

MEMBRANES OF THE HUMAN PEPSINOGEN GRANULE

CLINTON B. LILLIBRIDGE. From the Departments of Anatomy and Medicine, University of Washington, Seattle

Robertson's comprehensive study of the ultrastructure of cell membranes and membranous cell organelles (Robertson, 1957) demonstrated that many cytoplasmic membranes have basically a similar appearance. The appearance of these membranes when cut in cross-section and seen in the electron microscope is two dark lines about 20 Å thick, separated by a less dense space of about 35 Å. Conceiving of these membranes as universal in cells, Robertson called them "unit membranes." Robertson reported unit membranes in the plasma membranes of many types of cells, in the membranes of the nuclear envelope, in the vesicles of the smooth-surfaced endoplasmic reticulum, and in mitochondria. Numerous other investigators have also reported density patterns similar to Robertson's unit membrane.

Recently, however, Suganuma (1961) has shown clearly with the electron microscope that the plasma membrane of *Staphylococcus aureus* displays in sections a density pattern characterized by a single layer about 50 Å thick. Karrer (1960) has reported that the membranes of the cisternae of the rough-surfaced endoplasmic reticulum in the lung phagocytes are composed of a narrow, single-layered membrane unlike the unit membrane (Karrer, 1960). The present study reports another example of a variant from the unit membrane appearance. This variant is seen by electron microscopy in the envelope of certain pepsinogen granules from the "chief cells" in the human stomach.

Although the fine structure of gastric mucosa has been studied by several other investigators (Challice *et al.*, 1957; Dalton, 1951; Hally, 1959; Helander and Ekholm, 1959; Kurosumi *et al.*, 1958; Lawn, 1960; Sedar, 1955, 1959; and Vial and Orrego, 1960), the envelope of the pepsinogen granule has been clearly visualized only by Helander and Ekholm.

Tissue for the present study was obtained by peroral passage of a biopsy instrument (Brandborg *et al.*, 1955) into the stomach of human subjects without anesthesia. Biopsies of the full thickness of the mucosa were obtained. They were fixed in collidine-buffered OsO₄ (Bennett and

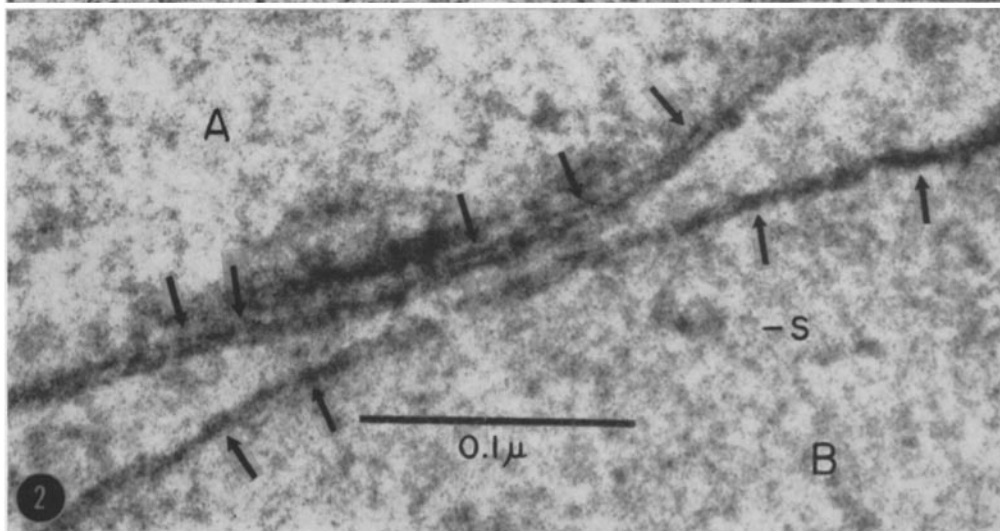
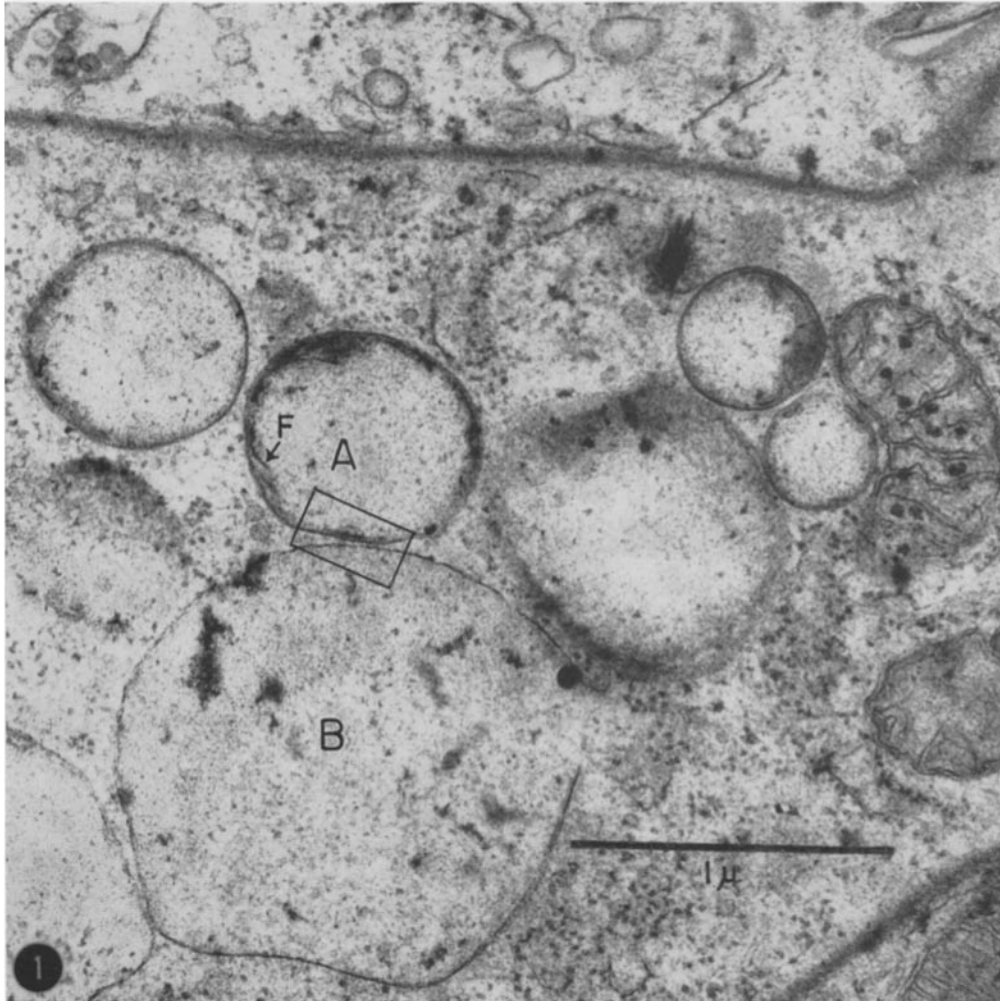
Luft, 1959) and embedded in Epon resin (Luft, 1961). The ultrathin sections were stained with lead hydroxide (Watson, 1958) and examined with a modified RCA EMU-2A electron microscope. Cells were identified by criteria developed in a previous study (Lillibridge, 1960). Distribution of membrane densities were evaluated with a modified Kipp and Zonen densitometer (Bennett *et al.*, 1953).

The secretion granules in human pepsiniferous (chief) cells appear in electron micrographs of thin sections as pale, homogeneous, oval structures enclosed by a membrane. These granules range in diameter from ½ to 3 microns. The smaller granules which are less than about 1 micron across usually are enveloped in a typical unit membrane. The larger granules have a thin, single-layered membrane which occasionally appears not to enclose the granules completely (Fig. 1).

Since measurements of the width of membranes with calipers give data which depend in considerable degree on photographic factors such as grain and contrast, the membranes in this study were measured by a densitometer. Fig. 2 illustrates two adjacent pepsinogen granules, one large, one small, each enclosed by differing membranes. Densitometric tracings from these membranes were taken at the sites indicated by the arrows. The tracings taken across each membrane were superimposed and are shown in Figs. 3 and 4.

The tracings across the membrane of the smaller granule (labeled *A* in Figs. 1 and 2), show two peaks of density with an average peak-to-peak distance on the tracings of about 50 Å (Fig. 3). The over-all width is about 80 Å. This is characteristic of tracings of typical unit membranes. In addition, tracings from membranes of the smaller granules show subsidiary peaks of density superimposed on the two main peaks and in the intervening trough.

In contrast, tracings across the membrane of the larger granule (labeled *B* in Figs. 1 and 2) display a single peak having an over-all width of about 50 Å (Fig. 4). The single-layered appearance of the membranes enclosing the larger gran-



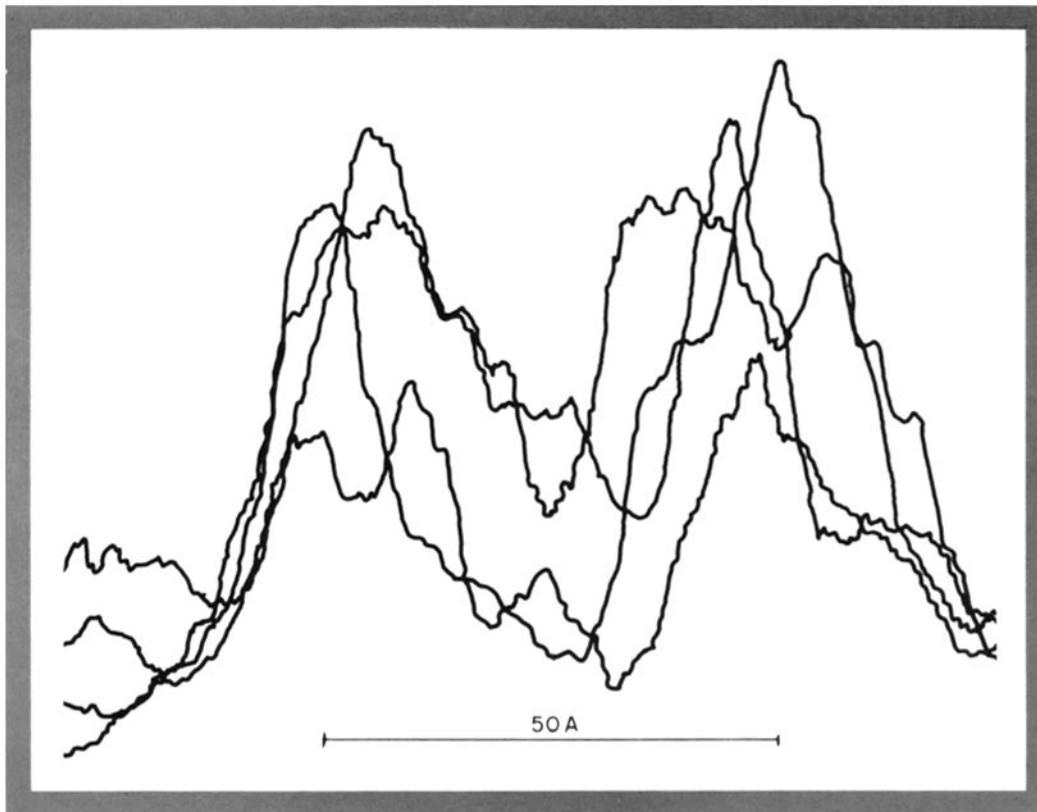


FIGURE 3

Superimposed densitometric tracings taken across the membrane enclosing granule *A* at the sites indicated by arrows in Fig. 2.

ules similar to granule *B* cannot be explained by poor resolution because the granule immediately adjacent does display the typical unit membrane appearance. Nor would oblique alignment of a

typical unit membrane in a 500-Å-thick section account for the appearance of a single dense line of the same width as the usual center-to-center spacing (50 Å) of the perpendicularly oriented

FIGURE 1

An electron micrograph of the apex of a human pepsiniferous (chief) cell showing secretion granules of various sizes enclosed by different types of membranes. The smaller pepsinogen granules (*e.g.* *A*) are enclosed by typical unit membranes. Small fragments, *F*, of membrane may be found lying unattached within the smaller granule near the enclosing membrane. The larger granules (*e.g.* *B*) have thin, apparently single-layered membranes, which, in the case of granule *B*, appear not to enclose the granule completely. $\times 42,000$.

FIGURE 2

An enlargement of the area of granules *A* and *B* enclosed within the rectangle in Fig. 1. Densitometric tracings (Figs. 3 and 4) were made across each membrane at the sites indicated by the arrows. The size of the slit (1.2×0.5 mm. actual size) relative to the micrograph is indicated by line *S*. $\times 360,000$.

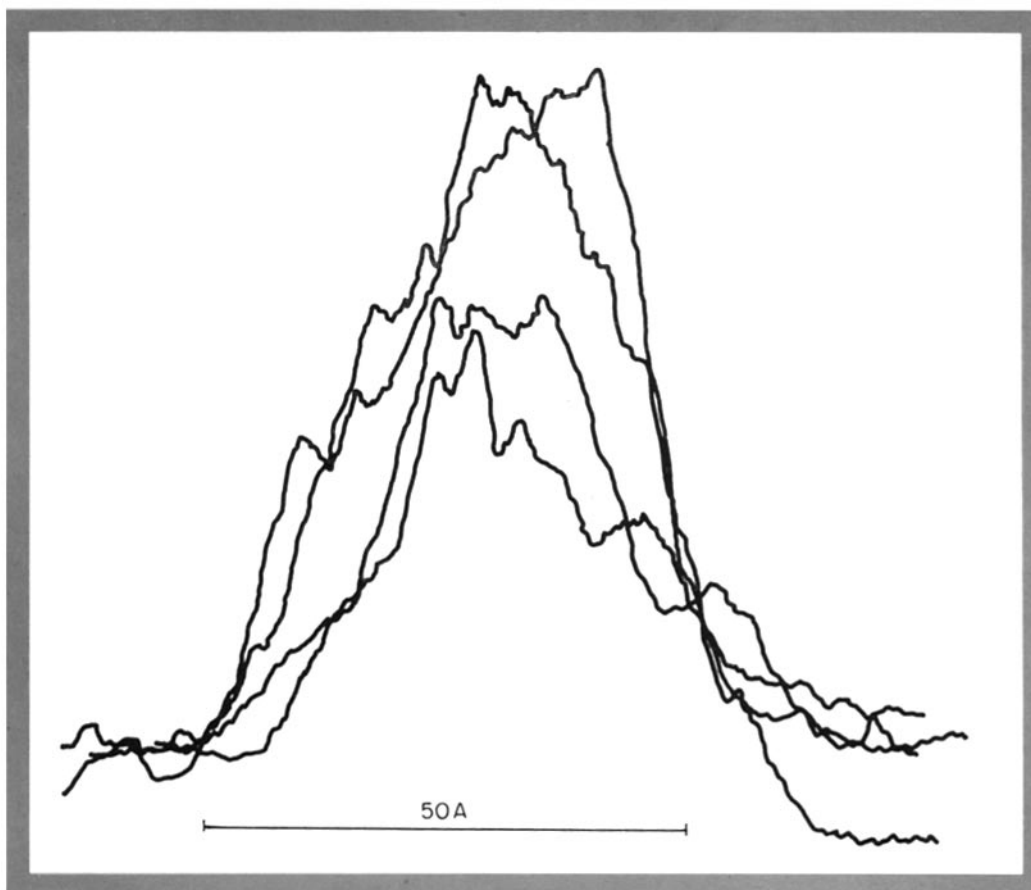


FIGURE 4

Superimposed densitometric tracings taken across the membrane enclosing granule *B* at the sites indicated by arrows in Fig. 2.

unit membrane. Since each picture was selected for focus from a series of five through-focus pictures, spurious images from diffraction effects are minimal.

In conclusion it appears that in the human pepsiniferous (chief) cell of the gastric mucosa the pepsinogen granules of less than 1 micron in diameter are enveloped in typical unit membranes, but the granules of more than 1 micron across have a different type of membrane which is thinner and has a single-layered appearance. These findings are supported by densitometry.

Whether this difference in appearance is due to alteration in basic molecular configuration or to some more subtle change reflected only in reactivity with OsO_4 remains to be answered.

Grateful appreciation is expressed to Dr. H. Stanley Bennett for his encouragement and criticism, and to Dr. Atsushi Suganuma, whose observations stimulated this investigation.

This work was supported in part by a post-sophomore Fellowship Grant No. PX-ss3-7, National Institutes of Health, and by Grant H-2698 (C4) from the National Institutes of Health, United States Public Health Service, Department of Health Education and Welfare.

Received for publication, November 21, 1960.

BIBLIOGRAPHY

1. BENNETT, H. S., QUINTON, W. E., and MUELLER, V. C., A convenient modification of the Moll recording microphotometer, *Appl. Spectros.*, 1953, 7, 127.

2. BENNETT, H. S., and LUFT, J. H., s-Collidine as a basis for buffering fixatives, *J. Biophysic. and Biochem. Cytol.*, 1959, **6**, 113.
3. BRANDBORG, L. L., RUBIN, C. E., and QUINTON, W. E., A multipurpose instrument for suction biopsy of esophagus, stomach, small bowel and colon, *Gastroenterology*, 1955, **121**, 365.
4. CHALLICE, C. E., BULLIVANT, S., and SCOTT, D. B., The fine structure of some cytoplasmic inclusions of oxyntic cells, *Exp. Cell Research*, 1957, **13**, 488.
5. DALTON, A. J., Electron micrography of the epithelium cells of the gastro-intestinal tract and pancreas, *Am. J. Anat.*, 1951, **89**, 109.
6. HALLY, A. D., The fine structure of the gastric parietal cell in the mouse, *J. Anat.*, 1959, **93**, 217.
7. HELANDER, H., and EKHOLM, R., Ultrastructure of epithelial cells in the fundus glands of the mouse gastric mucosa, *J. Ultrastruct. Research*, 1959, **3**, 74.
8. KARRER, H. E., Electron microscopic study of the phagocytosis process in the lung, *J. Biophysic. and Biochem. Cytol.*, 1960, **7**, 357.
9. KUROSUMI, F., SHIBASAKI, S., UCHIDA, G., and TANAKA, Y., Electron microscope studies on the gastric mucosa of normal rats, *Arch. Hist. Jap.*, 1958, **15**, 587.
10. LAWN, A. M., Observations on the fine structure of the gastric parietal cell of the rat, *J. Biophysic. and Biochem. Cytol.*, 1960, **7**, 161.
11. LILLIBRIDGE, C. B., Electron microscopic criteria for identifying human oxyntic and pepsin producing cells in biopsy material, *Anat. Rec.*, 1960, **136**, 234.
12. LUFT, J. H., An improved epoxy resin embedding method, *J. Biophysic. and Biochem. Cytol.*, 1961, **9**, 409.
13. ROBERTSON, J. D., The ultrastructure of cell membranes and their derivatives, in *The Structure and Function of Subcellular Components*, (E. M. Crook, editor), Biochemical Society Symposium No. 16, Cambridge, University Press, Cambridge, 1957, 3.
14. SEDAR, A. W., Fine structure of the parietal cells, *Anat. Rec.*, 1955, **121**, 365.
15. SEDAR, A. W., An attempt to correlate the fine structure of the parietal cell with the functional state of the gastric mucosa, *Anat. Rec.*, 1959, **133**, 337.
16. SUGANUMA, A., The plasma membrane of *Staphylococcus aureus*, *J. Biophysic. and Biochem. Cytol.*, in press.
17. VIAL, J. D. and ORREGO, H., Electron microscope observations on the fine structure of parietal cells, *J. Biophysic. and Biochem. Cytol.*, 1960, **7**, 367.
18. WATSON, M. L., Staining of tissue sections for electron microscopy with heavy metals. II. Application of solutions containing lead and barium, *J. Biophysic. and Biochem. Cytol.*, 1958, **4**, 727.