Sakurai et al. Vol. 154, No. 6, September 17, 2001. Pages 1259–1273.

Due to typographical errors the following corrections should be noted:

The seventh author's name was misspelled on page 1259. The corrected author line appears below:

Takeshi Sakurai, Marc Lustig, Joanne Babiarz, Andrew J.W. Furley, Steven Tait, Peter J. Brophy, Stephen A. Brown, Lucia Y. Brown, Carol A. Mason, and Martin Grumet

In the first row of Table IV "Nr-CAM 1/1" should read "Nr-CAM −/−." The corrected table appears below:

Table IV. **Measurement of thickness of EGL and IGL of Nr-CAM/L1 double knockout mice and their littermates**

	Layer	Lobe V	Lobe IX
Nr-CAM −/−, L1−/y[a]	EGL	42.5 ± 2.3 (70.6%)	31.6 ± 3.2 (90.8%)
	IGL	30.8 ± 3.0 (51.0%)	35.6 ± 4.8 (58.9%)
Littermates[a]	EGL	64.0 ± 3.2 (100%)	34.8 ± 2.6 (100%)
	IGL	60.4 ± 2.1 (100%)	60.4 ± 3.0 (100%)

Cerebellar midsagittal sections from P5–P6 Nr-CAM/L1 double knockout mice and their littermates were processed for Nissl staining and images were captured by CCD camera. Thickness of EGL and IGL were measured in lobe V and IX using NIH image. Numbers are average + SD μm. Numbers in parentheses are the percentage of littermate control.
[a]$n = 6$.

The following paragraph is being reprinted with two corrections. The proper gift reference for rabbit anti-TAG-1 antisera is now included; also, the provider of rabbit anti-Zic2 antisera has been removed:

General histology

Frozen sections from embryos and neonatal pups were prepared and immunohistochemistry was performed as described (Lustig et al., 2001). Antibodies used were 837/838, rabbit anti–Nr-CAM polyclonal antisera, (Lustig et al., 2001) 1:300; 371/372, rabbit anti-L1 polyclonal antisera (Lustig et al., 2001), 1:300; 4D7, mouse anti-TAG-1 monoclonal antibody (IgM, culture supernatant from Developmental Study Hybridoma Bank, University of Iowa, Iowa City, IA), 1:2; TG-3, rabbit anti-TAG-1 antisera (gift of Dr. Thomas Jessel and Susan Morton, Columbia University, New York, NY), 1:1,000; rabbit anti-neurofascin polyclonal antisera (Tait et al., 2000), 1:1000; affinity-purified rabbit anti-contactin polyclonal antibody (gift of Dr. John Hemperly, Becton Dickinson Research Center, Research Triangle Park, NC) (Rios et al., 2000), 1:100; SMI31, mouse anti-neurofilament monoclonal antibody (Sternberger); 262, rabbit anti-Zic2 antisera, 1:10,000; and mouse anti-calbindin antibody (Sigma-Aldrich), 1:300. Secondary antibodies were from The Jackson Laboratory and used at 1:100–1:300. Adult mice were perfused transcardially with PBS followed by 4% paraformaldehyde and brains were dissected out and kept in 4% paraformaldehyde at 4°C overnight. Brains were embedded in 3% agarose and 100–150 mm vibratome sections were prepared. Immunostaining was performed with 2H3, mouse antineurofilament monoclonal antibody (culture supernatant from Developmental Study Hybridoma Bank) followed by HRP conjugated anti–mouse antibody with DAB as a substrate. Some frozen sections and vibratome sections were stained with cresyl violet, washed with water, and mounted on the Superfrost slide (Fisher Scientific) with gel mount (Biomeda) or Permount. Slides were observed under a Zeiss Axiophot microscope. Photographs were taken using either Kodak Ektachrome 400 (for fluorescence) or Kodak 64T (for bright field). In some cases, images were captured by CCD camera with AG-5 image grabber or Spot camera on a Nikon Diaphot microscope using NIH image software or Adobe PhotoShop software.