Golgi apparatus. The subcellular localization of β 4GalNAcTs and GABPI was analyzed with N-terminally tagged proteins expressed in HEK293 cells. (A–C) Flag- β 4GalNAcTA, -B, and Myc-GABPI were visualized by indirect immunofluorescence (red). Subcellular compartments were labeled with anti- α -mannosidase II (Golgi) or calnexin (ER; green). (D–F) Flag- β 4GalNAcTB (red) coexpressed with Myc-GABPI (F, green). (G) Subcellular localization of Myc-GABPI containing an N-terminal KKTN sequence. (H) Flag- β 4GalNAcTB (green) coexpressed with Myc-GABPI-KKTN (red). Nuclei were stained with Hoechst 33258 (blue).

Depletion of GABPI in *Drosophila* S2 cells delocalizes GalNAcTB and reduces lacdiNAc-containing glycolipid formation

To additionally evaluate the influence of GABPI on β 4GalNAcTB localization in the natural environment, RNAi experiments were performed. S2 cells were transiently transfected

with N-terminally tagged $\beta 4\text{GalNAcTs}$, and localization of the enzymes was monitored. As shown in Fig. 4 A, both $\beta 4\text{GalNAcTs}$ were colocalized in vesicular structures presumed to be the Golgi. $\beta 4\text{GalNAcTB}$ did not show any overlap with the ER-specific antibody anti-HDEL (Fig. 4 B). Incubation of cells with double-stranded RNA (dsRNA; Clemens et al., 2000) corresponding to a central coding region of GABPI dissected the HA- $\beta 4\text{GalNAcTB}$ signal from Flag- $\beta 4\text{GalNAcTA}$ (Fig. 4 C) and shifted the signal to structures that are part of the ER (Fig. 4 D).

To answer whether the RNAi-induced redistribution of the enzyme is also followed by a change in activity, a second knockdown experiment was performed in which dsRNAs were designed to down-regulate $\beta 4\text{GalNAcTA}$, $\beta 4\text{GalNAcTB}$, or GABPI. S2 cells were cultured for 3 d in the absence or presence of dsRNA, after which the expression of lacdiNAc was displayed by immunocytochemistry and matrix-assisted laser desorption/ionization (MALDI) time of flight (TOF) mass spectrometry (MS), as illustrated in Fig. 5. The intense staining of control cells with antibody 259-2A1 was in accordance with the detection of lacdiNAc-containing glycolipid structures by negative-ion mode MALDI-TOF-MS. $\beta 4\text{GalNAcTA}$ knockdown did not change the signal pattern in comparison with control cells, whereas depletion of either $\beta 4\text{GalNAcTB}$ or GABPI had comparably strong effects on lacdiNAc expression. In the negative-ion mode, reduction of glycolipids carrying lacdiNAc repeats was accompanied by an enrichment of GlcNAc β ,3Gal β ,3GalNAc α ,4GalNAc β ,4(PE-6)GlcNAc β ,3Man β ,4Glc β Cer species with a molecular mass of 1,958.4 D, an acceptor structure for $\beta 4\text{GalNAcT}$. The positive-ion mode analyses clearly demonstrated the accumulation of a second $\beta 4\text{GalNAcT}$ acceptor structure, GlcNAc β ,3Man β ,4Glc β Cer, having a molecular mass of 1,087.6 D. Changes in the glycolipid structures are very similar to changes observed in $\beta 4\text{GalNAcTB}$ knockout flies (Stolz et al., 2008) and thus are not further addressed in this paper. In addition, a new glycolipid species carrying lacdiNAc repeats with a molecular mass of 1,796.3 D has been identified and characterized by MALDI-TOF/TOF-MS (Table S1 and Figs. S1 and S2, available at <http://www.jcb.org/cgi/content/full/jcb.200801071/DC1>) as well as two extended species (Table S1 and Fig. S3). In summary, the results presented in Figs. 4 and 5 allow the conclusions that (a) $\beta 4\text{GalNAcTB}$ is the major lacdiNAc-synthesizing enzyme in S2 cells as it is in the fly, (b) GABPI enables Golgi targeting of $\beta 4\text{GalNAcTB}$, and (c) $\beta 4\text{GalNAcTB}$ is essentially required to convey functionality.

Pull down of the complex formed between $\beta 4\text{GalNAcTB}$ and GABPI

Because the data shown so far consistently argue for the existence of $\beta 4\text{GalNAcTB}$ and GABPI as a complex in the Golgi, we examined this contact in pull-down experiments. HEK293 cells transiently transfected with cDNA constructs encoding Myc-GABPI-HA and Flag- $\beta 4\text{GalNAcTA}$ and -B were lysed with buffer containing 1% NP-40. The anti-HA antibody 12CA5 coupled to Sepharose beads was used to precipitate Myc-GABPI-HA. To control the expression of recombinant proteins, total cell lysates were analyzed by Western blotting in parallel to precipitated proteins (Fig. 6). Both proteins were well ex-

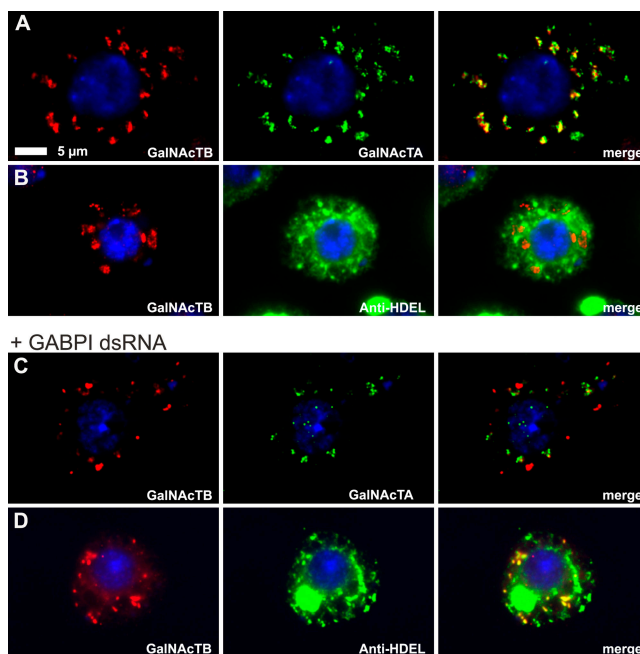


Figure 4. Knockdown of GABPI in S2 cells interferes with Golgi localization of $\beta 4\text{GalNAcTB}$. (A–D) S2 cells after 3 d of culture in the absence (A and B) or presence (C and D) of dsRNA directed against GABPI were transiently transfected with Flag- $\beta 4\text{GalNAcTA}$ and HA- $\beta 4\text{GalNAcTB}$. The Flag and HA epitopes were detected 2 d after transfection using respective antibodies, and the ER was marked with anti-HDEL. Nuclei were stained with Hoechst 33258.

pressed, as shown in the total cell lysates, whereas only Flag- $\beta 4\text{GalNAcTB}$ and not Flag- $\beta 4\text{GalNAcTA}$ was precipitated via Myc-GABPI-HA. This provided additional evidence for a tight interaction between $\beta 4\text{GalNAcTB}$ and GABPI.

The DHHC family-related protein GABPI is not an acyltransferase

Characterized DHHC protein family members are palmitoyl-transferases in which the cysteine residue in the conserved DHHC motif is essential for activity (Lobo et al., 2002; Roth et al., 2002; Valdez-Taubas and Pelham, 2005). GABPI, in contrast to all mammalian and the other *Drosophila* members in the family, has exchanged this motif from DHHC to DHHS. This was already an argument against its function as acyltransferase. To validate this assumption, a series of mutants was constructed with which a potential involvement of the DHHS motif in GABPI functions could be tested. The ability of GABPI to install a functional $\beta 4\text{GalNAcTB}$ was not abolished by reconstruction of the DHHC motif, by replacement of the serine by alanine, or by successive replacement to AAAA (unpublished data). In addition, the critical cysteine residue (C29) that may serve as acyl residue acceptor in $\beta 4\text{GalNAcTB}$ was mutated. Again, no effect on GABPI- $\beta 4\text{GalNAcTB}$ Golgi localization and activity was found (unpublished data).

The stem region of $\beta 4\text{GalNAcTB}$ is needed for activation by GABPI

Because the $\beta 4\text{GalNAcTs}$ isolated from *Drosophila* are highly homologous proteins, it was of relevance to identify primary sequence elements responsible for the strict GABPI dependency

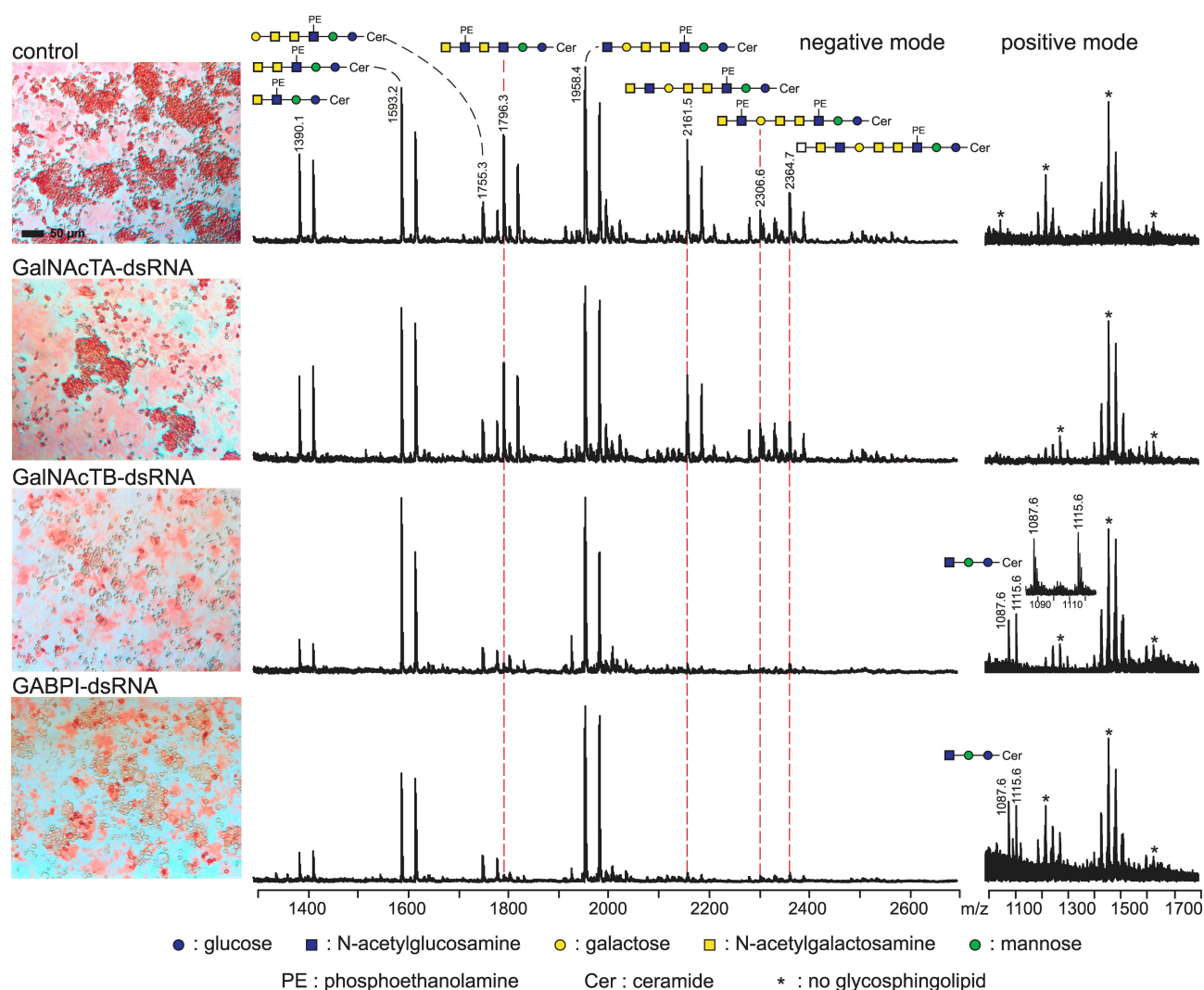


Figure 5. **Knockdown of GABPI in S2 cells abrogates $\beta 4$ GalNAcTB activity.** S2 cells before and after dsRNA down-regulation of $\beta 4$ GalNAcTs and GABPI as indicated were stained with antibody 259-2A1 to display lacdiNAc structures on the surface. Extracted glycolipid fractions were analyzed by MALDI-TOF-MS in negative- and positive-ion mode. Glycolipid species carrying multiple lacdiNAc structures that show differential expression are connected by dashed red lines. The positive-mode spectra show accumulation of the $\beta 4$ GalNAcT trisaccharide precursor.

of $\beta 4$ GalNAcTB. The aligned primary sequences indicated the stem region to be the domain of highest variability. Consequently, hybrids were made by domain swapping, as shown in Fig. 7. The chimera in which cytoplasmic and transmembrane domains of $\beta 4$ GalNAcTA were added to stem and catalytic regions of $\beta 4$ GalNAcTB (hybrid A-B-B) remained GABPI dependent for Golgi localization (Fig. 7, A and B) and activity (not depicted). However, additional replacement of the stem region destroyed activation by GABPI. The resulting protein was inactive and retained in the ER (Fig. 7, C and D). Because the stem region in $\beta 4$ GalNAcTA is considerably longer than in $\beta 4$ GalNAcTB, additional constructs were prepared in which the size was trimmed from the N and C termini to the exact length of the $\beta 4$ GalNAcTB stem region. All constructs remained inactive (unpublished data), allowing the conclusion that information contained in the stem region of $\beta 4$ GalNAcTB is essential for its function. In contrast, $\beta 4$ GalNAcTA remained Golgi localized and active independently of GABPI when fused

to the cytoplasmic and stem region of $\beta 4$ GalNAcTB (construct B-B-A; Fig. 7 E). This is in agreement with the fact that the catalytic domain of $\beta 4$ GalNAcTA can be produced as soluble enzyme and, therefore, is an independent active entity.

Discussion

Using a heterologous expression cloning approach, we isolated $\beta 4$ GalNAcTB as the major enzyme responsible for the biosynthesis of lacdiNAc structures in *Drosophila*. In this study, this enzyme was demonstrated to depend on the cooperation of a multimembrane-spanning protein related to the DHHC protein family. To point out the complexity of its involvement in forming a functionally active $\beta 4$ GalNAcTB, it was called GABPI. GABPI was cloned simultaneously to $\beta 4$ GalNAcTB in a classical expression cloning approach. This demonstrates the power of this technique, which exclusively screens for activity. The fact that the second $\beta 4$ GalNAc transferase in *Drosophila*,

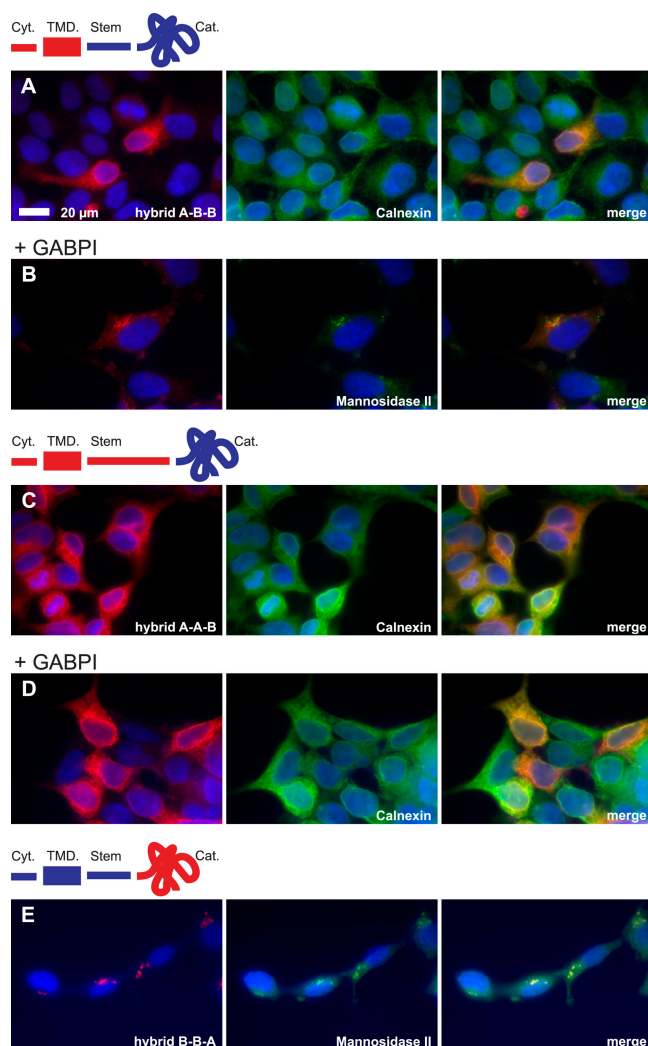


Figure 7. Protein domains of $\beta 4$ GalNAcTB involved in the interaction with GABPI. Hybrids of Flag- $\beta 4$ GalNAcTA (red) and -B (blue) were cloned by domain swapping of the cytoplasmic (Cyt) and transmembrane domain (TMD), stem region, and catalytic domain (Cat) as illustrated and expressed in HEK293 with and without GABPI. The intracellular localization was analyzed by indirect immunofluorescence using α -mannosidase II and calnexin as markers for Golgi and ER, respectively.

acyltransferase. All residues critical for a potential acyltransferase activity in GABPI (Mitchell et al., 2006) as well as the only cysteine residue that may serve as acyl acceptors in $\beta 4$ GalNAcTB were point mutated. None influenced the functionality of GABPI or activity of $\beta 4$ GalNAcTB. The functionally crucial cysteine in the name-giving DHHC motif is exchanged by serine in GABPI, which argues against its function as a palmitoyltransferase.

In experiments aimed at understanding how $\beta 4$ GalNAcTB and GABPI interact, we demonstrated that the selectivity with which GABPI activates $\beta 4$ GalNAcTB and not the highly homologous $\beta 4$ GalNAcTA is attributed to a structural element in the stem region. However, this area cannot be the solely responsible element. Additional sequences in the catalytic domain must be involved in determining GABPI dependency.

The exact function of GABPI in priming activity of $\beta 4$ GalNAcTB in the Golgi is difficult to address. However, several glycosyltransferases acting exclusively in the glycolipid bio-

synthetic pathways need membrane anchorage and cannot be expressed as soluble recombinant proteins (Amado et al., 1998; Steffensen et al., 2000; Schwientek et al., 2002). One of these enzymes is brainiac (Schwientek et al., 2002), a $\beta 3$ GlcNAc transferase acting right upstream of $\beta 4$ GalNAcTB in glycolipid biosynthesis of *Drosophila*. The factors determining membrane dependency of brainiac are not yet identified. Because we found the product of brainiac accumulated in S2 cells treated with RNAi against GABPI (Fig. 5), an involvement of GABPI for brainiac function in vivo is not likely. However, because mammalian lipid-modifying enzymes have been suggested to form multienzyme complexes (Giraud and Maccioni, 2003), GABPI, being an essential part of $\beta 4$ GalNAcTB, might be an anchor position in the pathway without being essential for the activity of all enzymes. A striking parallel exists between $\beta 4$ GalNAcTB and $\beta 4$ GalT-V and -VI described in the Introduction. These mammalian galactosyltransferases are members of the same gene family and are essentially dependent on membrane contact for transfer of Gal onto glucosyl Cer (van Die et al., 1999; Sato et al., 2000). As soluble enzymes, $\beta 4$ GalT-V and -VI recognize terminal GlcNAc residues instead of glucose. Therefore, it can be speculated that these enzymes require a cofactor similar to GABPI, which mediates glycolipid acceptor recognition. Orthologues of GABPI are found in arthropod and vertebrate species but not in nematodes, indicating that GABPI homologues might play a role in higher eukaryotes as well. In summary, it can be concluded that the identification of GABPI reveals a novel mechanism to generate specificity in the complex glycosylation pathway.

Materials and methods

Expression cloning

A cDNA library from *Drosophila* larval poly(A)⁺ RNA (Clontech Laboratories, Inc.) was constructed in pCMV-Script using the pCMV-ScriptXR cDNA library construction kit (Agilent Technologies). The library was divided into pools of 10,000 independent clones and used for expression cloning after the sibling selection strategy described previously (Bakker et al., 1997, 2005). The CHO cell line Lec8 (Deutscher and Hirschberg, 1986) grown in α -MEM supplemented with 10% FCS (both obtained from Biochrom AG) was used as the host. Pools or clones of the cDNA library were transfected into Lec8 cells using Metafectane (Biontex). After 2 d, cells grown in 6-well plates were fixed with 1.5% glutaraldehyde, incubated with the anti-dGalNAc monoclonal antibody 259-2A1 (van Remoortere et al., 2000) followed by HRP-conjugated goat anti-mouse antibody (Jackson ImmunoResearch Laboratories), and detected by tyramide signal amplification using biotin-tyramide (Speel et al., 2006), streptavidin-AP (Invitrogen), and Fast-Red (Sigma-Aldrich) as chromogenic substrate.

Plasmid constructs

All tagged mammalian expression constructs were made in pcDNA3 (Invitrogen). Myc-GABPI (flybase gene number CG17257) contains an N-terminal Myc tag (MAQKLISEEDLNIRPLE [antibody-bound sequence underlined]) and Myc-GABPI-HA, an additional C-terminal HA tag (SRYPYDVPDYASL). Flag- $\beta 4$ GalNAcTB (CG14517), Flag- $\beta 4$ GalNAcTA (CG8536), and *C. elegans* GalNAcT (Kawar et al., 2002) contain N-terminal Flag tags (MDYKDDDDKGS). The Myc-GABPI-KKTN construct was cloned by PCR using Myc-GABPI as a template. For expression in *Drosophila* S2 cells, Flag- $\beta 4$ GalNAcTA and HA- $\beta 4$ GalNAcTB (N-terminal HA tag; MYPYDVPDYAGS) were cloned in pB/V5-His (Invitrogen). Hybrids of $\beta 4$ GalNAcTA and $\beta 4$ GalNAcTB are identified by a three-letter code, whereby the first letter indicates the cytoplasmic plus transmembrane region, the second letter indicates the stem region, and the third letter indicates the catalytic domain (e.g., A-B-B). Borders between the three regions are after amino acids 29 and 135 in $\beta 4$ GalNAcTA and after 33 and 65 in $\beta 4$ GalNAcTB. Flag- or Myc-tagged constructs were used for all experiments unless indicated.

Preparation of ER and Golgi fractions from transfected HEK293 cells

HEK293 cells were grown in DME/HAM's F-12 supplemented with 10% FCS (both obtained from Biocrom AG). Cells transiently transfected as described in the Expression cloning section for CHO cells were washed with PBS and collected by centrifugation (5 min at 1,500 g). The cell pellets from three 175-cm² plates (9 × 10⁷ cells) were resuspended in 7 ml of lysis buffer (10 mM Hepes-Tris, pH 7.4, 0.8 M sorbitol, and 1 mM EDTA) containing an EDTA-free protease inhibitor mixture (Roche). After 10 strokes in a Dounce homogenizer, the lysate was centrifuged (10 min at 1,500 g). The supernatant was collected, and the pellet was subjected to a second homogenization/centrifugation round. The ER/Golgi-rich fraction was obtained by centrifugation of the combined supernatants at 100,000 g for 1 h. Pelleted vesicles were resuspended in 500 µl of assay buffer (0.1 M MOPS, pH 7.5) and 20-µl aliquots kept at −80°C. Protein concentrations were determined using a bicinchoninic acid kit (Thermo Fisher Scientific).

In vitro β4GalNAcT assays

Standard enzyme assays were performed with 20 µl of the ER/Golgi preparations in 50 µl of assay buffer (0.1 MOPS, pH 7.5, 20 mM MnCl₂, 10 mM ATP, 100 mM GalNAc, 0.1% BSA, and 0.01% saponin). Therefore, 20-µl aliquots of the ER/Golgi vesicle preparation were supplemented to obtain the appropriate buffer composition and 0.5 mM of the radio-labeled nucleotide sugars UDP-6[³H]Gal (specific activity of 32 Bq/nmol; GE Healthcare) or UDP-1[³H]GalNAc (specific activity of 36 Bq/nmol [PerkinElmer]; diluted with cold nucleotide sugars [Sigma-Aldrich]). Reactions were started by adding the acceptor substrate GlcNAc-pNP (Sigma-Aldrich) at 1 mM and were incubated for 2 h at 28°C. Control samples were incubated in the absence of GlcNAc-O-pNP and subtracted from measured values. Reactions were stopped by addition of 1 ml of ice-cold water, and products were isolated on columns (Sep Pak Plus C₁₈; Waters Corporation) as described previously (Palcic et al., 1988). The elutes were dried and counted in 2 ml of scintillation cocktail (Luma Safe Plus; Lumac LSC). Incorporated radioactivity was measured in a counter (LS 6500; Beckman Coulter).

Analyses of glycosphingolipids and proteins from transfected HEK293 cells

Transiently transfected HEK293 cells were washed with PBS, scraped off the plates, and collected by centrifugation (10 min at 1,500 g). *Drosophila* S2 cells were harvested by centrifugation and extracted in the same way. The cell pellets (10⁷ cells) were resuspended in 300 µl of water and sonicated for 5 min in a bath sonicator. 2-propanol and hexane were added to obtain a solvent ratio of 55:25:20 (2-propanol/hexane/water), and the mixtures were sonicated again for 5 min. Samples were centrifuged for 10 min at 1,500 g, and supernatants were dried under nitrogen. The extracts were resuspended in chloroform/methanol/water (3:47:48) and desalted by reverse-phase chromatography (Sep Pak Plus C₁₈ columns; Williams and McChuer, 1980). The eluted glycosphingolipids were dried under nitrogen, and one fourth of each sample was spotted onto a TLC plate (Nano-Durasil-20; Macherey-Nagel) and developed in running solvent composed of chloroform/methanol/0.25% aqueous KCl (5:4:1). For immunostaining, the silica plate was fixed in 0.1% polyisobutylmethacrylate (Sigma-Aldrich) in acetone. The plate was blocked overnight with 1% BSA in TBS at 4°C followed by incubation with primary antibody (mouse anti-lacdiNAc 259-2A1) for 2 h at room temperature and with secondary antibody goat anti-mouse IRDye 800 (LI-COR Biosciences) for 30 min. After washing, the plate was analyzed on an infrared imaging system (Odyssey; LI-COR Biosciences).

Protein samples for Western blotting were isolated from the same cells by dissolving 10⁷ cells in 750 µl of lysis buffer (2 mM EDTA, 50 mM Tris-HCl, pH 8.0, 1 mM MgCl₂, and 1% NP-40 supplemented with a protease inhibitor mixture [Roche]) and analyzing 20 µl of these samples by standard Western blotting techniques. The blot was incubated with the same antibodies as the TLC plate and analyzed in the same way.

Subcellular localization studies by immunofluorescence

Subcellular localizations of recombinant Flag-β4GalNAcTA, Flag-β4GalNAcTB, and Myc-GABPI were performed with stably transfected HEK293 cells (without selecting clones). Therefore, transfected cells were cultured for 3 wk in the presence of G-418 (EMD). For staining, cells were seeded onto glass coverslips, fixed in 4% PFA, and permeabilized for 30 min with 0.1% saponin in PBS containing 0.1% BSA. Samples were incubated with the respective primary antibodies (anti-Flag tag M5, anti-Flag tag F7425, anti-HA tag 12CA5 or anti-Myc tag 9E10, and rabbit anti-α-mannosidase II or -calnexin as a Golgi or ER marker) for 1.5 h at room temperature. After

three washings (in PBS, 0.1% BSA, and 0.1% Tween 20), cells were incubated with anti-mouse IgCy3 and anti-rabbit IgG Alexa Fluor 488 for 1 h at room temperature. After staining with the nuclear dye (Hoechst 33258; Hoechst Pharmaceuticals), the slides were washed with water, mounted (Dako), and analyzed under a microscope (Axiovert 200M; Carl Zeiss, Inc.) using a Plan Apochromat 63×/1.40 oil differential interference contrast objective (M27; Carl Zeiss, Inc.) at room temperature. Images (1,388 × 1,040 pixels) were taken using a camera (AxioCam MRm; Carl Zeiss, Inc.) and Axiovision 4.4 software (Carl Zeiss, Inc.). Images taken in an automatic exposure setting with filter sets for Hoechst 33258, Alexa Fluor 488, and Cy3 were converted in blue, green, and red, respectively; intensities were adapted to be equal for the three colors, and images were reduced to 600 dots per inch for display in Figs. 3 and 7.

Knockdown experiments in *Drosophila* S2 cells

dsRNA was made using the MEGAscript T7 transcription kit (Applied Biosystems). Each primer used in the PCR contained a 5' T7 RNA polymerase-binding site followed by sequences specific for the target genes: GABPI (5'-CCG-GCACCTCCAATTCTTTC-3' and 5'-GTCCATATCCCCACCTCGTCA-3'), β4GalNAcTA (5'-ATGTACCTCTTCACCAAGGCGA-3' and 5'-ATAACCAA-TGTCATCATGGCA-3'), and β4GalNAcTB (5'-TCAACTTTCTGCGCAA-CAATG-3' and 5'-ACCACGCCGCCGAAAAGACC-3').

Drosophila Schneider (S2) cells were grown in Schneider's *Drosophila* medium (Invitrogen) supplemented with 10% FCS and 4 mM L-glutamine (Biocrom AG). For RNAi knockdown experiments (Clemens et al., 2000), 10⁶ cells were plated per 6 wells in serum-free medium, and dsRNA of β4GalNAcTA, β4GalNAcTB, and/or GABPI was added directly to the media in a final concentration of 37 nM (15 µg). After 30 min at room temperature, 2 ml of Schneider's medium containing FCS was added, and incubation was continued for 3 d at 27°C. For the immunocytochemical analysis of surface-expressed lacdiNAc structures, the protocol described in the Expression cloning section for CHO cells was used. Light microscopic images of Fig. 5 were taken using the aforementioned microscope and software using a camera (AxioCam MRc; Carl Zeiss, Inc.) and a Plan Apochromat 10×/0.45 objective (Ph1M27; Carl Zeiss, Inc.). To determine the subcellular localization of β4GalNAcTA and β4GalNAcTB in GABPI dsRNA-treated cells, the N-terminally Flag- and HA-tagged enzymes were transiently transfected with Eugene (Roche) into cultures that had been treated for 3 d with dsRNA. The day after transfection, cells were washed with serum-free medium, and RNAi treatment was repeated with a concentration of 18.5 nM dsRNA. 2 d after transfection, cells were transferred to concanavalin A-coated coverslips for 1 h, fixed in 4% PFA, and further processed as described in the previous section for HEK293 cells except that 0.1% saponin was kept in all incubation and washing solutions. Mouse anti-Flag M5 (Sigma-Aldrich) in combination with rabbit anti-HA (Sigma-Aldrich) was used to visualize the tagged β4GalNAc transferases, whereas the ER was stained with mouse anti-HDEL (Santa Cruz Biotechnology, Inc.). Secondary antibodies used were goat anti-mouse, Alexa Fluor 488, and goat anti-rabbit Cy3. Fluorescent images were made using a microscope (Axiovert 200M) as for the aforementioned HEK293 cells except that the ApoTome mode was used and five images were averaged. Fig. 4 shows 330 × 236-pixel sections of the original images. In addition, glycosphingolipid extracts (prepared as described in Analyses of glycosphingolipids and proteins...) from S2 cells before and after dsRNA treatment were analyzed by MALDI-TOF-MS in a TOF/TOF mass spectrometer (Ultraflex II; Bruker Daltonics) as described previously (Wuhrer and Deelder, 2005; Stolz et al., 2008).

Immunoprecipitation

Transiently transfected HEK293 cells were lysed for 30 min at 4°C using 750 µl of lysis buffer (2 mM EDTA, 50 mM Tris-HCl, pH 8.0, 1 mM MgCl₂, and 1% NP-40 supplemented with protease inhibitor mixture). After centrifugation for 30 min at 12,000 g, anti-HA antibody 12CA5 coupled to Sepharose A beads was added to supernatants and incubated for 3 h at 4°C on a rotating wheel. Immunocomplexes were pelleted by centrifugation (300 g for 5 min) and washed twice with 50 mM Tris-HCl, pH 8.0, and 1% NP-40, twice with 50 mM Tris-HCl, pH 8.0, 500 mM NaCl, and 1% NP-40, and once with the first washing buffer. Immunoprecipitated proteins were separated in SDS-PAGE, blotted onto polyvinylidene difluoride membranes (Waters Corporation), and stained with mouse anti-Flag M5 or rat anti-HA antibody.

Online supplemental material

Table S1 shows newly registered zwitterionic glycosphingolipid species. Fig. S1 shows negative-mode MALDI-TOF-MS of S2 cell glycosphingolipids.

Fig. S2 shows MALDI-TOF/TOF-MS fragmentation analysis of two zwitterionic glycolipid species containing lacdiNAc tandem repeats. Fig. S3 shows MALDI-TOF/TOF-MS analysis of two zwitterionic glycolipid species. Online supplemental material is available at <http://www.jcb.org/cgi/content/full/jcb.200801071/DC1>.

We would like to thank Drs. Françoise Routier and Birgit Lüttig for critical reading of the manuscript.

Supporting financial resources for this study were obtained from the Hannover Medical School bonus system Leistungsorientierte Mittel and Regenerative Biology to Reconstructive Therapy, a Cluster of Excellence financed by the Deutsche Forschungsgemeinschaft.

Submitted: 11 January 2008

Accepted: 9 December 2008

References

- Amado, M., R. Almeida, F. Carneiro, S.B. Levery, E.H. Holmes, M. Nomoto, M.A. Hollingsworth, H. Hassan, T. Schwientek, P.A. Nielsen, et al. 1998. A family of human β 3-galactosyltransferases. Characterization of four members of a UDP-galactose: β -N-acetyl-glucosamine/ β -N-acetyl-galactosamine β -1,3-galactosyltransferase family. *J. Biol. Chem.* 273:12770–12778.
- Bakker, H., I. Friedmann, S. Oka, T. Kawasaki, N. Nifant'ev, M. Schachner, and N. Mantei. 1997. Expression cloning of a cDNA encoding a sulfotransferase involved in the biosynthesis of the HNK-1 carbohydrate epitope. *J. Biol. Chem.* 272:29942–29946.
- Bakker, H., F. Routier, S. Oelmann, W. Jordi, A. Lommen, R. Gerardy-Schahn, and D. Bosch. 2005. Molecular cloning of two *Arabidopsis* UDP-galactose transporters by complementation of a deficient Chinese hamster ovary cell line. *Glycobiology*. 15:193–201.
- Brew, K., T.C. Vanaman, and R.L. Hill. 1968. The role of α -lactalbumin and the A protein in lactose synthetase: a unique mechanism for the control of a biological reaction. *Proc. Natl. Acad. Sci. USA*. 59:491–497.
- Clemens, J.C., C.A. Worby, N. Simonson-Leff, M. Muda, T. Machama, B.A. Hemmings, and J.E. Dixon. 2000. Use of double-stranded RNA interference in *Drosophila* cell lines to dissect signal transduction pathways. *Proc. Natl. Acad. Sci. USA*. 97:6499–6503.
- Coutinho, P.M., E. Deleury, G.J. Davies, and B. Henrissat. 2003. An evolving hierarchical family classification for glycosyltransferases. *J. Mol. Biol.* 328:307–317.
- de Graffenried, C.L., and C.R. Bertozzi. 2004. The roles of enzyme localisation and complex formation in glycan assembly within the Golgi apparatus. *Curr. Opin. Cell Biol.* 16:356–363.
- de Vries, T., C.A. Srnka, M.M. Palcic, S.J. Swiedler, D.H. van den Eijnden, and B.A. Macher. 1995. Acceptor specificity of different length constructs of human recombinant α 1,3/4-fucosyltransferases. Replacement of the stem region and the transmembrane domain of fucosyltransferase V by protein A results in an enzyme with GDP-fucose hydrolyzing activity. *J. Biol. Chem.* 270:8712–8722.
- Deutscher, S.L., and C.B. Hirschberg. 1986. Mechanism of galactosylation in the Golgi apparatus. A Chinese hamster ovary cell mutant deficient in translocation of UDP-galactose across Golgi vesicle membranes. *J. Biol. Chem.* 261:96–100.
- Giraudo, C.G., and H.J. Maccioni. 2003. Ganglioside glycosyltransferases organize in distinct multienzyme complexes in CHO-K1 cells. *J. Biol. Chem.* 278:40262–40271.
- Guo, S., T. Sato, K. Shirane, and K. Furukawa. 2001. Galactosylation of N-linked oligosaccharides by human β -1,4-galactosyltransferases I, II, III, IV, V, and VI expressed in Sf-9 cells. *Glycobiology*. 11:813–820.
- Haines, N., and K.D. Irvine. 2005. Functional analysis of *Drosophila* β 1,4-N-acetylglucosaminyltransferases. *Glycobiology*. 15:335–346.
- Ju, T., and R.D. Cummings. 2002. A unique molecular chaperone Cosmc required for activity of the mammalian core 1 β 3-galactosyltransferase. *Proc. Natl. Acad. Sci. USA*. 99:16613–16618.
- Ju, T., and R.D. Cummings. 2005. Protein glycosylation: chaperone mutation in Tn syndrome. *Nature*. 437:1252.
- Ju, T., K. Brewer, A. D'Souza, R.D. Cummings, and W.M. Canfield. 2002a. Cloning and expression of human core 1 β 1,3-galactosyltransferase. *J. Biol. Chem.* 277:178–186.
- Ju, T., R.D. Cummings, and W.M. Canfield. 2002b. Purification, characterization, and subunit structure of rat core 1 β 1,3-galactosyltransferase. *J. Biol. Chem.* 277:169–177.
- Ju, T., R.P. Aryal, C.J. Stowell, and R.D. Cummings. 2008. Regulation of protein O-glycosylation by the endoplasmic reticulum-localized molecular chaperone Cosmc. *J. Cell Biol.* 182:531–542.
- Kawar, Z.S., I. van Die, and R.D. Cummings. 2002. Molecular cloning and enzymatic characterization of a UDP-GalNAc:GlcNAc(β)-R β 1,4-N-acetyl-galactosaminyltransferase from *Caenorhabditis elegans*. *J. Biol. Chem.* 277:34924–34932.
- Kolmakova, A., and S. Chatterjee. 2005. Platelet derived growth factor recruits lactosylceramide to induce cell proliferation in UDP Gal:GlcCer: β 1 \rightarrow 4Galactosyltransferase (GalT-V) mutant Chinese hamster ovary cells. *Glycoconj. J.* 22:401–407.
- Kolter, T., and K. Sandhoff. 2005. Principles of lysosomal membrane digestion: stimulation of sphingolipid degradation by sphingolipid activator proteins and anionic lysosomal lipids. *Annu. Rev. Cell Dev. Biol.* 21:81–103.
- Lobo, S., W.K. Greentree, M.E. Linder, and R.J. Deschenes. 2002. Identification of a Ras palmitoyltransferase in *Saccharomyces cerevisiae*. *J. Biol. Chem.* 277:41268–41273.
- Manya, H., A. Chiba, A. Yoshida, X. Wang, Y. Chiba, Y. Jigami, R.U. Margolis, and T. Endo. 2004. Demonstration of mammalian protein O-mannosyltransferase activity: coexpression of POMT1 and POMT2 required for enzymatic activity. *Proc. Natl. Acad. Sci. USA*. 101:500–505.
- McCormick, C., G. Duncan, K.T. Goutsos, and F. Tufaro. 2000. The putative tumor suppressors EXT1 and EXT2 form a stable complex that accumulates in the Golgi apparatus and catalyzes the synthesis of heparan sulfate. *Proc. Natl. Acad. Sci. USA*. 97:668–673.
- Mitchell, D.A., A. Vasudevan, M.E. Linder, and R.J. Deschenes. 2006. Protein palmitoylation by a family of DHHC protein S-acyltransferases. *J. Lipid Res.* 47:1118–1127.
- Münster, A.K., M. Eckhardt, B. Potvin, M. Mühlenhoff, P. Stanley, and R. Gerardy-Schahn. 1998. Mammalian cytidine 5'-monophosphate N-acetylneuraminic acid synthetase: a nuclear protein with evolutionarily conserved structural motifs. *Proc. Natl. Acad. Sci. USA*. 95:9140–9145.
- Nomura, T., M. Takizawa, J. Aoki, H. Arai, K. Inoue, E. Wakisaka, N. Yoshizuka, G. Imokawa, N. Dohmae, K. Takio, et al. 1998. Purification, cDNA cloning, and expression of UDP-Gal: glucosylceramide β -1,4-galactosyltransferase from rat brain. *J. Biol. Chem.* 273:13570–13577.
- North, S.J., K. Koles, C. Hembd, H.R. Morris, A. Dell, V.M. Panin, and S.M. Haslam. 2006. Glycomic studies of *Drosophila melanogaster* embryos. *Glycoconj. J.* 23:345–354.
- Palcic, M.M., L.D. Heerze, M. Pierce, and O. Hindsgaul. 1988. The use of hydrophobic synthetic glycosides as acceptors in glycosyltransferase assays. *Glycoconj. J.* 5:49–63.
- Ramakrishnan, B., E. Boeggeman, and P.K. Qasba. 2002. β -1,4-galactosyltransferase and lactose synthase: molecular mechanical devices. *Biochem. Biophys. Res. Commun.* 291:1113–1118.
- Roth, A.F., Y. Feng, L. Chen, and N.G. Davis. 2002. The yeast DHHC cysteine-rich domain protein Akr1p is a palmitoyl transferase. *J. Cell Biol.* 159:23–28.
- Sasaki, N., H. Yoshida, T.J. Fuwa, A. Kinoshita-Toyoda, H. Toyoda, Y. Hirabayashi, H. Ishida, R. Ueda, and S. Nishihara. 2007. *Drosophila* β 1,4-N-acetylglucosaminyltransferase-A synthesizes the LacdiNAc structures on several glycoproteins and glycosphingolipids. *Biochem. Biophys. Res. Commun.* 354:522–527.
- Sato, T., S. Guo, and K. Furukawa. 2000. Involvement of recombinant human β -1, 4-galactosyltransferase V in lactosylceramide biosynthesis. *Res. Commun. Biochem. Cell Mol. Biol.* 4:3–10.
- Sato, T., M. Gotoh, K. Kiyohara, A. Kameyama, T. Kubota, N. Kikuchi, Y. Ishizuka, H. Iwasaki, A. Togayachi, T. Kudo, et al. 2003. Molecular cloning and characterization of a novel human β 1,4-N-acetylglucosaminyltransferase, β 4GalNAc-T3, responsible for the synthesis of N,N'-diacetyl-lactosediamine, GalNAc β 1-4GlcNAc. *J. Biol. Chem.* 278:47534–47544.
- Schulz, I. 1990. Permeabilizing cells: some methods and applications for the study of intracellular processes. *Methods Enzymol.* 192:280–300.
- Schutzbach, J.S. 1997. The role of the lipid matrix in the biosynthesis of dolichyl-linked oligosaccharides. *Glycoconj. J.* 14:175–182.
- Schwientek, T., B. Keck, S.B. Levery, M.A. Jensen, J.W. Pedersen, H.H. Wandall, M. Stroud, S.M. Cohen, M. Amado, and H. Clausen. 2002. The *Drosophila* gene brainiac encodes a glycosyltransferase putatively involved in glycosphingolipid synthesis. *J. Biol. Chem.* 277:32421–32429.
- Seppo, A., M. Moreland, H. Schweingruber, and M. Tiemeyer. 2000. Zwitterionic and acidic glycosphingolipids of the *Drosophila melanogaster* embryo. *Eur. J. Biochem.* 267:3549–3558.
- Smith, P.L., and J.U. Baenziger. 1988. A pituitary N-acetylglucosamine transferase that specifically recognizes glycoprotein hormones. *Science*. 242:930–933.
- Speel, E.J., A.H. Hopman, and P. Komminoth. 2006. Tyramide signal amplification for DNA and mRNA in situ hybridization. *Methods Mol. Biol.* 326:33–60.
- Steffensen, R., K. Carlier, J. Wiels, S.B. Levery, M. Stroud, B. Cedergren, S.B. Nilsson, E.P. Bennett, C. Jersild, and H. Clausen. 2000. Cloning and

expression of the histo-blood group Pk UDP-galactose: Ga1 β -4Glc β 1-cer α 1, 4-galactosyltransferase. Molecular genetic basis of the p phenotype. *J. Biol. Chem.* 275:16723–16729.

- Stolz, A., N. Haines, A. Pich, K.D. Irvine, C.H. Hokke, A.M. Deelder, R. Gerardy-Schahn, M. Wuhrer, and H. Bakker. 2008. Distinct contributions of β 4GalNAcTA and β 4GalNAcTB to *Drosophila* glycosphingolipid biosynthesis. *Glycoconj. J.* 25:167–175.
- Taniguchi, N., K. Honke, and M. Fukuda. 2002. Handbook of Glycosyltransferases and Related Genes. Springer-Verlag New York Inc., New York. 670 pp.
- Togayachi, A., T. Akashima, R. Ookubo, T. Kudo, S. Nishihara, H. Iwasaki, A. Natsume, H. Mio, J. Inokuchi, T. Irimura, et al. 2001. Molecular cloning and characterization of UDP-GlcNAc:lactosylceramide β 1,3-N-acetylglucosaminyltransferase (β 3Gn-T5), an essential enzyme for the expression of HNK-1 and Lewis X epitopes on glycolipids. *J. Biol. Chem.* 276:22032–22040.
- Vadaie, N., R.S. Hulinsky, and D.L. Jarvis. 2002. Identification and characterization of a *Drosophila melanogaster* ortholog of human β 1,4-galactosyltransferase VII. *Glycobiology*. 12:589–597.
- Valdez-Taubas, J., and H. Pelham. 2005. Swf1-dependent palmitoylation of the SNARE Tlg1 prevents its ubiquitination and degradation. *EMBO J.* 24:2524–2532.
- van Die, I., H. Bakker, and D.H. van den Eijnden. 1997. Identification of conserved amino acid motifs in members of the β 1 \rightarrow 4-galactosyltransferase gene family. *Glycobiology*. 7:v–viii.
- van Die, I., A. van Tetering, W.E. Schiphorst, T. Sato, K. Furukawa, and D.H. van den Eijnden. 1999. The acceptor substrate specificity of human β 4-galactosyltransferase V indicates its potential function in O-glycosylation. *FEBS Lett.* 450:52–56.
- Van Remoortere, A., C.H. Hokke, G.J. van Dam, I. van Die, A.M. Deelder, and D.H. van den Eijnden. 2000. Various stages of *schistosoma* express Lewis(x), LacdiNAc, GalNAc β 1-4 (Fuc α 1-3)GlcNAc and GalNAc β 1-4(Fuc α 1-2Fuc α 1-3)GlcNAc carbohydrate epitopes: detection with monoclonal antibodies that are characterized by enzymatically synthesized neoglycoproteins. *Glycobiology*. 10:601–609.
- Wandall, H.H., J.W. Pedersen, C. Park, S.B. Levery, S. Pizette, S.M. Cohen, T. Schwientek, and H. Clausen. 2003. *Drosophila* egghead encodes a β 1,4-mannosyltransferase predicted to form the immediate precursor glycosphingolipid substrate for brainiac. *J. Biol. Chem.* 278:1411–1414.
- Williams, M.A., and R.H. McCluer. 1980. The use of Sep-Pak C18 cartridges during the isolation of gangliosides. *J. Neurochem.* 35:266–269.
- Wuhrer, M., and A.M. Deelder. 2005. Negative-mode MALDI-TOF/TOF-MS of oligosaccharides labeled with 2-aminobenzamide. *Anal. Chem.* 77:6954–6959.
- Zerangue, N., M.J. Malan, S.R. Fried, P.F. Dazin, Y.N. Jan, L.Y. Jan, and B. Schwappach. 2001. Analysis of endoplasmic reticulum trafficking signals by combinatorial screening in mammalian cells. *Proc. Natl. Acad. Sci. USA.* 98:2431–2436.
- Zhu, G., M.L. Allende, E. Jaskiewicz, R. Qian, D.S. Darling, C.A. Worth, K.J. Colley, and W.W. Young Jr. 1998. Two soluble glycosyltransferases glycosylate less efficiently in vivo than their membrane bound counterparts. *Glycobiology*. 8:831–840.